

---

**Cleo**

**Kyle Johnsen, Nathan Cruzado**

**Mar 20, 2024**



# CONTENTS

- 1 Electrode recording 3**
- 2 1P/2P optogenetic stimulation 5**
- 3 2P imaging 7**
- 4 Getting started 9**
- 5 Related resources 11**
  - 5.1 Publications . . . . . 11
- 6 Documentation contents 13**
  - 6.1 Overview . . . . . 13
  - 6.2 Tutorials . . . . . 20
  - 6.3 Reference . . . . . 108
- 7 Indices and tables 159**
- Python Module Index 161**
- Index 163**



Hello there! Cleo has the goal of bridging theory and experiment for mesoscale neuroscience, facilitating electrode recording, optogenetic stimulation, and closed-loop experiments (e.g., real-time input and output processing) with the [Brian 2](#) spiking neural network simulator. We hope users will find these components useful for prototyping experiments, innovating methods, and testing observations about a hypotheses *in silico*, incorporating into spiking neural network models laboratory techniques ranging from passive observation to complex model-based feedback control. Cleo also serves as an extensible, modular base for developing additional recording and stimulation modules for Brian simulations.

This package was developed by [Kyle Johnsen](#) and Nathan Cruzado under the direction of [Chris Rozell](#) at Georgia Institute of Technology. See the preprint [here](#).

Cleo allows for flexible I/O processing in real time, enabling the simulation of closed-loop experiments such as event-triggered or feedback control. The user can also add latency to the stimulation to study the effects of computation delays.



## **ELECTRODE RECORDING**

Cleo provides functions for configuring electrode arrays and placing them in arbitrary locations in the simulation. The user can then specify parameters for probabilistic spike detection or a spike-based LFP approximation developed by [Teleńczuk et al., 2020](#).





## **1P/2P OPTOGENETIC STIMULATION**

By modeling light propagation and opsins, Cleo enables users to flexibly add photostimulation to their model. Both a four-state Markov state model of opsin kinetics is available, as well as a minimal proportional current option for compatibility with simple neuron models. Cleo also accounts for opsin action spectra to model the effects of multi-light/wavelength/opsin crosstalk and heterogeneous expression. Parameters are for multiple opsins, and blue optic fiber (1P) and infrared spot (for 2P) illumination.



## 2P IMAGING

Users can also inject a microscope into their model, selecting neurons on the specified plane of imaging or elsewhere, with signal and noise strength determined by indicator expression levels and position with respect to the focal plane. The calcium indicator model of [Song et al., 2021](#) is implemented, with parameters included for GCaMP6 variants.



## GETTING STARTED

Just use pip to install—the name on PyPI is `cleosim`:

```
pip install cleosim
```

Then head to the [overview section of the documentation](#) for a more detailed discussion of motivation, structure, and basic usage.



## RELATED RESOURCES

Those using Cleo to simulate closed-loop control experiments may be interested in software developed for the execution of real-time, *in-vivo* experiments. Developed by members of [Chris Rozell's](#) and [Garrett Stanley's](#) labs at Georgia Tech, the [CLOCTools repository](#) can serve these users in two ways:

1. By providing utilities and interfaces with experimental platforms for moving from simulation to reality.
2. By providing performant control and estimation algorithms for feedback control. Although Cleo enables closed-loop manipulation of network simulations, it does not include any advanced control algorithms itself. The `ldsCtrlEst` library implements adaptive linear dynamical system-based control while the `hmm` library can generate and decode systems with discrete latent states and observations.

### 5.1 Publications

**Cleo: A testbed for bridging model and experiment by simulating closed-loop stimulation, electrode recording, and optophysiology** K.A. Johnsen, N.A. Cruzado, Z.C. Menard, A.A. Willats, A.S. Charles, and C.J. Rozell. *bioRxiv*, 2023.

**CLOC Tools: A Library of Tools for Closed-Loop Neuroscience** A.A. Willats, M.F. Bolus, K.A. Johnsen, G.B. Stanley, and C.J. Rozell. *In prep*, 2023.

**State-Aware Control of Switching Neural Dynamics** A.A. Willats, M.F. Bolus, C.J. Whitmire, G.B. Stanley, and C.J. Rozell. *In prep*, 2023.

**Closed-Loop Identifiability in Neural Circuits** A. Willats, M. O'Shaughnessy, and C. Rozell. *In prep*, 2023.

**State-space optimal feedback control of optogenetically driven neural activity** M.F. Bolus, A.A. Willats, C.J. Rozell and G.B. Stanley. *Journal of Neural Engineering*, 18(3), pp. 036006, March 2021.

**Design strategies for dynamic closed-loop optogenetic neurocontrol in vivo** M.F. Bolus, A.A. Willats, C.J. Whitmire, C.J. Rozell and G.B. Stanley. *Journal of Neural Engineering*, 15(2), pp. 026011, January 2018.





## DOCUMENTATION CONTENTS

### 6.1 Overview

#### 6.1.1 Introduction

##### Who is this package for?

Cleo (Closed-Loop, Electrophysiology, and Optophysiology experiment simulation testbed) is a Python package developed to bridge theory and experiment for mesoscale neuroscience. We envision two primary uses cases:

1. For prototyping closed-loop control of neural activity *in silico*. Animal experiments are costly to set up and debug, especially with the added complexity of real-time intervention—our aim is to enable researchers, given a decent spiking model of the system of interest, to assess whether the type of control they desire is feasible and/or what configuration(s) would be most conducive to their goals.
2. The complexity of experimental interfaces means it's not always clear what a model would look like in a real experiment. Cleo can help anyone interested in observing or manipulating a model while taking into account the constraints present in real experiments. Because Cleo is built around the [Brian simulator](#), we especially hope this is helpful for existing Brian users who for whatever reason would like a convenient way to inject recorders (e.g., electrodes or 2P imaging) or stimulators (e.g., optogenetics) into the core network simulation.

---

##### What is closed-loop control?

In short, determining the inputs to deliver to a system from its outputs. In neuroscience terms, making the stimulation parameters a function of the data recorded in real time.

---

##### Structure and design

Cleo wraps a spiking network simulator and allows for the injection of stimulators and/or recorders. The models used to emulate these devices are often non-trivial to implement or use in a flexible manner, so Cleo aims to make device injection and configuration as painless as possible, requiring minimal modification to the original network.

Cleo also orchestrates communication between the simulator and a user-configured [IOProcessor](#) object, modeling how experiment hardware takes samples, processes signals, and controls stimulation devices in real time.

For an explanation of why we choose to prioritize spiking network models and how we chose Brian as the underlying simulator, see [Design rationale](#).

---

##### Why closed-loop control in neuroscience?

---

Fast, real-time, closed-loop control of neural activity enables intervention in processes that are too fast or unpredictable to control manually or with pre-defined stimulation, such as sensory information processing, motor planning, and oscillatory activity. Closed-loop control in a *reactive* sense enables the experimenter to respond to discrete events of interest, such as the arrival of a traveling wave or sharp wave ripple, whereas *feedback* control deals with driving the system towards a desired point or along a desired state trajectory. The latter has the effect of rejecting noise and disturbances, reducing variability across time and across trials, allowing the researcher to perform inference with less data and on a finer scale. Additionally, closed-loop control can compensate for model mismatch, allowing it to reach more complex targets where open-loop control based on imperfect models is bound to fail.

## 6.1.2 Installation

Make sure you have Python 3.7, then use pip: `pip install cleosim`.

**Caution:** The name on PyPI is `cleosim` since `cleo` was already taken, but in code it is still used as `import cleo`. The other Cleo appears to actually be a fairly well developed package, so I'm sorry if you need to use it along with this Cleo in the same environment. In that case, [there are workarounds](#).

Or, if you're a developer, install `poetry` and run `poetry install` from the repository root.

## 6.1.3 Usage

### Brian network model

The starting point for using Cleo is a Brian spiking neural network model of the system of interest. For those new to Brian, the [docs](#) are a great resource. If you have a model built with another simulator or modeling language, you may be able to [import it to Brian via NeuroML](#).

Perhaps the biggest change you may have to make to an existing model to make it compatible with Cleo's optogenetics and electrode recording is to give the neurons of interest coordinates in space. See the tutorials or the `cleo.coords` module for more info.

You'll need your model in a Brian `Network` object before you move on. E.g.,:

```
import brian2 as b2
ng = b2.NeuronGroup( # a simple population of 100 LIF neurons
    500,
    """dv/dt = (-v - 70*mV + (500*Mohm)*Iopto + 2*xi*sqrt(tau_m)*mvolt) / tau_m : volt
    Iopto : amp""",
    threshold='v>-50*mV',
    reset='v=-70*mV',
    namespace={'tau_m': 20*b2.ms},
)
ng.v = -70*b2.mV
net = b2.Network(ng)
```

Your neurons need x, y, and z coordinates for Cleo's spatially defined recording and stimulation models to work:

```
import cleo
cleo.coords.assign_coords_rand_rect_prism(
    ng, xlim=(-0.2, 0.2), ylim=(-0.2, 0.2), zlim=(0.2, 1), unit=b2.mm
)
```

## CLSimulator

Once you have a network model, you can construct a *CLSimulator* object:

```
sim = cleo.CLSimulator(net)
```

The simulator object wraps the Brian network and coordinates device injection, processing input and output, and running the simulation.

## Recording

Recording devices take measurements of the Brian network. To use a *Recorder*, you must inject it into the simulator via *cleo.CLSimulator.inject()*. The recorder will only record from the neuron groups specified on injection, allowing for such scenarios as singling out a cell type to record from. Some extremely simple implementations (which do little more than wrap Brian monitors) are available in the *cleo.recorders* module. See the *Electrode recording* and *All-optical control* tutorials for more detail on how to record from a simulation more realistically, but here's a quick example of how to record multi-unit spiking activity with an electrode:

```
# configure and inject a 32-channel shank
coords = cleo.ephys.linear_shank_coords(
    array_length=1*b2.mm, channel_count=32, start_location=(0, 0, 0.2)*b2.mm
)
mua = cleo.ephys.MultiUnitSpiking(
    r_perfect_detection=20 * b2.um,
    r_half_detection=40 * b2.um,
)
probe = cleo.ephys.Probe(coords, signals=[mua])
sim.inject(probe, ng)
```

```
-----
DimensionMismatchError                                Traceback (most recent call last)
Cell In[4], line 10
      5 mua = cleo.ephys.MultiUnitSpiking(
      6     r_perfect_detection=20 * b2.um,
      7     r_half_detection=40 * b2.um,
      8 )
      9 probe = cleo.ephys.Probe(coords, signals=[mua])
--> 10 sim.inject(probe, ng)

File ~/checkouts/readthedocs.org/user_builds/cleosim/envs/v0.14.0/lib/python3.12/site-
packages/cleo/base.py:383, in CLSimulator.inject(self, device, *neuron_groups,
**kwargs)
    381     device.sim = self
    382     device.init_for_simulator(self)
--> 383     device.connect_to_neuron_group(ng, **kwargs)
    384     for brian_object in device.brian_objects:
    385         if brian_object not in self.network.objects:

File ~/checkouts/readthedocs.org/user_builds/cleosim/envs/v0.14.0/lib/python3.12/site-
packages/cleo/ephys/probes.py:158, in Probe.connect_to_neuron_group(self, neuron_group,
**kwargs)
    146 """Configure probe to record from given neuron group
    147
```

(continues on next page)

(continued from previous page)

```

148 Will call :meth:`Signal.connect_to_neuron_group` for each signal
(...)
155     Passed in to signals' connect functions, needed for some signals
156 """
157 for signal in self.signals:
--> 158     signal.connect_to_neuron_group(neuron_group, **kwparams)
159     self.brian_objects.update(signal.brian_objects)

File ~/checkouts/readthedocs.org/user_builds/cleosim/envs/v0.14.0/lib/python3.12/site-
packages/cleo/ephys/spiking.py:208, in MultiUnitSpiking.connect_to_neuron_group(self,
neuron_group, **kwparams)
200 def connect_to_neuron_group(self, neuron_group: NeuronGroup, **kwparams) -> None:
201     """Configure signal to record from specified neuron group
202
203     Parameters
204     (...)
206         group to record from
207     """
--> 208     neuron_channel_dtct_probs = super(
209         MultiUnitSpiking, self
210     ).connect_to_neuron_group(neuron_group, **kwparams)
211     if self._dtct_prob_array is None:
212         self._dtct_prob_array = neuron_channel_dtct_probs

File ~/checkouts/readthedocs.org/user_builds/cleosim/envs/v0.14.0/lib/python3.12/site-
packages/cleo/ephys/spiking.py:99, in Spiking.connect_to_neuron_group(self, neuron_
group, **kwparams)
97 distances = np.sqrt(dist2) * meter # since units stripped
98 # probs is n_neurons by n_channels
--> 99 probs = self._detection_prob_for_distance(distances)
100 # cut off to get indices of neurons to monitor
101 # [0] since nonzero returns tuple of array per axis
102 i_ng_to_keep = np.nonzero(np.all(probs > self.cutoff_probability, axis=1))[0]

File ~/checkouts/readthedocs.org/user_builds/cleosim/envs/v0.14.0/lib/python3.12/site-
packages/cleo/ephys/spiking.py:148, in Spiking._detection_prob_for_distance(self, r)
146 h = b - a
147 with np.errstate(divide="ignore"):
--> 148     decaying_p = h / (r - c)
149 decaying_p[decaying_p == np.inf] = 1 # to fix NaNs caused by /0
150 p = 1 * (r <= a) + decaying_p * (r > a)

File ~/checkouts/readthedocs.org/user_builds/cleosim/envs/v0.14.0/lib/python3.12/site-
packages/brian2/units/fundamentalunits.py:1193, in Quantity.__array_ufunc__(self, uf,
method, *inputs, **kwargs)
1190 elif uf.__name__ in UFUNCS_MATCHING_DIMENSIONS + UFUNCS_COMPARISONS:
1191     # Ok if dimension of arguments match (for reductions, they always do)
1192     if method == "__call__":
-> 1193         fail_for_dimension_mismatch(
1194             inputs[0],
1195             inputs[1],
1196             error_message=(

```

(continues on next page)

(continued from previous page)

```

1197         "Cannot calculate {val1} %s {val2}, the units do not match"
1198     )
1199     % uf.__name__,
1200     val1=inputs[0],
1201     val2=inputs[1],
1202 )
1203 if uf.__name__ in UFUNCS_COMPARISONS:
1204     return uf_method(*[np.asarray(i) for i in inputs], **kwargs)

```

File ~/checkouts/readthedocs.org/user\_builds/cleosim/envs/v0.14.0/lib/python3.12/site-packages/brian2/units/fundamentalunits.py:266, in fail\_for\_dimension\_mismatch(obj1, obj2, error\_message, \*\*error\_quantities)

```

264     raise DimensionMismatchError(error_message, dim1)
265 else:
--> 266     raise DimensionMismatchError(error_message, dim1, dim2)
267 else:
268     return dim1, dim2

```

DimensionMismatchError: Cannot calculate [[439.36017494 411.88178876 ... 629.77906691 660.04990818]  
[ 79.65293991 64.32448351 ... 919.47975127 951.66749704]  
...  
[681.92926009 651.00913615 ... 365.9178535 393.84796392]  
[222.8764748 204.71210451 ... 849.69087103 881.2733189 ]] mm^2 subtract 0. m, the units do not match (units are m^2 and m).

## Stimulation

*Stimulator* devices manipulate the Brian network, and are likewise *inject()*ed into the simulator, into specified neuron groups. Optogenetics (1P and 2P) is the main stimulation modality currently implemented by Cleo. This requires injection of both a light source and an opsin—see the *Optogenetic stimulation* and *All-optical control* tutorials for more detail.

```

fiber = cleo.light.Light(
    coords=(0, 0, 0.5)*b2.mm,
    light_model=cleo.light.fiber473nm(),
    wavelength=473*b2.nmeter
)
chr2 = cleo.opto.chr2_4s()
sim.inject(fiber, ng).inject(chr2, ng)

```

**Note:** Markov opsin kinetics models require target neurons to have membrane potentials in realistic ranges and an *Iopto* term defined in amperes. If you need to interface with a model without these features, you may want to use the simplified *ProportionalCurrentOpsin*. You can find more details, including a comparison between the two model types, in the *optogenetics tutorial*.

**Note:** Recorders and stimulators need unique names which serve as keys to access/set values in the *IOProcessor*'s input/output dictionaries. The name defaults to the class name, but you can specify it on construction.

## I/O Processor

Just as in a real experiment where the experiment hardware must be connected to signal processing equipment and/or computers for recording and control, the *CLSimulator* must be connected to an *IOProcessor*. If you are only recording, you may want to use the *RecordOnlyProcessor*. Otherwise you will want to implement the *LatencyIOProcessor*, which not only takes samples at the specified rate, but processes the data and delivers input to the network after a user-defined delay, emulating the latency inherent in real experiments. This is done by creating a subclass and defining the *process()* function:

```
class MyProcessor(cleo.ioproc.LatencyIOProcessor):
    def process(self, state_dict, sample_time_ms):
        # state_dict contains a {'recorder_name': value} dict of network.
        i_spikes, t_ms_spikes, y_spikes = state_dict['Probe']['MultiUnitSpiking']
        # on-off control
        irr0_mW_per_mm2 = 5 if len(i_spikes) < 10 else 0
        # output is a {'stimulator_name': value} dict and output time
        return {'Light': irr0_mW_per_mm2}, sample_time_ms + 3 # (3 ms delay)

sim.set_io_processor(MyProcessor(sample_period_ms=1))
```

The *On-off control*, *PI control*, and *LQR optimal control using ldscstrlest* tutorials give examples of closed-loop control ranging from simple to complex.

## Visualization

*cleo.viz.plot()* allows you to easily visualize your experimental configuration:

```
cleo.viz.plot(ng, colors=['#c500cc'], sim=sim, zlim=(200, 1000))
```

Cleo also features some *video visualization capabilities*.

## Running experiments

Use *cleo.CLSimulator.run()* function with the desired duration. This wrap's Brian's *brian2.core.network.Network.run()* function:

```
sim.run(50 * b2.ms) # kwargs are passed to Brian's run function
```

Use *reset()* to restore the default state (right after initialization/injection) for the network and all devices. This could be useful for running a simulation multiple times under different conditions.

To facilitate access to data after the simulation, devices offer a *cleo.InterfaceDevice.save\_history* option on construction, by default True. If true, that object will store relevant variables as attributes. For example:

```
import matplotlib.pyplot as plt
fig, (ax1, ax2) = plt.subplots(2, 1, sharex=True)
ax1.scatter(mua.t_ms, mua.i, marker='.', c='white', s=2)
ax1.set(ylabel='channel index', title='spikes')
ax2.step(fiber.t_ms, fiber.values, c='#72b5f2')
ax2.set(xlabel='time (ms)', ylabel='irradiance (mW/mm²)', title='photostimulation')
```

## 6.1.4 Design rationale

### Why not prototype with more abstract models?

Cleo aims to be practical, and as such provides models at the level of abstraction corresponding to the variables the experimenter has available to manipulate. This means models of spatially defined, spiking neural networks.

Of course, neuroscience is studied at many spatial and temporal scales. While other projects may be better suited for larger segments of the brain and/or longer timescales (such as [HNN](#) or BMTK's [PopNet](#) or [FilterNet](#)), this project caters to finer-grained models because they can directly simulate the effects of alternate experimental configurations. For example, how would the model change when swapping one opsin for another, using multiple opsins simultaneously, or with heterogeneous expression? How does recording or stimulating one cell type vs. another affect the experiment? Would using a more sophisticated control algorithm be worth the extra compute time, and thus later stimulus delivery, compared to a simpler controller?

Questions like these could be answered using an abstract dynamical system model of a neural circuit, but they would require the extra step of mapping the afore-mentioned details to a suitable abstraction—e.g., estimating a transfer function to model optogenetic stimulation for a given opsin and light configuration. Thus, we haven't emphasized these sorts of models so far in our development of Cleo, though they should be possible to implement in Brian if you are interested. For example, one could develop a Poisson linear dynamical system (PLDS), record spiking output, and configure stimulation to act directly on the system's latent state.

And just as experiment prototyping could be done on a more abstract level, it could also be done on an even more realistic level, which we did not deem necessary. That brings us to the next point...

### Why Brian?

Brian is a relatively new spiking neural network simulator written in Python. Here are some of its advantages:

- Flexibility: allowing (and requiring!) the user to define models mathematically rather than selecting from a pre-defined library of cell types and features. This enables us to define arbitrary models for recorders and stimulators and easily interface with the simulation
- Ease of use: it's all just Python
- Speed (at least when compiled to C++ or GPU—see [Brian2GENN](#), [Brian2CUDA](#))

[NEST](#) is a popular alternative to Brian also strong in point neuron simulations. However, it appears to be less flexible, and thus harder to extend. [NEURON](#) is another popular alternative to Brian. Its main advantage is its first-class support of detailed, morphological, multi-compartment neurons. In fact, strong alternatives to Brian for this project were BioNet ([docs](#), [paper](#)) and NetPyNE ([docs](#), [paper](#)), which already offer a high-level interface to NEURON with extracellular potential recording. Optogenetics could be incorporated with [pre-existing .hoc code](#), though the light model would need to be implemented. From brief examination of the [source code of BioNet](#), it appears that closed-loop stimulation would not be too difficult to add. It is unclear for NetPyNE.

In the end, we chose Brian since our priority was to model circuit/population-level dynamics over molecular/intra-neuron dynamics. Also, Brian does have support for multi-compartment neurons, albeit less fully featured, if that is needed.

[PyNN](#) would have been an ideal interface to build around, as it supports multiple simulator backends. The difficulty is, implementing objects not natively supported by SNN simulators (e.g., opsins, calcium indicators, and light source) has required bespoke, idiosyncratic code applicable only to one simulator. To do so in a native, efficient way, as we have attempted to do with Brian, would require significant work. A collaborative effort extending a multi-simulator framework such as PyNN for this purpose may be of value if there is enough community interest in expanding the open-source SNN experiment simulation toolbox.

## 6.1.5 Future development

Here are some features which are missing but could be useful to add:

- Electrode microstimulation
- A more accurate LFP signal (only usable for morphological neurons) based on the volume conductor forward model as in [LFPy](#) or [Vertex](#)
- The [Mazzoni-Lindén LFP approximation](#) for LIF point-neuron networks
- Voltage indicators
- An expanded calcium indicator library—currently the parameter set for the full [NAOMi](#) model is only available for GCaMP6f. The [phenomenological S2F model](#) should be easy to implement and fit to data, has parameters for several of the latest and greatest GECIs (jGCaMP7 and jGCaMP8 varieties), and should be cheaper to simulate as well.

## 6.2 Tutorials

### 6.2.1 Electrode recording

How to insert electrodes to measure different spiking and extracellular signals from a Brian network simulation.

Preamble:

```
import brian2.only as b2
from brian2 import np
import matplotlib.pyplot as plt
import cleo

# the default cython compilation target isn't worth it for
# this trivial example
b2.prefs.codegen.target = "numpy"
b2.seed(1919)
np.random.seed(1919)

cleo.utilities.style_plots_for_docs()

# colors
c = {
    "light": "#df87e1",
    "main": "#C500CC",
    "dark": "#8000B4",
    "exc": "#d6755e",
    "inh": "#056eee",
    "accent": "#36827F",
}
```



## Network setup

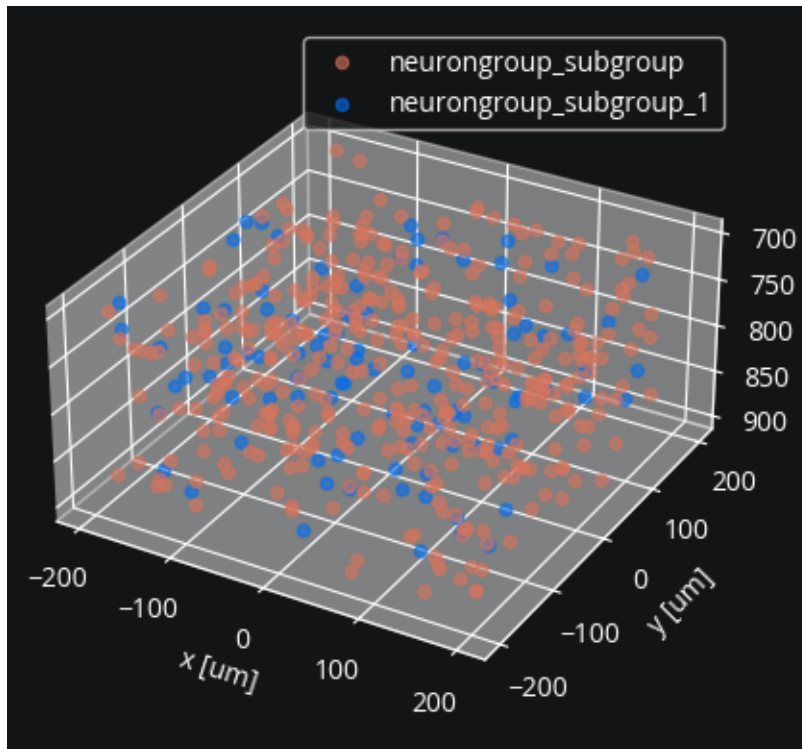
First we create a simple E-I network with external Poisson input and assign coordinates. We only need spiking neurons to record *TKLFPSignal*, but we need synapses onto pyramidal cells for *RWSLFPSignalFromSpikes*.

```
N = 500
n_e = int(N * 0.8)
n_i = int(N * 0.2)
n_ext = N

neurons = b2.NeuronGroup(
    N,
    "dv/dt = -v / (10*ms) : 1",
    threshold="v > 1",
    reset="v = 0",
    refractory=2 * b2.ms,
)
ext_input = b2.PoissonGroup(n_ext, 23 * b2.Hz, name="ext_input")
cleo.coords.assign_coords_rand_rect_prism(
    neurons, xlim=(-0.2, 0.2), ylim=(-0.2, 0.2), zlim=(0.7, 0.9)
)
# need to create subgroups after assigning coordinates
exc = neurons[:n_e]
inh = neurons[n_e:]

w0 = 0.06
syn_exc = b2.Synapses(
    exc,
    neurons,
    f"w = {w0} : 1",
    on_pre="v_post += w",
    name="syn_exc",
    delay=1.5 * b2.ms,
)
syn_exc.connect(p=0.1)
syn_inh = b2.Synapses(
    inh,
    neurons,
    f"w = -4*{w0} : 1",
    on_pre="v_post += w",
    name="syn_inh",
    delay=1.5 * b2.ms,
)
syn_inh.connect(p=0.1)
syn_ext = b2.Synapses(
    ext_input, neurons, "w = .05 : 1", on_pre="v_post += w", name="syn_ext"
)
syn_ext.connect(p=0.1)

net = b2.Network([neurons, exc, inh, syn_exc, syn_inh, ext_input, syn_ext])
sim = cleo.CLSimulator(net)
cleo.viz.plot(exc, inh, colors=[c["exc"], c["inh"]], scatterargs={"alpha": 0.6});
```



### Specifying electrode coordinates

Now we insert an electrode shank probe in the center of the population by injecting a *Probe* device. Note that *Probe* takes arbitrary coordinates as arguments, so you can place contacts wherever you wish. However, the *cleo.ephys* module provides convenience functions to easily generate coordinates common in NeuroNexus probes. Here are some examples:

```
from cleo import ephys
from mpl_toolkits.mplot3d import Axes3D

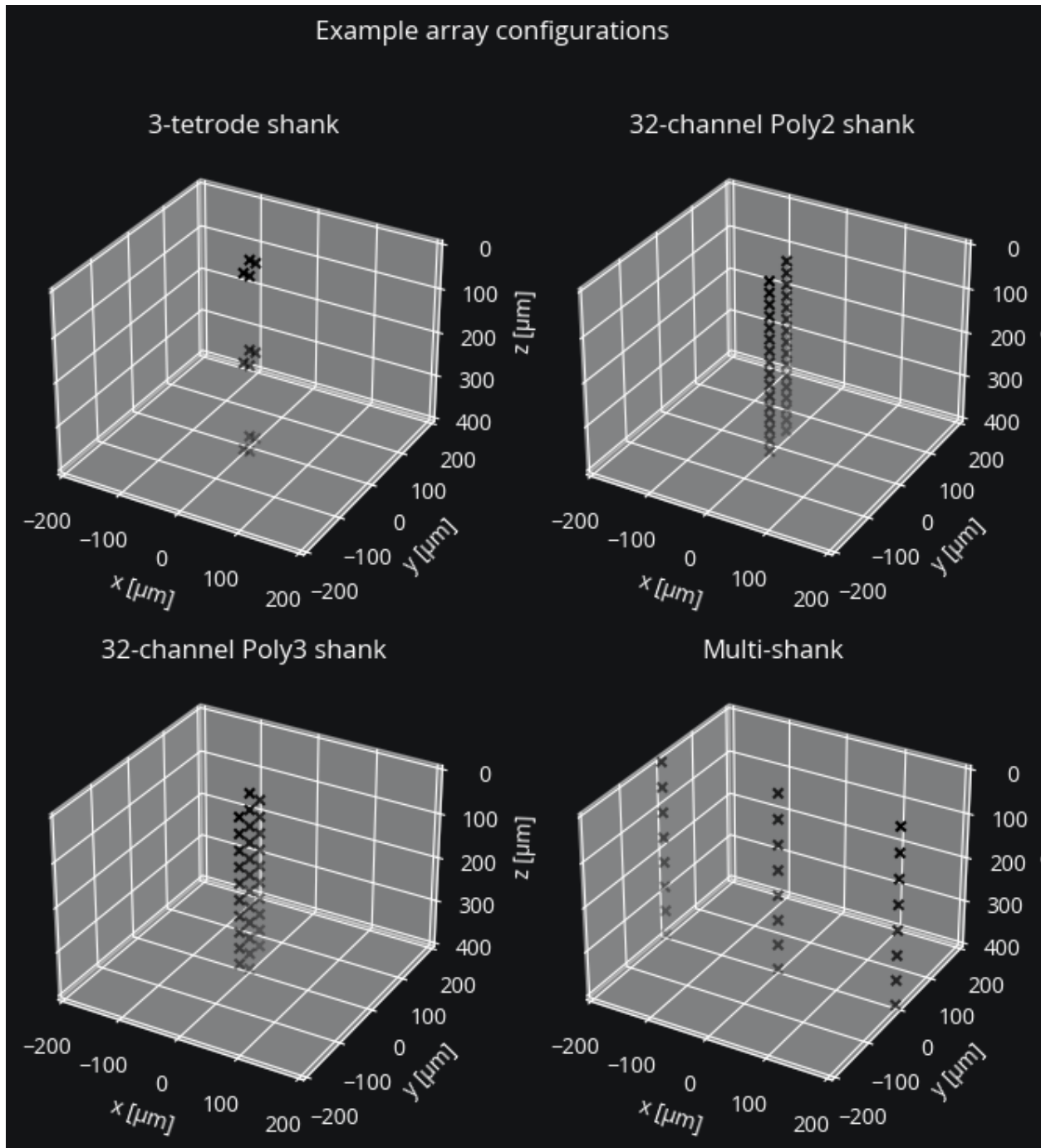
array_length = 0.4 * b2.mm # length of the array itself, not the shank
tetr_coords = ephys.tetrode_shank_coords(array_length, tetrode_count=3)
poly2_coords = ephys.poly2_shank_coords(
    array_length, channel_count=32, intercol_space=50 * b2.um
)
poly3_coords = ephys.poly3_shank_coords(
    array_length, channel_count=32, intercol_space=30 * b2.um
)
# by default start_location (location of first contact) is at (0, 0, 0)
single_shank = ephys.linear_shank_coords(
    array_length, channel_count=8, start_location=(-0.2, 0, 0) * b2.mm
)
# tile vector determines length and direction of tiling (repeating)
multishank = ephys.tile_coords(
    single_shank, num_tiles=3, tile_vector=(0.4, 0, 0) * b2.mm
)

fig = plt.figure(figsize=(8, 8))
```

(continues on next page)

(continued from previous page)

```
fig.suptitle("Example array configurations")
for i, (coords, title) in enumerate(
    [
        (tetr_coords, "3-tetrode shank"),
        (poly2_coords, "32-channel Poly2 shank"),
        (poly3_coords, "32-channel Poly3 shank"),
        (multishank, "Multi-shank"),
    ],
    start=1,
):
    ax = fig.add_subplot(2, 2, i, projection="3d")
    x, y, z = coords.T / b2.um
    ax.scatter(x, y, z, marker="x", c="black")
    ax.set(
        title=title,
        xlabel="x [μm]",
        ylabel="y [μm]",
        zlabel="z [μm]",
        xlim=(-200, 200),
        ylim=(-200, 200),
        zlim=(400, 0),
    )
)
```



As seen above, the `tile_coords()` function can be used to repeat a single shank to produce coordinates for a multi-shank probe. Likewise it can be used to repeat multi-shank coordinates to achieve a 3D recording array (what NeuroNexus calls a `MatrixArray`).

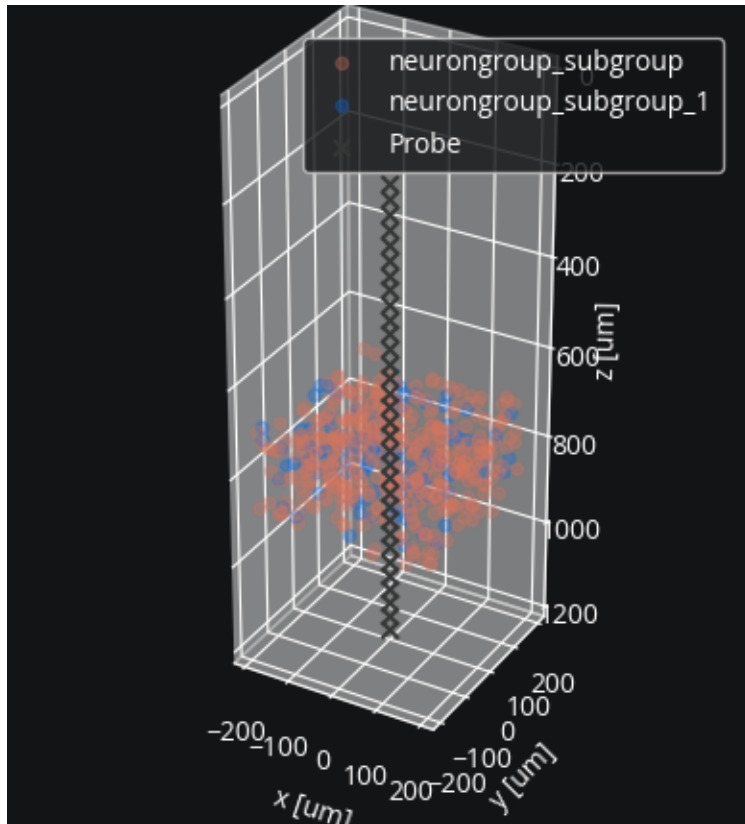
For our example we will use a simple linear array. We configure the probe so it has 32 contacts ranging from 0.2 to 1.2 mm in depth. We could specify the orientation, but by default shank coordinates extend downwards (in the positive  $z$  direction).

We can add the electrode to the plotting function to visualize it along with the neurons:

```

coords = ephys.linear_shank_coords(1 * b2.mm, 32, start_location=(0, 0, 0.2) * b2.mm)
probe = ephys.Probe(coords, save_history=True)
cleo.viz.plot(
    exc,
    inh,
    colors=[c["exc"], c["inh"]],
    zlim=(0, 1200),
    devices=[probe],
    scatterargs={"alpha": 0.3},
);

```



## Specifying signals to record

This looks right, but we need to specify what signals we want to pick up with our electrode. Let's try the two basic spiking signals and an LFP approximation for point neurons.

The two spiking signals (sorted and multi-unit) take the same parameters, mainly `r_perfect_detection`, within which all spikes will be detected, and `r_half_detection`, at which distance a spike has only a 50% chance of being detected. My choice to set these parameters at 50 and 100  $\mu\text{m}$  is arbitrary, though from [at least some published data](#) that seems reasonable.

We use default parameters for the [Teleńczuk kernel LFP approximation method \(TKLFP\)](#), but will need to specify cell type (excitatory or inhibitory) and sampling period (if unavailable from a connected IO processor) upon injection.

The [reference weighted sum of synaptic currents LFP proxy \(RWSLFP\)](#) requires synaptic currents, which we synthesize by convolving spikes with a biexponential kernel. We need to pass in `ampa_syns` and `gaba_syns` on injection to compute the effect of incoming spikes on the population we're recording from. We only need to do this for pyramidal

cells, since RWSLFP ignores currents onto interneurons.

```
mua = ephys.MultiUnitSpiking(
    r_perfect_detection=0.05 * b2.mm,
    r_half_detection=0.1 * b2.mm,
)
ss = ephys.SortedSpiking(0.05 * b2.mm, 0.1 * b2.mm)
tklfp = ephys.TKLFPSignal()
rwslfp = ephys.RWSLFPSignalFromSpikes()

probe.add_signals(mua, ss, tklfp, rwslfp)

sim.set_io_processor(cleo.ioproc.RecordOnlyProcessor(sample_period_ms=1))
sim.inject(
    probe,
    exc,
    tklfp_type="exc",
    ampa_syns=[syn_exc[f"j < {n_e}"], syn_ext[f"j < {n_e}"]],
    gaba_syns=[syn_inh[f"j < {n_e}"]],
)
sim.inject(probe, inh, tklfp_type="inh")
```

```
CLSimulator(io_processor=RecordOnlyProcessor(sample_period_ms=1, sampling='fixed',
    ↪processing='parallel'), devices={Probe(name='Probe', save_history=True,
    ↪signals=[MultiUnitSpiking(name='MultiUnitSpiking', brian_objects={<SpikeMonitor,
    ↪recording from 'spikemonitor_6'>, <SpikeMonitor, recording from 'spikemonitor'>},
    ↪probe=..., r_perfect_detection=50. * umetre, r_half_detection=100. * umetre, cutoff_
    ↪probability=0.01), SortedSpiking(name='SortedSpiking', brian_objects={<SpikeMonitor,
    ↪recording from 'spikemonitor_1'>, <SpikeMonitor, recording from 'spikemonitor_7'>},
    ↪probe=..., r_perfect_detection=50. * umetre, r_half_detection=100. * umetre, cutoff_
    ↪probability=0.01), TKLFPSignal(name='TKLFPSignal', brian_objects={<SpikeMonitor,
    ↪recording from 'spikemonitor_2'>, <SpikeMonitor, recording from 'spikemonitor_8'>},
    ↪probe=..., uLFP_threshold_uV=0.001, _lfp_unit=uvolt), RWSLFPSignalFromSpikes(name=
    ↪'RWSLFPSignalFromSpikes', brian_objects={<SpikeMonitor, recording from 'spikemonitor_5
    ↪'>, <SpikeMonitor, recording from 'spikemonitor_4'>, <SpikeMonitor, recording from
    ↪'spikemonitor_3'>}, probe=..., amp_func=<function mazzoni15_nrn at 0x7faa69039080>,
    ↪pop_aggregate=False, _lfp_unit=1, tau1_ampa=2. * msecond, tau2_ampa=0.4 * msecond,
    ↪tau1_gaba=5. * msecond, tau2_gaba=250. * usecond, syn_delay=1. * msecond, I_
    ↪threshold=0.001, weight='w')], probe=NOTHING)})
```

## Simulation and results

Now we'll run the simulation:

```
sim.run(250 * b2.ms)
```

```
INFO          No numerical integration method specified for group 'neurongroup', using
    ↪method 'exact' (took 0.18s). [brian2.stateupdaters.base.method_choice]
```

And plot the output of the four signals we've recorded:

```

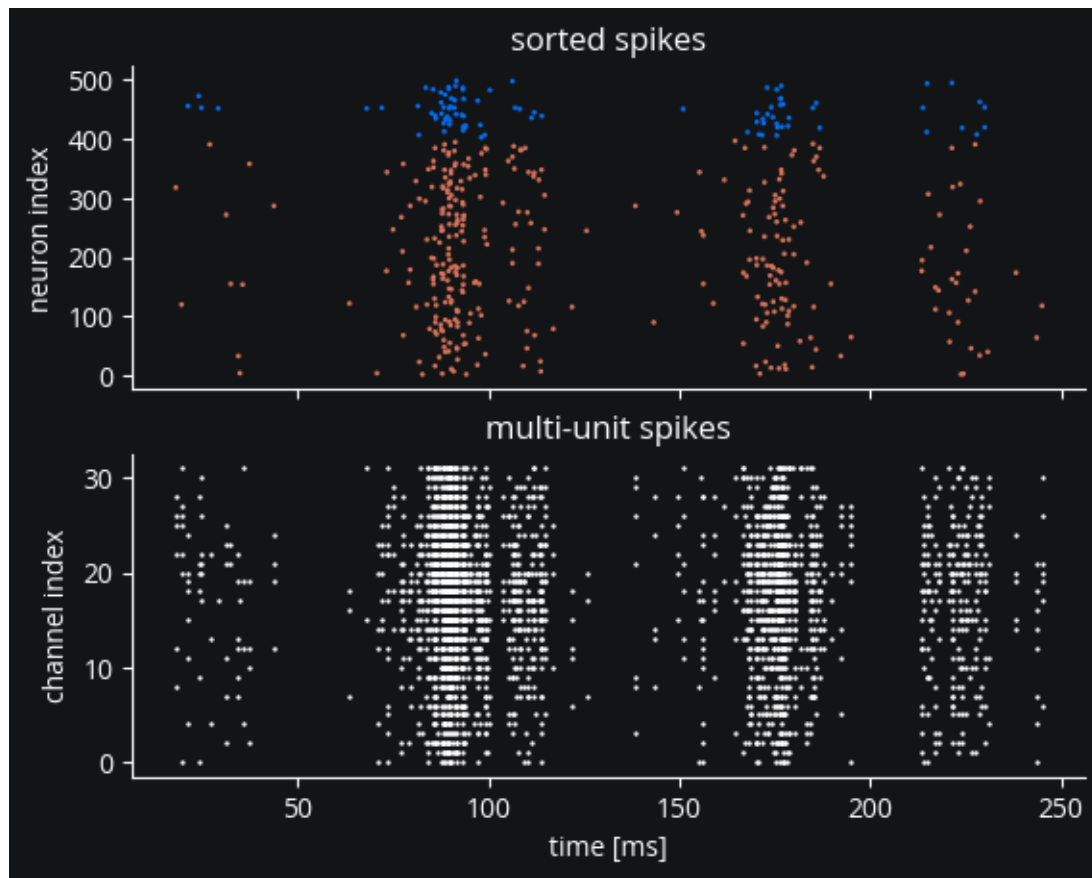
from matplotlib.colors import ListedColormap, LinearSegmentedColormap

fig, axs = plt.subplots(2, 1, sharex=True)

# assuming all neurons are detectable for c=ss.i >= n_e to work
# in practice this will often not be the case and we'd have to map
# from probe index to neuron group index using ss.i_probe_by_i_ng.inverse
exc_inh_cmap = ListedColormap([c["exc"], c["inh"]])
axs[0].scatter(ss.t_ms, ss.i, marker=".", c=ss.i >= n_e, cmap=exc_inh_cmap, s=3)
axs[0].set(title="sorted spikes", ylabel="neuron index")

axs[1].scatter(mua.t_ms, mua.i, marker=".", s=2, c="white")
axs[1].set(title="multi-unit spikes", ylabel="channel index", xlabel="time [ms]");

```



TKLFP supposedly outputs a value with an absolute scale in terms of V, though it is quite high compared to  $\pm 0.1$  V scale of RWSLFP as given in Mazzoni, Lindén et al., 2015. RWSLFP outputs unnormalized LFP instead of this  $\pm 0.1$  V range to sidestep the complications of normalizing in a causal, stepwise manner.

```

fig, axs = plt.subplots(1, 2, figsize=(6, 7), sharey=True, layout="constrained")
channel_offsets = -12 * np.arange(probe.n)
lfp_to_plot = tkLFP.lfp / b2.uvolt + channel_offsets
axs[0].plot(lfp_to_plot, color="w", lw=1)
axs[0].set(
    yticks=channel_offsets,
    yticklabels=range(1, 33),

```

(continues on next page)

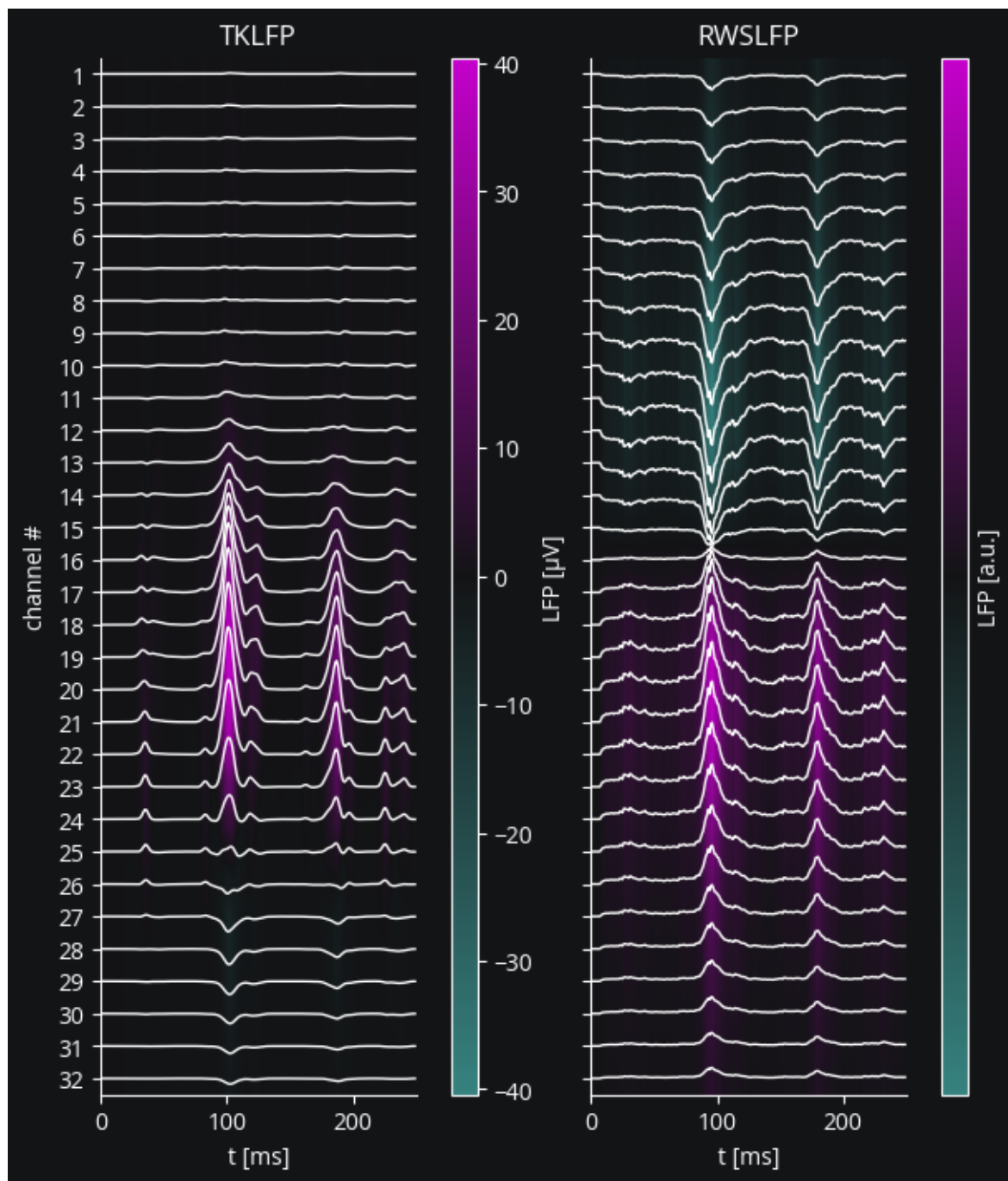
(continued from previous page)

```
        xlabel="t [ms]",
        ylabel="channel #",
        title="TKLFP",
    )

    axs[1].plot(rwslfp.lfp + channel_offsets, color="w", lw=1)
    axs[1].set(title="RWSLFP", xlabel="t [ms]")

    extent = (0, 250, channel_offsets.min() - 6, channel_offsets.max() + 6)
    cmap = LinearSegmentedColormap.from_list("lfp", [c["accent"], "#131416", c["main"]])
    im1 = axs[0].imshow(
        tkf.lfp.T / b2.uvolt,
        aspect="auto",
        cmap=cmap,
        extent=extent,
        vmin=-np.max(np.abs(tkf.lfp / b2.uvolt)),
        vmax=np.max(np.abs(tkf.lfp / b2.uvolt)),
    )
    fig.colorbar(im1, aspect=40, label="LFP [V]")
    im2 = axs[1].imshow(
        rwslfp.lfp.T,
        aspect="auto",
        cmap=cmap,
        extent=extent,
        vmin=-np.max(np.abs(rwslfp.lfp)),
        vmax=np.max(np.abs(rwslfp.lfp)),
    )
    fig.colorbar(im2, aspect=40, label="LFP [a.u.]", ticks=[]);
```





See [Advanced LFP](#) for more options and comparison of different LFP signals.

## 6.2.2 Optogenetic stimulation

How to inject an optogenetic intervention (opsin and optic fiber) into a simulation.

Preamble:

```
%load_ext autoreload
%autoreload 2
from brian2 import *
import matplotlib.pyplot as plt

import cleo
from cleo import *

cleo.utilities.style_plots_for_docs()

# numpy faster than cython for lightweight example
prefs.codegen.target = 'numpy'
# for reproducibility
np.random.seed(1866)
```

The autoreload extension is already loaded. To reload it, use:

```
%reload_ext autoreload
```

### Create a Markov opsin-compatible network

Cleo enables two basic approaches to modeling opsin currents. One is a fairly accurate Markov state model and the other is a simple proportional current model. We will look at the Markov model here.

The established Markov opsin models (as presented in [Evans et al., 2016](#)), are conductance-based and so depend on somewhat realistic membrane voltages. Hence, we'll use the [exponential integrate-and-fire \(EIF\) model](#). Note that we follow the conventions used in neuron modeling, where current is positive, rather than the conventions in opsin modeling, where the photocurrent is negative.

We'll use a small neuron group, biased by Poisson input spikes.

```
n = 10
ng = NeuronGroup(
    n,
    """
    dv/dt = -(v - E_L) + Delta_T*exp((v-theta)/Delta_T) + Rm*I) / tau_m : volt
    I : amp
    """,
    threshold="v>30*mV",
    reset="v=-55*mV",
    namespace={
        "tau_m": 20 * ms,
        "Rm": 500 * Mohm,
        "theta": -50 * mV,
        "Delta_T": 2 * mV,
        "E_L": -70*mV,
    },
)
ng.v = -70 * mV
```

(continues on next page)

(continued from previous page)

```

input_group = PoissonInput(ng, "v", n, 100 * Hz, 1 * mV)

mon = SpikeMonitor(ng)

net = Network(ng, input_group, mon)
ng.equations

```

$$\frac{dv}{dt} = \frac{\Delta_T e^{\frac{-\theta+v}{\Delta_T}} + E_L + IRm - v}{\tau_m} \quad (\text{unit of } v: \text{V})$$

(unit: A)

### Assign coordinates and configure optogenetic model

The `OptogeneticIntervention` class implements the chosen opsin kinetics model with specified parameters. A standard four-state Markov model as well as channelrhodopsin-2 (ChR2) parameters are included with `cleo` and are accessible in the `cleo.opto` module. For extending to other models (such as three-state or six-state), see the [source code](#)—the state equations, opsin-specific parameters, and light wavelength-specific parameters (if not using 473-nm blue) would be needed.

For reference, `cleo` draws heavily on [Foutz et al., 2012](#) for the light propagation model and on [Evans et al., 2016](#) for the opsin kinetics model.

```

from cleo.coords import assign_coords_rand_rect_prism
from cleo.opto import *
from cleo.light import *

assign_coords_rand_rect_prism(ng, xlim=(-0.1, 0.1), ylim=(-0.1, 0.1), zlim=(0.4, 0.6))

opsin = chr2_4s()
fiber = Light(
    coords=(0, 0, 0.2) * mm,
    light_model=fiber473nm(),
    name="fiber",
)

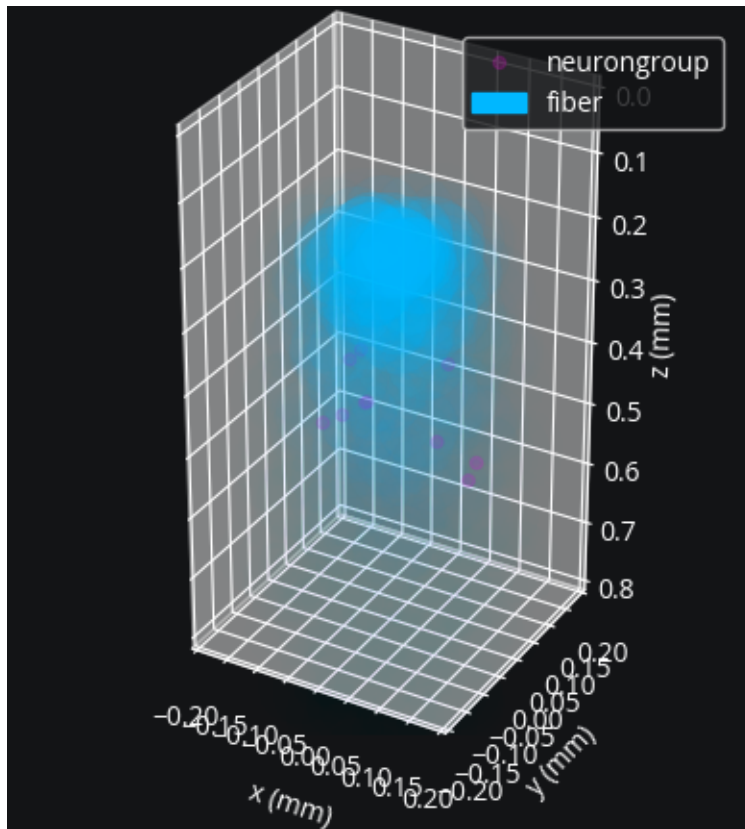
cleo.viz.plot(
    ng,
    colors=["xkcd:fuchsia"],
    xlim=(-0.2, 0.2),
    ylim=(-0.2, 0.2),
    zlim=(0, 0.8),
    devices=[fiber],
)

```

```

(<Figure size 640x480 with 1 Axes>,
 <Axes3D: xlabel='x (mm)', ylabel='y (mm)', zlabel='z (mm)'>)

```



## Open-loop optogenetic stimulation

We need to inject our optogenetic intervention into the simulator. `cleo` handles all the object creation and equations needed to interact with the existing Brian model without the need to alter it, with the possible exception of adding a variable to represent the opsin current. This needs to be specified upon injection with `Iopto_var_name=...` if not the default `Iopto`. The membrane potential variable name also needs to be specified (with `v_var_name=...`) if not the default `v`.

```
sim = CLSimulator(net)
sim.inject(opsin, ng, Iopto_var_name='I')
sim.inject(fiber, ng)
```

```
CLSimulator(io_processor=None, devices={FourStateOpsin(brian_objects=
→ {NeuronGroup(clock=Clock(dt=100. * usecond, name='defaultclock'), when=start, order=0,
→ name='light_agg_ChR2_neurongroup'), Synapses(clock=Clock(dt=100. * usecond, name=
→ 'defaultclock'), when=start, order=0, name='syn_ChR2_neurongroup')}), sim=..., name=
→ 'ChR2', save_history=True, spectrum=[(400, 0.34), (422, 0.65), (460, 0.96), (470, 1),
→ (473, 1), (500, 0.57), (520, 0.22), (540, 0.06), (560, 0.01)], required_vars=[('Iopto',
→ amp), ('v', volt)], g0=114. * nsiemens, gamma=0.00742, phim=2.33e+23 * (second ** -1)
→ / (meter ** 2), k1=4.15 * khertz, k2=0.868 * khertz, p=0.833, Gf0=37.3 * hertz, kf=58.
→ 1 * hertz, Gb0=16.1 * hertz, kb=63. * hertz, q=1.94, Gd1=105. * hertz, Gd2=13.8 *
→ hertz, Gr0=0.33 * hertz, E=0. * volt, v0=43. * mvolt, v1=17.1 * mvolt, model="\n
→ dC1/dt = Gd1*O1 + Gr0*C2 - Ga1*C1 : 1 (clock-driven)\n          dO1/dt = Ga1*C1 + Gb*O2
→ - (Gd1+Gf)*O1 : 1 (clock-driven)\n          dO2/dt = Ga2*C2 + Gf*O1 - (Gd2+Gb)*O2 : 1
→ (clock-driven)\n          C2 = 1 - C1 - O1 - O2 : 1\n\n          Theta = int(phi_pre >
```

(continues on next page)

(continued from previous page)

```

→ 0*phi_pre) : 1\n      Hp = Theta * phi_pre**p/(phi_pre**p + phim**p) : 1\n      ↵
→ Ga1 = k1*Hp : hertz\n      Ga2 = k2*Hp : hertz\n      Hq = Theta * phi_pre**q/(phi_
→ pre**q + phim**q) : 1\n      Gf = kf*Hq + Gf0 : hertz\n      Gb = kb*Hq + Gb0 : ↵
→ hertz\n\n      fphi = 01 + gamma*02 : 1\n      # v1/v0 when v-E == 0 via l'Hopital
→ 's rule\n      fv = f_unless_x0(\n      (1 - exp(-(V_VAR_NAME_post-E)/v0)) / ↵
→ ((V_VAR_NAME_post-E)/v1),\n      V_VAR_NAME_post - E,\n      v1/v0\n      ↵
→ ) : 1\n\n      IOPT0_VAR_NAME_post = -g0*fphi*f_v*(V_VAR_NAME_post-E)*rho_rel : ↵
→ ampere (summed)\n      rho_rel : 1", extra_namespace={'f_unless_x0': <brian2.core.
→ functions.Function object at 0x7fa988672c80>}), Light(brian_objects=set(), sim=..., ↵
→ name='fiber', save_history=True, value=array([0.]), light_model=OpticFiber(R0=100. * ↵
→ umetre, NAfib=0.37, K=125. * metre ** -1, S=7370. * metre ** -1, ntis=1.36), ↵
→ coords=array([[ 0., 0., 200.]]) * umetre, wavelength=0.473 * umetre, ↵
→ direction=array([0., 0., 1.]), max_Irr0_mW_per_mm2=None, max_Irr0_mW_per_mm2_viz=None, ↵
→ default_value=array([0.])))}

```

## IO processor setup

Here we design an IO processor that ignores measurements and simply sets the light intensity according to the stimulus(t) function:

```

from cleo.ioproc import LatencyIOProcessor

def stimulus(time_ms):
    f = 30
    return 1 * (1 + np.sin(2*np.pi*f * time_ms/1000))

class OpenLoopOpto(LatencyIOProcessor):
    def __init__(self):
        super().__init__(sample_period_ms=1)

    # since this is open-loop, we don't use state_dict
    def process(self, state_dict, time_ms):
        opto_intensity = stimulus(time_ms)
        # return output dict and time
        return ({"fiber": opto_intensity}, time_ms)

sim.set_io_processor(OpenLoopOpto())

```

```

CLSimulator(io_processor=<__main__.OpenLoopOpto object at 0x7fa986024280>, devices=
→ {FourStateOpsin(brian_objects={NeuronGroup(clock=Clock(dt=100. * usecond, name=
→ 'defaultclock'), when=start, order=0, name='light_agg_ChR2_neurongroup'), ↵
→ Synapses(clock=Clock(dt=100. * usecond, name='defaultclock'), when=start, order=0, ↵
→ name='syn_ChR2_neurongroup')}, sim=..., name='ChR2', save_history=True, spectrum=[(400,
→ 0.34), (422, 0.65), (460, 0.96), (470, 1), (473, 1), (500, 0.57), (520, 0.22), (540, ↵
→ 0.06), (560, 0.01)], required_vars=[('Iopto', amp), ('v', volt)], g0=114. * nsiemens, ↵
→ gamma=0.00742, phim=2.33e+23 * (second ** -1) / (meter ** 2), k1=4.15 * khertz, k2=0.
→ 868 * khertz, p=0.833, Gf0=37.3 * hertz, kf=58.1 * hertz, Gb0=16.1 * hertz, kb=63. * ↵
→ hertz, q=1.94, Gd1=105. * hertz, Gd2=13.8 * hertz, Gr0=0.33 * hertz, E=0. * volt, ↵
→ v0=43. * mvolt, v1=17.1 * mvolt, model="\n      dC1/dt = Gd1*01 + Gr0*C2 - Ga1*C1 : ↵
→ 1 (clock-driven)\n      d01/dt = Ga1*C1 + Gb*02 - (Gd1+Gf)*01 : 1 (clock-driven)\n ↵

```

(continues on next page)

(continued from previous page)

```

→ dO2/dt = Ga2*C2 + Gf*O1 - (Gd2+Gb)*O2 : 1 (clock-driven)\n          C2 = 1 - C1 -_
→ O1 - O2 : 1\n\n          Theta = int(phi_pre > 0*phi_pre) : 1\n          Hp = Theta * phi_
→ pre**p/(phi_pre**p + phim**p) : 1\n          Ga1 = k1*Hp : hertz\n          Ga2 = k2*Hp :_
→ hertz\n          Hq = Theta * phi_pre**q/(phi_pre**q + phim**q) : 1\n          Gf = kf*Hq_
→ + Gf0 : hertz\n          Gb = kb*Hq + Gb0 : hertz\n\n          fphi = O1 + gamma*O2 : 1\n_
→ # v1/v0 when v-E == 0 via l'Hopital's rule\n          fv = f_unless_x0(\n          _
→ (1 - exp(-(V_VAR_NAME_post-E)/v0)) / ((V_VAR_NAME_post-E)/v1),\n          V_VAR_
→ NAME_post - E,\n          v1/v0\n          ) : 1\n\n          IOPTO_VAR_NAME_post = -
→ g0*fphi*fvy*(V_VAR_NAME_post-E)*rho_rel : ampere (summed)\n          rho_rel : 1", extra_
→ namespace={'f_unless_x0': <brian2.core.functions.Function object at 0x7fa988672c80>}},_
→ Light(brian_objects=set(), sim=..., name='fiber', save_history=True, value=array([0.]),_
→ light_model=OpticFiber(R0=100. * umetre, NAfib=0.37, K=125. * metre ** -1, S=7370. *_
→ metre ** -1, ntis=1.36), coords=array([[ 0.,  0., 200.]]) * umetre, wavelength=0.473_
→ * umetre, direction=array([0., 0., 1.]), max_Irr0_mW_per_mm2=None, max_Irr0_mW_per_mm2_
→ viz=None, default_value=array([0.])))}

```

## Run simulation and plot results

```

sim.run(100*ms)

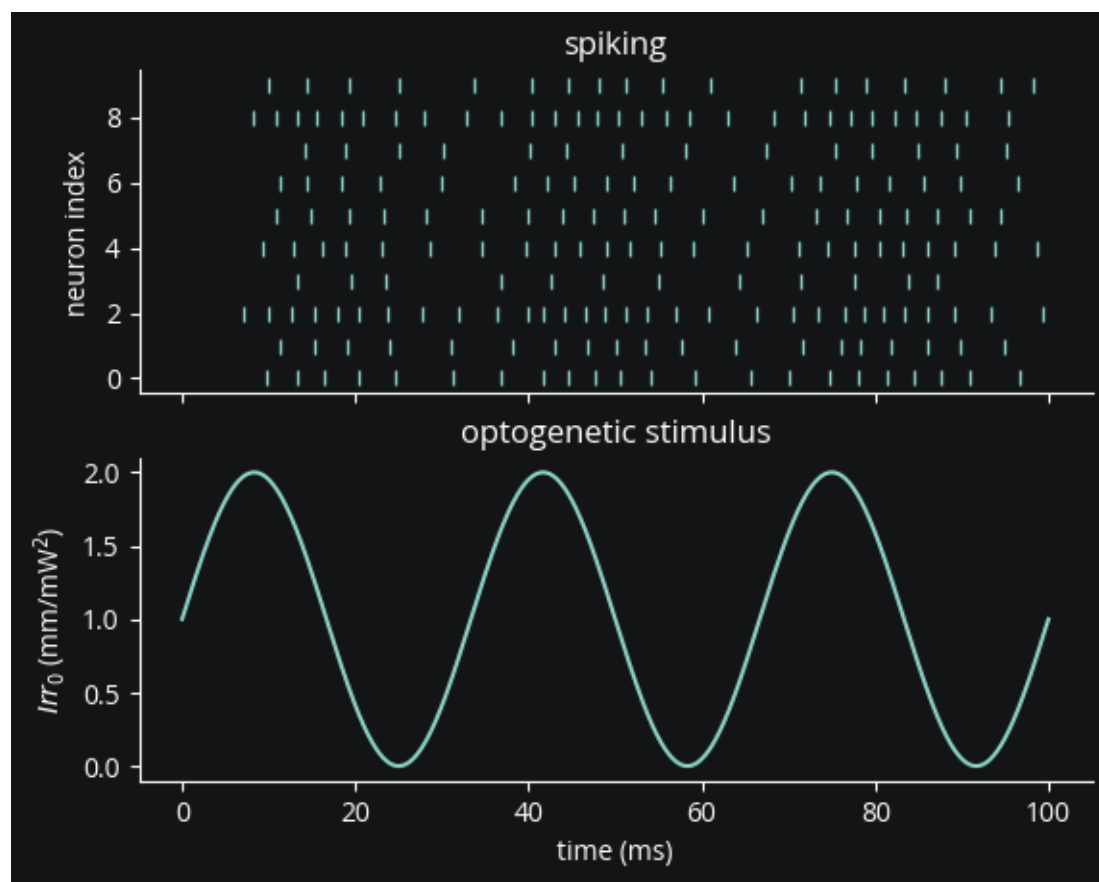
fig, (ax1, ax2) = plt.subplots(2, 1, sharex=True)
ax1.plot(mon.t / ms, mon.i[:, '|'])
ax1.set(ylabel='neuron index', title='spiking')
t_sim=np.linspace(0, 100, 1000)
ax2.plot(t_sim, stimulus(t_sim))
ax2.set(ylabel=r'$Irr_0$ (mm/mW$^2$)', title='optogenetic stimulus', xlabel='time (ms)');

```

```

INFO          No numerical integration method specified for group 'neurongroup', using_
→ method 'euler' (took 0.01s, trying other methods took 0.04s). [brian2.stateupdaters.
→ base.method_choice]
INFO          No numerical integration method specified for group 'syn_ChR2_neurongroup',_
→ using method 'euler' (took 0.02s, trying other methods took 0.05s). [brian2.
→ stateupdaters.base.method_choice]

```



```
opsin.synapses[ng.name].equations
```

$$\begin{aligned}
C2 &= -C_1 - O_1 - O_2 + 1 && \text{(unit of } C_2: \text{ rad)} \\
Theta &= \text{int}(\phi_{pre} > 0) && \text{(unit of } \Theta: \text{ rad)} \\
fphi &= O_1 + O_2\gamma && \text{(unit of } fphi: \text{ rad)} \\
fv &= f_{unlessx0} \left( \frac{v_1 \cdot \left(1 - e^{\frac{E - v_{post}}{v_0}}\right)}{-E + v_{post}}, -E + v_{post}, \frac{v_1}{v_0} \right) && \text{(unit of } fv: \text{ rad)} \\
Hp &= \frac{\Theta \phi_{pre}^p}{\phi_{pre}^p + phimp} && \text{(unit of } Hp: \text{ rad)} \\
Hq &= \frac{\Theta \phi_{pre}^q}{\phi_{pre}^q + phimq} && \text{(unit of } Hq: \text{ rad)} \\
Ga1 &= Hpk_1 && \text{(unit of } Ga_1: \text{ Hz)} \\
Ga2 &= Hpk_2 && \text{(unit of } Ga_2: \text{ Hz)} \\
Gf &= Gf_0 + Hqkf && \text{(unit of } Gf: \text{ Hz)} \\
Gb &= Gb_0 + Hqkb && \text{(unit of } Gb: \text{ Hz)} \\
\frac{dC_1}{dt} &= -C_1 Ga_1 + C_2 Gr_0 + Gd_1 O_1 && \text{(unit of } C_1: \text{ rad, flags: clock-driven)} \\
\frac{dO_1}{dt} &= C_1 Ga_1 + Gb O_2 - O_1 (Gd_1 + Gf) && \text{(unit of } O_1: \text{ rad, flags: clock-driven)} \\
\frac{dO_2}{dt} &= C_2 Ga_2 + Gf O_1 - O_2 (Gb + Gd_2) && \text{(unit of } O_2: \text{ rad, flags: clock-driven)} \\
\rho_{rel} & && \text{(unit: rad)}
\end{aligned}$$

## Conclusion

We can see clearly that firing rate correlates with light intensity as expected.

As a recap, in this tutorial we've seen how to:

- configure an `OptogeneticIntervention`,
- inject it into the simulation,
- and control its light intensity in an open-loop fashion.

## Appendix: alternative opsin and neuron models

Because it would be a pain and an obstacle to reproducibility to have to replace all pre-existing simple neuron models with more sophisticated ones with proper voltage ranges and units, we provide an approximation that is much more flexible, requiring only a current term, of any unit, in the target neurons.

The Markov models of opsin dynamics we've used so far produce a rise, peak, and fall to a steady-state plateau current when subjected to sustained light. Since they are conductance-based, the current also varies with membrane voltage, including during spikes. The `ProportionalCurrentModel`, on the other hand, simply delivers current proportional to light intensity. This should be adequate for a wide range of use cases where the exact opsin current dynamics on short timescales don't matter so much and a sort of average current-light relationship will suffice.

Speaking of realistic membrane voltages, does the Markov model's voltage-dependent current render it unsuitable for the most basic leaky integrate-and-fire (LIF) neuron model? LIF neurons reset on reaching their rheobase threshold,



staying perpetually in a subthreshold region producing exaggerated opsin currents. How much does this affect the output? We will explore this question by comparing a variety of opsin/neuron model combinations.

First, we introduce exponential integrate-and-fire neurons, which maintain simplicity while modeling an upward membrane potential swing during a spike. For more info, see the [related section in the Neuronal Dynamics online textbook](#) and their [example parameters table](#).

```
neuron_params = {
    "tau_m": 20 * ms,
    "Rm": 500 * Mohm,
    "theta": -50 * mV,
    "Delta_T": 2 * mV,
    "E_L": -70*mV,
}

def prep_ng(ng, neuron_type, markov_opsin):
    ng.v = neuron_params['E_L']
    assign_coords_rect_prism(ng, xlim=(0, 0), ylim=(0, 0), zlim=(0, 0))
    state_mon = StateMonitor(ng, ("Iopto", "v"), record=True)
    spike_mon = SpikeMonitor(ng)
    return neuron_type, ng, state_mon, spike_mon, markov_opsin

experiments = []

eif = NeuronGroup(
    1,
    """
    dv/dt = (-(v - E_L) + Delta_T*exp((v-theta)/Delta_T) + Rm*Iopto) / tau_m : volt
    Iopto : amp
    """,
    threshold="v > -10*mV",
    reset="v = E_L - 0*mV",
    namespace=neuron_params,
)

experiments.append(prepare_ng(eif, 'EIF', True))
```

## Configure LIF models

Here we define LIF neurons with biological parameters for the sake of comparison, but the ProportionalCurrentModel is compatible with models of any voltage range and units, so long as it has an Iopto term.

```
def prep_lif(markov_opsin):
    ng = NeuronGroup(
        1,
        """dv/dt = (-(v - E_L) + Rm*Iopto) / tau_m : volt
        Iopto : amp""",
        threshold="v > theta + 4*mV",
        reset="v = E_L - 0*mV",
        namespace=neuron_params,
    )
    return prep_ng(ng, "LIF", markov_opsin)
```

(continues on next page)

(continued from previous page)

```
experiments.append(prepare_lif(True))
experiments.append(prepare_lif(False))
```

## Comparing to more realistic models

To see how well simplified neuron and opsin models do, we'll also compare to the more complex `AdEx neuron` (with “tonic” firing pattern parameters) and a Hodgkin-Huxley model (code from `Neuronal Dynamics`).

```
adex = NeuronGroup(
    1,
    """dv/dt = -(v - E_L) + 2*mV*exp((v-theta)/Delta_T) + Rm*(Iopto-w) / tau_m : volt
    dw/dt = (0*nsiemens*(v-E_L) - w) / (100*ms) : amp
    Iopto : amp""",
    threshold="v>=-10*mV",
    reset="v=-55*mV; w+=5*pamp",
    namespace=neuron_params,
)
experiments.append(prepare_ng(adex, "AdEx", True))

# Parameters
# Cm = 1*ufarad*cm**-2 * area
Cm = neuron_params["tau_m"] / neuron_params["Rm"]
# area = 5000*umetre**2
area = Cm / (1*ufarad*cm**-2)
gl = 0.3*msiemens*cm**-2 * area
El = -65*mV
EK = -90*mV
ENa = 50*mV
g_na = 40*msiemens*cm**-2 * area
g_kd = 35*msiemens*cm**-2 * area
VT = -63*mV

# The model
eqs = Equations('''
dv/dt = (gl*(El-v) - g_na*(m*m*m)*h*(v-ENa) - g_kd*(n*n*n*n)*(v-EK) + Iopto)/Cm : volt
dm/dt = 0.32*(mV**-1)*4*mV/exprel((13.*mV-v+VT)/(4*mV))/ms*(1-m)-0.28*(mV**-1)*5*mV/
    exprel((v-VT-40.*mV)/(5*mV))/ms*m : 1
dn/dt = 0.032*(mV**-1)*5*mV/exprel((15.*mV-v+VT)/(5*mV))/ms*(1-n)-.5*exp((10.*mV-v+VT)/
    (40.*mV))/ms*n : 1
dh/dt = 0.128*exp((17.*mV-v+VT)/(18.*mV))/ms*(1-h)-4./(1+exp((40.*mV-v+VT)/(5.*mV)))/
    ms*h : 1
Iopto : amp
''')
# Threshold and refractoriness are only used for spike counting
hh = NeuronGroup(1, eqs,
    threshold='v > -40*mV',
    reset='',
    method='exponential_euler')
experiments.append(prepare_ng(hh, "HH", True))
```

## Opsin configuration

Note that the only parameter we need to set for the simple opsin model is the gain on light intensity, `I_per_Irr`. This term defines what the neuron receives for every 1 mW/mm2 of light intensity. Here that term is defined in amperes, but it could have been unitless for a simpler model.

The gain is tuned somewhat by hand (in relation to the membrane resistance and the 20 mV gap between rest and threshold potential) to achieve similar outputs to the Markov model.

```
light = Light(light_model=fiber473nm())
simple_opsin = ProportionalCurrentOpsin(
    name="simple_opsin",
    # handpicked gain to make firing rate roughly comparable to EIF
    I_per_Irr=140/neuron_params['Rm']*20*mV,
)
markov_opsin = chr2_4s()
markov_opsin.name = "markov_opsin"
```

## Simulation

And we set up the simulator:

```
net = Network()
sim = CLSimulator(net)
for ng_type, ng, state_mon, spike_mon, use_markov_opsin in experiments:
    net.add(ng, state_mon, spike_mon)
    sim.inject(light, ng)
    if use_markov_opsin:
        sim.inject(markov_opsin, ng)
    else:
        sim.inject(simple_opsin, ng)
```

We'll now run the simulation with light pulses of increasing amplitudes to observe the effect on the current.

```
# hand-picked range of amplitudes to show 0 to moderate firing rates
for Irr0_mW_per_mm2 in np.linspace(0.015, 0.05, 5):
    light.update(Irr0_mW_per_mm2)
    sim.run(60 * ms)
    light.update(0)
    sim.run(60 * ms)
```

```
INFO      No numerical integration method specified for group 'neurongroup_1', using
↳method 'euler' (took 0.01s, trying other methods took 0.02s). [brian2.stateupdaters.
↳base.method_choice]
INFO      No numerical integration method specified for group 'neurongroup_2', using
↳method 'exact' (took 0.04s). [brian2.stateupdaters.base.method_choice]
INFO      No numerical integration method specified for group 'neurongroup_3', using
↳method 'exact' (took 0.02s). [brian2.stateupdaters.base.method_choice]
INFO      No numerical integration method specified for group 'neurongroup_4', using
↳method 'euler' (took 0.01s, trying other methods took 0.05s). [brian2.stateupdaters.
↳base.method_choice]
```

```

WARNING      'n' is an internal variable of group 'neurongroup_5', but also exists in the_
↳run namespace with the value 10. The internal variable will be used. [brian2.groups.
↳group.Group.resolve.resolution_conflict]
INFO         No numerical integration method specified for group 'syn_markov_opsin_
↳neurongroup_1', using method 'euler' (took 0.01s, trying other methods took 0.01s).
↳[brian2.stateupdaters.base.method_choice]
INFO         No numerical integration method specified for group 'syn_markov_opsin_
↳neurongroup_2', using method 'euler' (took 0.01s, trying other methods took 0.01s).
↳[brian2.stateupdaters.base.method_choice]
INFO         No numerical integration method specified for group 'syn_markov_opsin_
↳neurongroup_4', using method 'euler' (took 0.01s, trying other methods took 0.01s).
↳[brian2.stateupdaters.base.method_choice]
INFO         No numerical integration method specified for group 'syn_markov_opsin_
↳neurongroup_5', using method 'euler' (took 0.01s, trying other methods took 0.01s).
↳[brian2.stateupdaters.base.method_choice]

```

## Results

```

c1 = '#8000b4'
c2 = '#df87e1'

fig, axs = plt.subplots(
    len(experiments), 1, figsize=(8, 2*len(experiments)), sharex=True
)

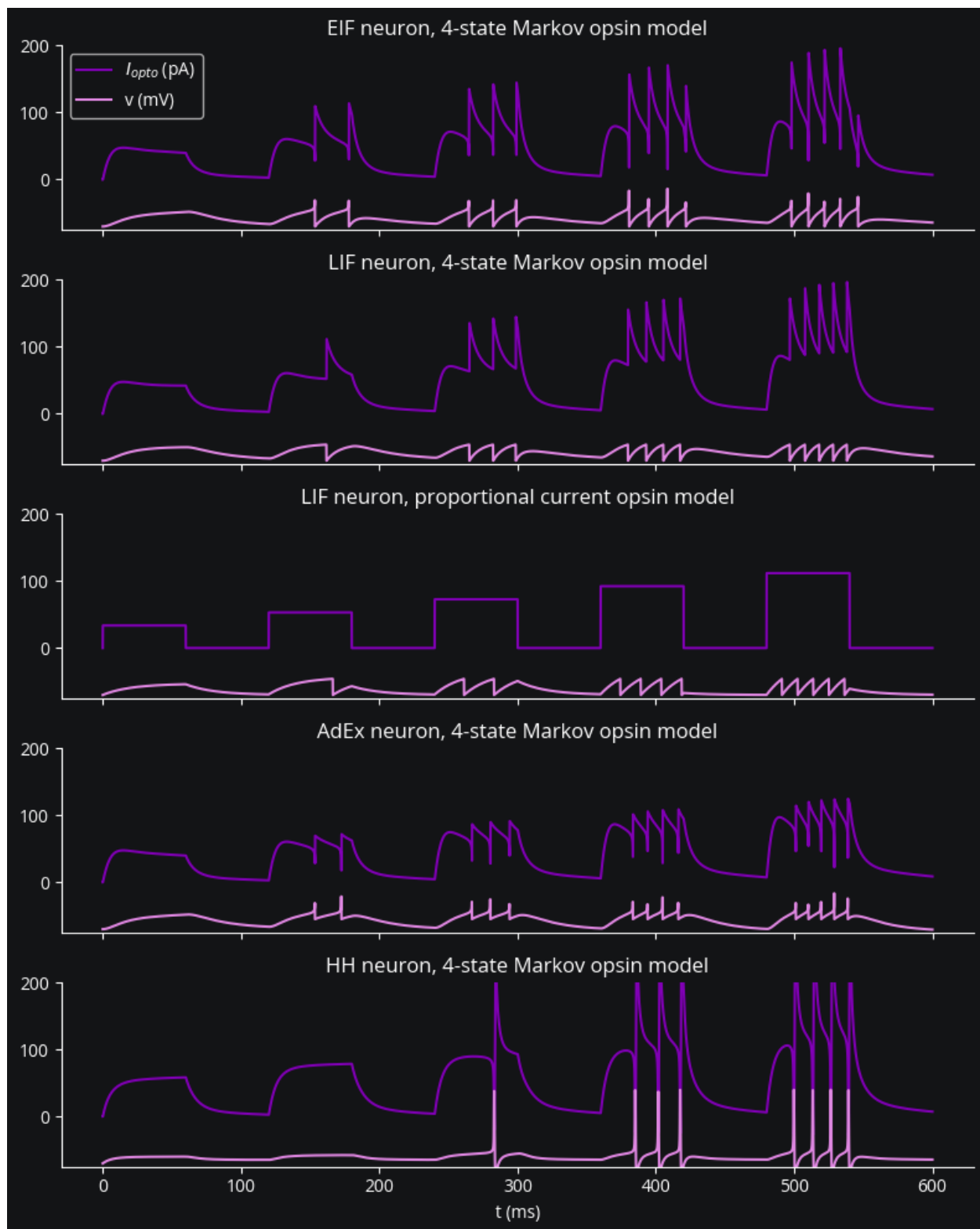
for ax, (ng_type, _, state_mon, spike_mon, markov_opsin) in zip(axs, experiments):
    ax.plot(state_mon.t / ms, state_mon.Iopto[0] / pamp, c=c1, label="$I_{opto}$ (pA)")
    ax.plot(state_mon.t / ms, state_mon.v[0] / mV, c=c2, label="v (mV)")
    opsin_name = "4-state Markov" if markov_opsin else "proportional current"
    ax.set(title=f"{ng_type} neuron, {opsin_name} opsin model")

axs[-1].set_xlabel('t (ms)')
axs[0].legend();

max_ylim = max([ax.get_ylim()[1] for ax in axs])
for ax in axs:
    ax.set_ylim([-75, 200])

fig.tight_layout()

```



Qualitatively we can see that the proportional current model doesn't capture the rise, peak, plateau, and fall dynamics that a Markov model can produce, but is a reasonable approximation if all you need is a roughly linear light intensity-firing rate relationship. We also see that a variety of neuron/opsin model combinations all produce similar firing responses to light.

### 6.2.3 Multi-channel, bidirectional optogenetics

Cleo supports the simultaneous use of multiple Light devices, multiple channels per device, and multiple opsins per neuron group. Here we'll see how to use all these features.

```
# boilerplate
%load_ext autoreload
%autoreload 2
from brian2 import *
import matplotlib.pyplot as plt

import cleo
from cleo import *

cleo.utilities.style_plots_for_docs()

# numpy faster than cython for lightweight example
prefs.codegen.target = 'numpy'
# for reproducibility
np.random.seed(1866)

# colors
c = {
    'main': '#C500CC',
    '473nm': '#72b5f2',
    '590nm': (1, .875, 0),
}
```

The autoreload extension is already loaded. To reload it, use:

```
%reload_ext autoreload
```

#### Network setup

We'll use excitatory and inhibitory populations of exponential integrate-and-fire neurons.

```
n = 500
ng = NeuronGroup(
    n,
    """
    dv/dt = (
        -(v - E_L)
        + Delta_T*exp((v-theta)/Delta_T)
        + Rm*(I_exc + I_inh + I_bg)
    ) / tau_m + sigma*sqrt(2/tau_m)*xi: volt
    I_exc : amp
    I_inh : amp
    """,
    threshold="v > -10*mV",
    reset="v=E_L",
    namespace={
        "tau_m": 20 * ms,
        "Rm": 500 * Mohm,
```

(continues on next page)

(continued from previous page)

```

        "theta": -50 * mV,
        "Delta_T": 2 * mV,
        "E_L": -70*mV,
        "sigma": 5 * mV,
        "I_bg": 60 * pamp,
    },
)
ng.v = np.random.uniform(-70, -50, n) * mV

W = 1 * mV
p_S = 0.3
n_neighbors = 40
S_ee = Synapses(ng, model='w: 1', on_pre="v_post+=W*w/sqrt(N)")
S_ee.connect(condition='abs(i-j)<=n_neighbors and i!=j')
S_ee.w = np.exp(np.random.randn(int(S_ee.N - 0))) * np.random.choice([-1, 1], int(S_ee.N - 0))
    ↪ - 0))

spike_mon = SpikeMonitor(ng)
# for visualizing currents
Imon = StateMonitor(ng, ('I_exc', 'I_inh'), record=range(50))
net = Network(ng, S_ee, spike_mon, Imon)
sim = cleo.CLSimulator(net)

```

```

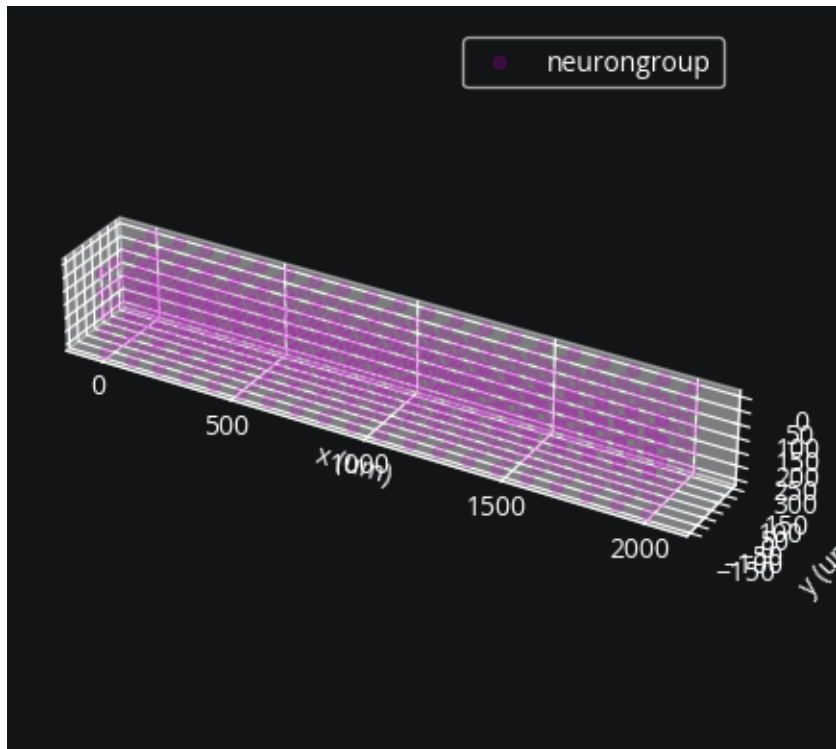
from cleo.coords import assign_coords_grid_rect_prism
xmax_mm = 2
assign_coords_grid_rect_prism(ng, xlim=(0, xmax_mm), ylim=(-.15, .15), zlim=(0, .3),
    ↪ shape=(20, 5, 5))
cleo.viz.plot(ng, colors=[c['main']])

```

```

(<Figure size 640x480 with 1 Axes>,
 <Axes3D: xlabel='x (um)', ylabel='y (um)', zlabel='z (um)'>)

```



### Injecting a multi-channel Light

A `Light` device can have multiple channels; all the user needs is to specify the coordinates (and optionally direction) of each light source (channel). A `LightModel` (e.g., that of an optical fiber) defines how light propagates from each source.

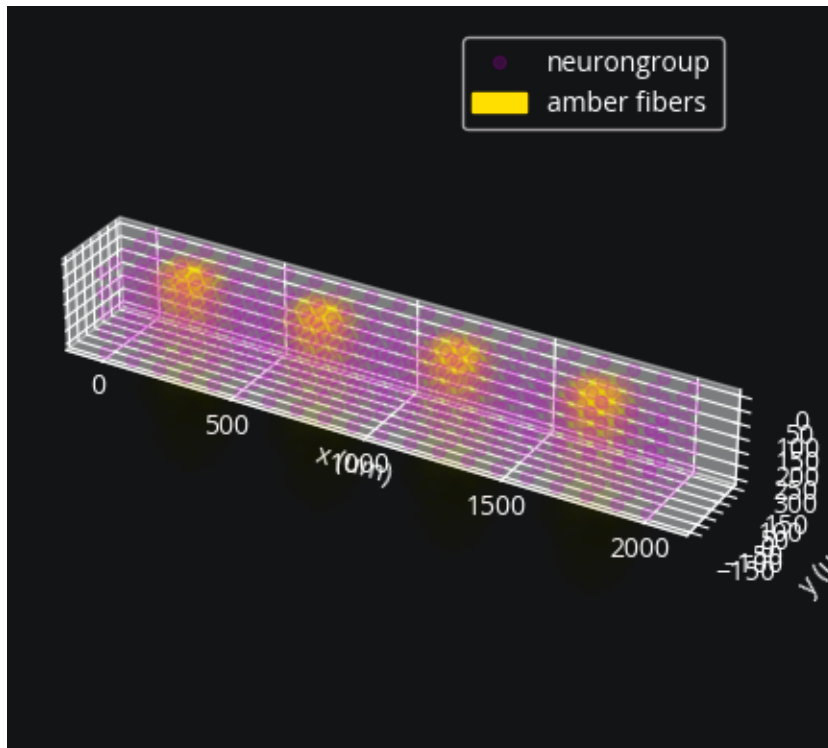
Here we inject 590 nm light for activating Vf-Chrimson. Lacking a more rigorous quantification, we assume absorption and scattering coefficients of 590 nm light in the brain are roughly 0.8 times that of 470 nm light (see [Jacques 2013](#) for some justification).

```
from cleo.light import Light, OpticFiber
from cleo.opto import vfchrimson_4s

n_fibers = 4
coords = np.zeros((n_fibers, 3))
end_space = 1 / (2 * n_fibers)
coords[:, 0] = np.linspace(end_space, 1 - end_space, n_fibers) * xmax_mm
coords[:, 1] = -0.025
amber_fibers = Light(
    name="amber fibers",
    coords=coords * mm,
    light_model=OpticFiber(K=0.125 * 0.8 / mm, S=7.37 * 0.8 / mm),
    wavelength=590 * nmeter,
)
sim.inject(amber_fibers, ng)
vfc = vfchrimson_4s()
sim.inject(vfc, ng, Iopto_var_name="I_exc")
cleo.viz.plot(ng, colors=[c["main"]], sim=sim)
```



```
(<Figure size 640x480 with 1 Axes>,
<Axes3D: xlabel='x (um)', ylabel='y (um)', zlabel='z (um)'>)
```



### Bidirectional control via a second opsin

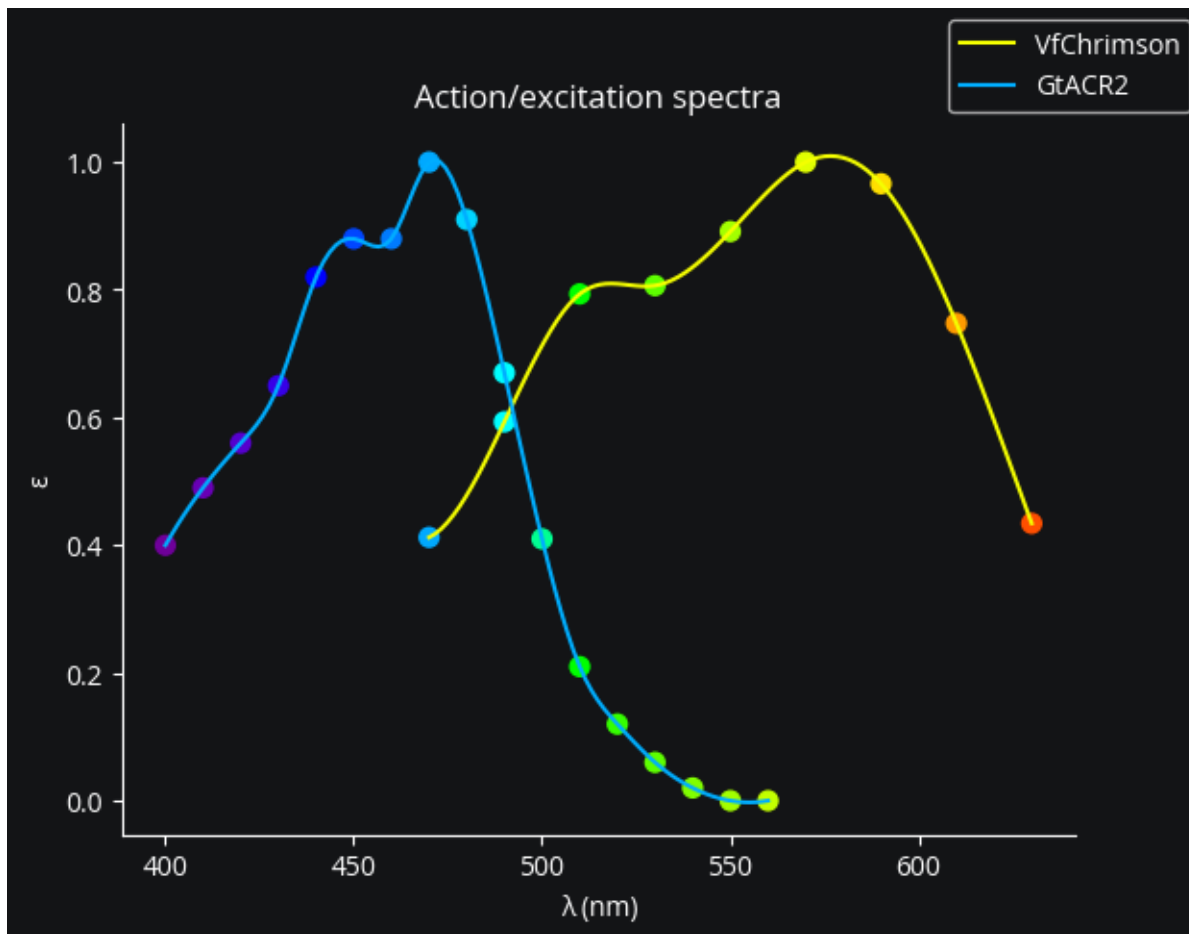
Here we will demonstrate increasing and decreasing activity in the same experiment by injecting an inhibitory opsin with a minimally overlapping activation spectrum. Alternatively, we could achieve bidirectional control with excitatory opsins on excitatory and inhibitory neurons.

We will use the anion channel GtACR2 which is maximally activated by 470 nm light.

Let's first visualize how the action spectra for the two opsins overlap:

```
gtacr2 = cleo.opto.gtacr2_4s()
cleo.light.plot_spectra(vfc, gtacr2)
```

```
(<Figure size 640x480 with 1 Axes>,
<Axes: title={'center': 'Action/excitation spectra'}, xlabel=' (nm)', ylabel=''>)
```

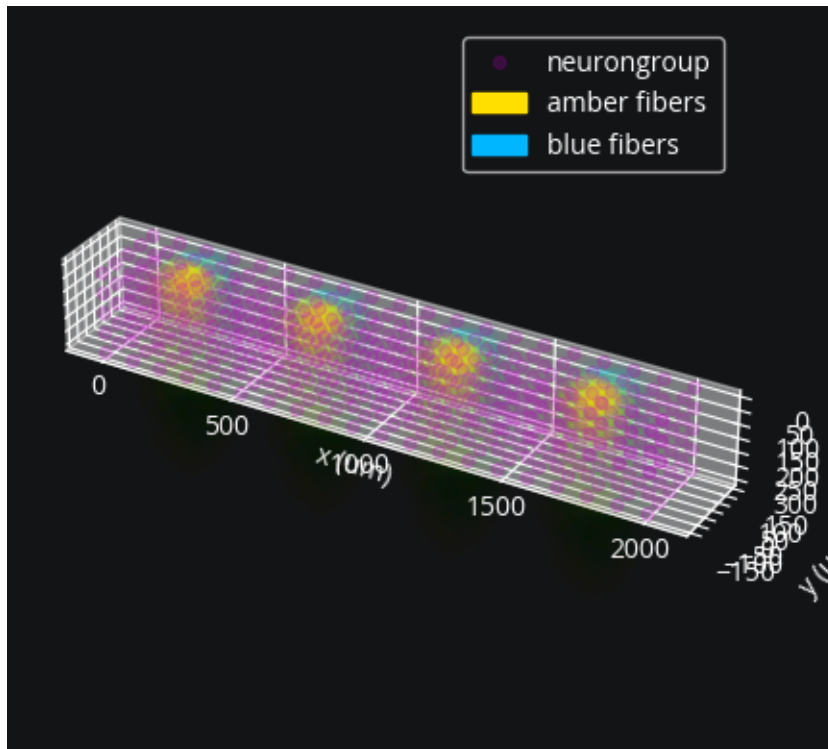


We see that GtACR2 will be totally unaffected by amber (590 nm) light, while Vf-Chrimson will be slightly activated by blue (470 nm) light.

```
coords[:, 1] = 0.025
blue_fibers = Light(
    name="blue fibers",
    coords=coords * mm,
    light_model=cleo.light.fiber473nm(),
)
sim.inject(blue_fibers, ng)
sim.inject(gtacr2, ng, Iopto_var_name="I_inh")
cleo.viz.plot(ng, colors=[c["main"]], sim=sim)
```

```
WARNING /home/kyle/Dropbox (GaTech)/projects/cleo/cleo/light/light_dependence.py:107:
↳UserWarning: = 590.0 nm is outside the range of the action spectrum data for GtACR2.
↳Assuming = 0.
    warnings.warn(
[py.warnings]
```

```
(<Figure size 640x480 with 1 Axes>,
<Axes3D: xlabel='x (um)', ylabel='y (um)', zlabel='z (um)'>)
```



## Open-loop stimulation

We will now design a stimulus pattern to demonstrate bidirectional control segregated by channel.

```
from cleo.ioproc import LatencyIOProcessor

class OpenLoopOpto(LatencyIOProcessor):
    def __init__(self):
        super().__init__(sample_period_ms=1)

    # since this is open-loop, we don't use state_dict
    def process(self, state_dict, time_ms):
        amplitude_mW_mm2 = 1
        time_offsets = np.array([0, -20, -40, -60])
        t = time_ms + time_offsets
        amber = ((t >= 20) & (t < 40)) * amplitude_mW_mm2
        blue = ((t >= 60) & (t < 63)) * amplitude_mW_mm2

        # return output dict and time
        return ({"amber fibers": amber, "blue fibers": blue}, time_ms)

sim.set_io_processor(OpenLoopOpto())
```

```
CLSimulator(io_processor=<__main__.OpenLoopOpto object at 0x7fe758a361d0>, devices=
→ {BansalFourStateOpsin(sim=..., name='GtACR2', save_history=True, on_pre='',
→ spectrum=[(400, 0.4), (410, 0.49), (420, 0.56), (430, 0.65), (440, 0.82), (450, 0.88),
→ (460, 0.88), (470, 1.0), (480, 0.91), (490, 0.67), (500, 0.41), (510, 0.21), (520, 0.
→ 12), (530, 0.06), (540, 0.02), (550, 0.0), (560, 0.0)], required_vars=['Iopto', amp],
```

(continues on next page)

(continued from previous page)

```

→('v', volt)], Gd1=17. * hertz, Gd2=10. * hertz, Gr0=0.58 * hertz, g0=44. * nsiemens,
→phim=2.e+23 * (second ** -1) / (meter ** 2), k1=40. * khertz, k2=20. * khertz, Gf0=1.
→* hertz, Gb0=3. * hertz, kf=1. * hertz, kb=5. * hertz, gamma=0.05, p=1, q=0.1, E=-69.5
→* mvolt, model='\n          dC1/dt = Gd1*O1 + Gr0*C2 - Ga1*C1 : 1 (clock-driven)\n
→dO1/dt = Ga1*C1 + Gb*O2 - (Gd1+Gf)*O1 : 1 (clock-driven)\n          dO2/dt = Ga2*C2 +
→Gf*O1 - (Gd2+Gb)*O2 : 1 (clock-driven)\n          C2 = 1 - C1 - O1 - O2 : 1\n\n
→Theta = int(phi_pre > 0*phi_pre) : 1\n          Hp = Theta * phi_pre**p/(phi_pre**p +
→phim**p) : 1\n          Ga1 = k1*Hp : hertz\n          Ga2 = k2*Hp : hertz\n          Hq =
→Theta * phi_pre**q/(phi_pre**q + phim**q) : 1\n          Gf = kf*Hq + Gf0 : hertz\n
→Gb = kb*Hq + Gb0 : hertz\n\n          fphi = O1 + gamma*O2 : 1\n\n          IOPTO_VAR_
→NAME_post = -g0*fphi*(V_VAR_NAME_post-E)*rho_rel : ampere (summed)\n          rho_rel : 1
→'), Light(sim=..., name='amber fibers', save_history=True, value=array([0., 0., 0., 0.
→]), light_model=OpticFiber(R0=100. * umetre, NAfib=0.37, K=100. * metre ** -1, S=5896.
→* metre ** -1, ntis=1.36), wavelength=0.59 * umetre, direction=array([0., 0., 1.]),
→max_Irr0_mW_per_mm2=None, max_Irr0_mW_per_mm2_viz=None), BansalFourStateOpsin(sim=...,
→name='VfChrimson', save_history=True, on_pre='', spectrum=[(470.0, 0.4123404255319149),
→(490.0, 0.593265306122449), (510.0, 0.7935294117647058), (530.0, 0.8066037735849055),
→(550.0, 0.8912727272727272), (570.0, 1.0), (590.0, 0.9661016949152542), (610.0, 0.
→7475409836065574), (630.0, 0.4342857142857143)], required_vars=[('Iopto', amp), ('v',
→volt)], Gd1=0.37 * khertz, Gd2=175. * hertz, Gr0=0.667 * mhertz, g0=17.5 * nsiemens,
→phim=1.5e+22 * (second ** -1) / (meter ** 2), k1=3. * khertz, k2=200. * hertz, Gf0=20.
→* hertz, Gb0=3.2 * hertz, kf=10. * hertz, kb=10. * hertz, gamma=0.05, p=1, q=1, E=0. *
→volt, model='\n          dC1/dt = Gd1*O1 + Gr0*C2 - Ga1*C1 : 1 (clock-driven)\n
→dO1/dt = Ga1*C1 + Gb*O2 - (Gd1+Gf)*O1 : 1 (clock-driven)\n          dO2/dt = Ga2*C2 +
→Gf*O1 - (Gd2+Gb)*O2 : 1 (clock-driven)\n          C2 = 1 - C1 - O1 - O2 : 1\n\n
→Theta = int(phi_pre > 0*phi_pre) : 1\n          Hp = Theta * phi_pre**p/(phi_pre**p +
→phim**p) : 1\n          Ga1 = k1*Hp : hertz\n          Ga2 = k2*Hp : hertz\n          Hq =
→Theta * phi_pre**q/(phi_pre**q + phim**q) : 1\n          Gf = kf*Hq + Gf0 : hertz\n
→Gb = kb*Hq + Gb0 : hertz\n\n          fphi = O1 + gamma*O2 : 1\n\n          IOPTO_VAR_
→NAME_post = -g0*fphi*(V_VAR_NAME_post-E)*rho_rel : ampere (summed)\n          rho_rel : 1
→'), Light(sim=..., name='blue fibers', save_history=True, value=array([0., 0., 0., 0.
→]), light_model=OpticFiber(R0=100. * umetre, NAfib=0.37, K=125. * metre ** -1, S=7370.
→* metre ** -1, ntis=1.36), wavelength=0.473 * umetre, direction=array([0., 0., 1.]),
→max_Irr0_mW_per_mm2=None, max_Irr0_mW_per_mm2_viz=None)})

```

## Run simulation and plot results

```

sim.reset()
sim.run(200*ms)

```

```

INFO          No numerical integration method specified for group 'neurongroup', using
→method 'euler' (took 0.15s, trying other methods took 0.00s). [brian2.stateupdaters.
→base.method_choice]
INFO          No numerical integration method specified for group 'syn_GtACR2_neurongroup',
→using method 'euler' (took 0.02s, trying other methods took 0.06s). [brian2.
→stateupdaters.base.method_choice]
INFO          No numerical integration method specified for group 'syn_VfChrimson_
→neurongroup', using method 'euler' (took 0.01s, trying other methods took 0.01s).
→[brian2.stateupdaters.base.method_choice]

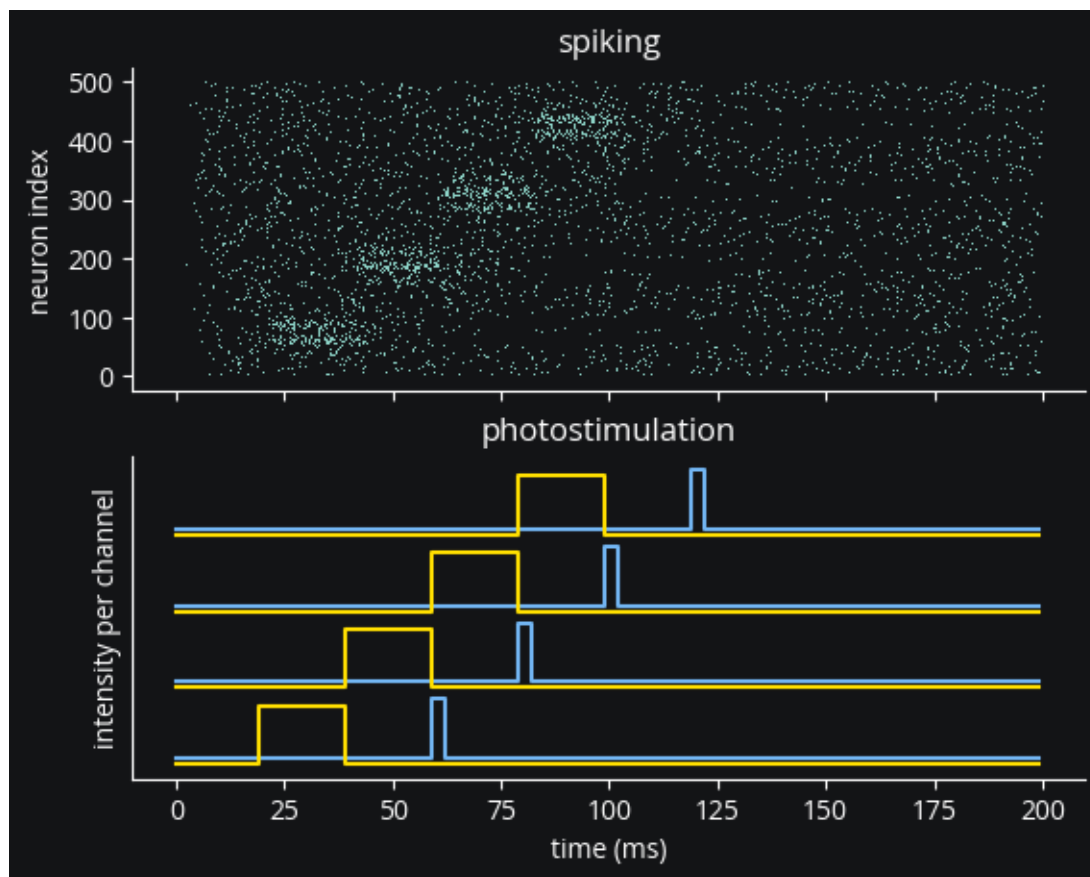
```

```
fig, (ax1, ax2) = plt.subplots(2, 1, sharex=True)

ax1.plot(spike_mon.t / ms, spike_mon.i[:, :], ",")
ax1.set(ylabel="neuron index", title="spiking")

ax2.step(
    blue_fibers.t_ms, blue_fibers.values + np.arange(n_fibers) * 1.3 + 0.1, c=c["473nm"]
)
ax2.step(amber_fibers.t_ms, amber_fibers.values + np.arange(n_fibers) * 1.3, c=c["590nm"])
ax2.set(
    yticks=[],
    ylabel="intensity per channel",
    title="photostimulation",
    xlabel="time (ms)",
)
```

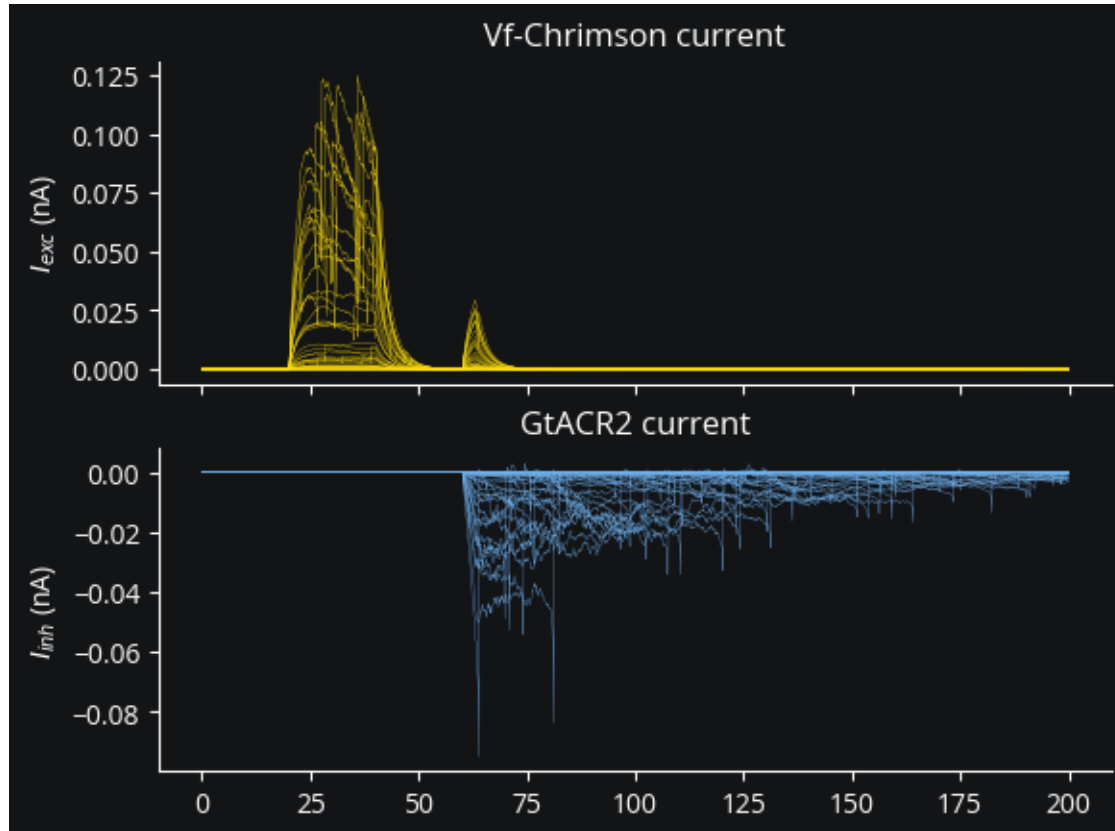
```
[[],
 Text(0, 0.5, 'intensity per channel'),
 Text(0.5, 1.0, 'photostimulation'),
 Text(0.5, 0, 'time (ms)')]
```



As you can see, Vf-Chrimson has fast dynamics, enabling high-frequency control. GtACR2, on the other hand, continues acting long after the light is removed due to its slow deactivation kinetics. We can confirm this by plotting the current from each of the opsins in the first segment of neurons:

```
fig, (ax1, ax2) = plt.subplots(2, 1, sharex=True)
ax1.plot(Imon.t / ms, Imon.I_exc.T / namp, lw=0.2, c=c["590nm"])
ax1.set(title="Vf-Chrimson current", ylabel="$I_{exc}$ (nA)")
ax2.plot(Imon.t / ms, Imon.I_inh.T / namp, lw=0.2, c=c["473nm"])
ax2.set(title="GtACR2 current", ylabel="$I_{inh}$ (nA)")
```

```
[Text(0.5, 1.0, 'GtACR2 current'), Text(0, 0.5, '$I_{inh}$ (nA)')]
```



We can also confirm that blue light has a non-negligible effect on Vf-Chrimson.

## Conclusion

As a recap, in this tutorial we've seen how to:

- configure a multi-channel `Light` device,
- use more than one of them simultaneously, and
- inject multiple opsins of overlapping action spectra into the same network.

## 6.2.4 On-off control

Here we will see how to set up a minimum, working closed loop with a very simple threshold-triggered control scheme.

Preamble:

```
from brian2 import *
from cleo import *

import matplotlib.pyplot as plt

utilities.style_plots_for_docs()

# the default cython compilation target isn't worth it for
# this trivial example
prefs.codegen.target = "numpy"
```

### Set up network

We will use a simple leaky integrate-and-fire network with Poisson spike train input. We use Brian's standard SpikeMonitor to view resulting spikes here for simplicity, but see the electrodes tutorial for a more realistic electrode recording scheme.

```
n = 10
population = NeuronGroup(n, '''
    dv/dt = (-v - 70*mV + Rm*I) / tau : volt
    tau: second
    Rm: ohm
    I: amp''',
    threshold='v>-50*mV',
    reset='v=-70*mV'
)
population.tau = 10*ms
population.Rm = 100*Mohm
population.I = 0*mA
population.v = -70*mV

input_group = PoissonGroup(n, np.linspace(0, 100, n)*Hz + 10*Hz)

S = Synapses(input_group, population, on_pre='v+=5*mV')
S.connect(condition='abs(i-j)<=3')

pop_mon = SpikeMonitor(population)

net = Network([population, input_group, S, pop_mon])

print("Recorded population's equations:")
population.user_equations
```

```
Recorded population's equations:
```

$$\frac{dv}{dt} = \frac{IRm - 70mV - v}{\tau}$$

(unit of  $v$ : V)

(unit: s)

(unit: ohm)

(unit: A)

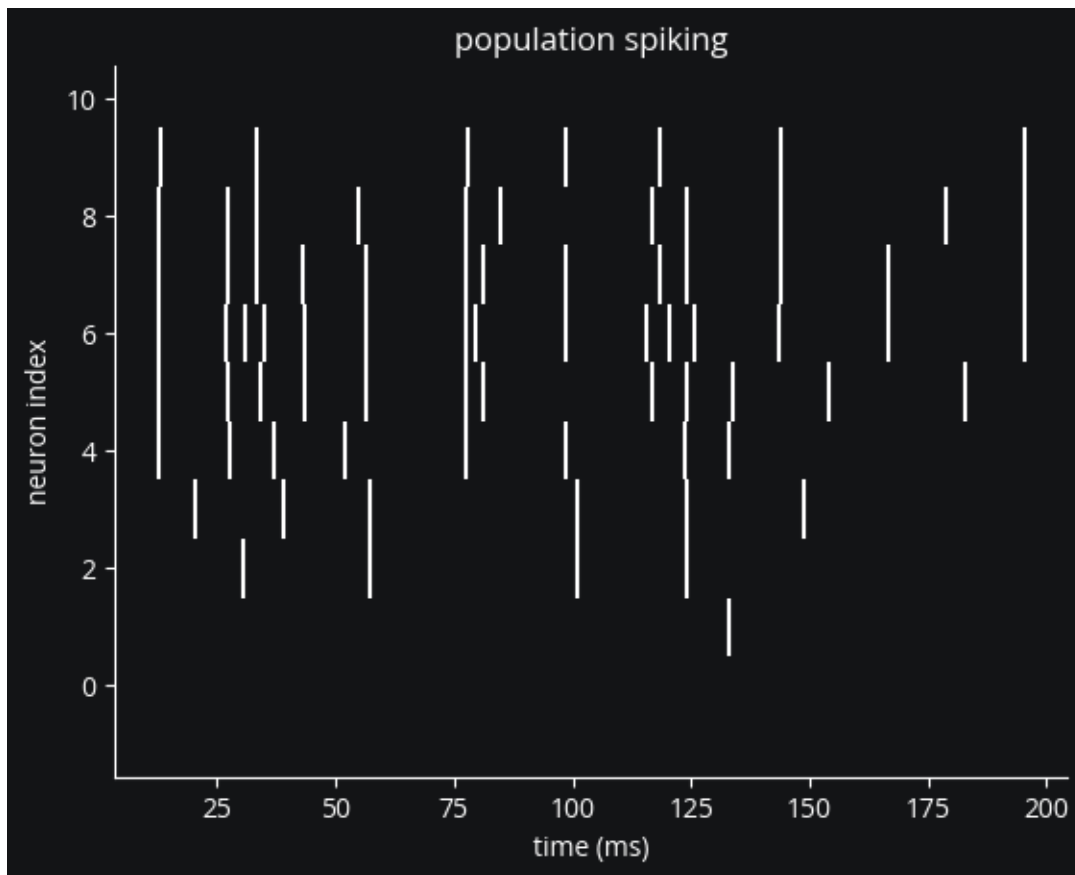
## Run simulation

```
net.run(200*ms)
```

```
INFO      No numerical integration method specified for group 'neurongroup', using
method 'exact' (took 0.15s). [brian2.stateupdaters.base.method_choice]
```

```
sptrains = pop_mon.spike_trains()
fig, ax = plt.subplots()
ax.eventplot([t / ms for t in sptrains.values()], lineoffsets=list(sptrains.keys()))
ax.set(title='population spiking', ylabel='neuron index', xlabel='time (ms)')
```

```
[Text(0.5, 1.0, 'population spiking'),
Text(0, 0.5, 'neuron index'),
Text(0.5, 0, 'time (ms)')]
```



Because lower neuron indices receive very little input, we see no spikes for neuron 0. Let's change that with closed-loop control.



## IO processor setup

We use the `IOProcessor` class to define interactions with the network. To achieve our goal of making neuron 0 fire, we'll use a contrived, simplistic setup where

1. the recorder reports the voltage of a given neuron (of index 5 in our case),
2. the controller outputs a pulse whenever that voltage is below a certain threshold, and
3. the stimulator applies that pulse to the specified neuron.

So if everything is wired correctly, we'll see bursts of activity in just the first neuron.

```
from cleo.recorders import RateRecorder, VoltageRecorder
from cleo.stimulators import StateVariableSetter

i_rec = int(n / 2)
i_ctrl = 0
sim = CLSimulator(net)
v_rec = VoltageRecorder(name="rec")
sim.inject(v_rec, population[i_rec])
sim.inject(
    StateVariableSetter(name="stim", variable_to_ctrl="I", unit=nA), population[i_ctrl]
)
```

```
CLSimulator(io_processor=None, devices={VoltageRecorder(brian_objects={<StateMonitor,
→ recording ['v'] from 'neurongroup_subgroup'>}, sim=..., name='rec', voltage_var_name='v
→ ', mon=<StateMonitor, recording ['v'] from 'neurongroup_subgroup'>),
→ StateVariableSetter(brian_objects=set(), sim=..., name='stim', value=0, default_
→ value=0, save_history=True, variable_to_ctrl='I', unit=namp, neuron_groups=[<Subgroup
→ 'neurongroup_subgroup_1' of 'neurongroup' from 0 to 1>])})})
```

We need to implement the `LatencyIOProcessor` object. For a more sophisticated case we'd use `ProcessingBlock` objects to decompose the computation in the process function.

```
from cleo.ioproc import LatencyIOProcessor

trigger_threshold = -60*mV
class ReactivePulseIOProcessor(LatencyIOProcessor):
    def __init__(self, pulse_current=1):
        super().__init__(sample_period_ms=1)
        self.pulse_current = pulse_current
        self.out = {}

    def process(self, state_dict, time_ms):
        v = state_dict['rec']
        if v is not None and v < trigger_threshold:
            self.out['stim'] = self.pulse_current
        else:
            self.out['stim'] = 0

        return (self.out, time_ms)

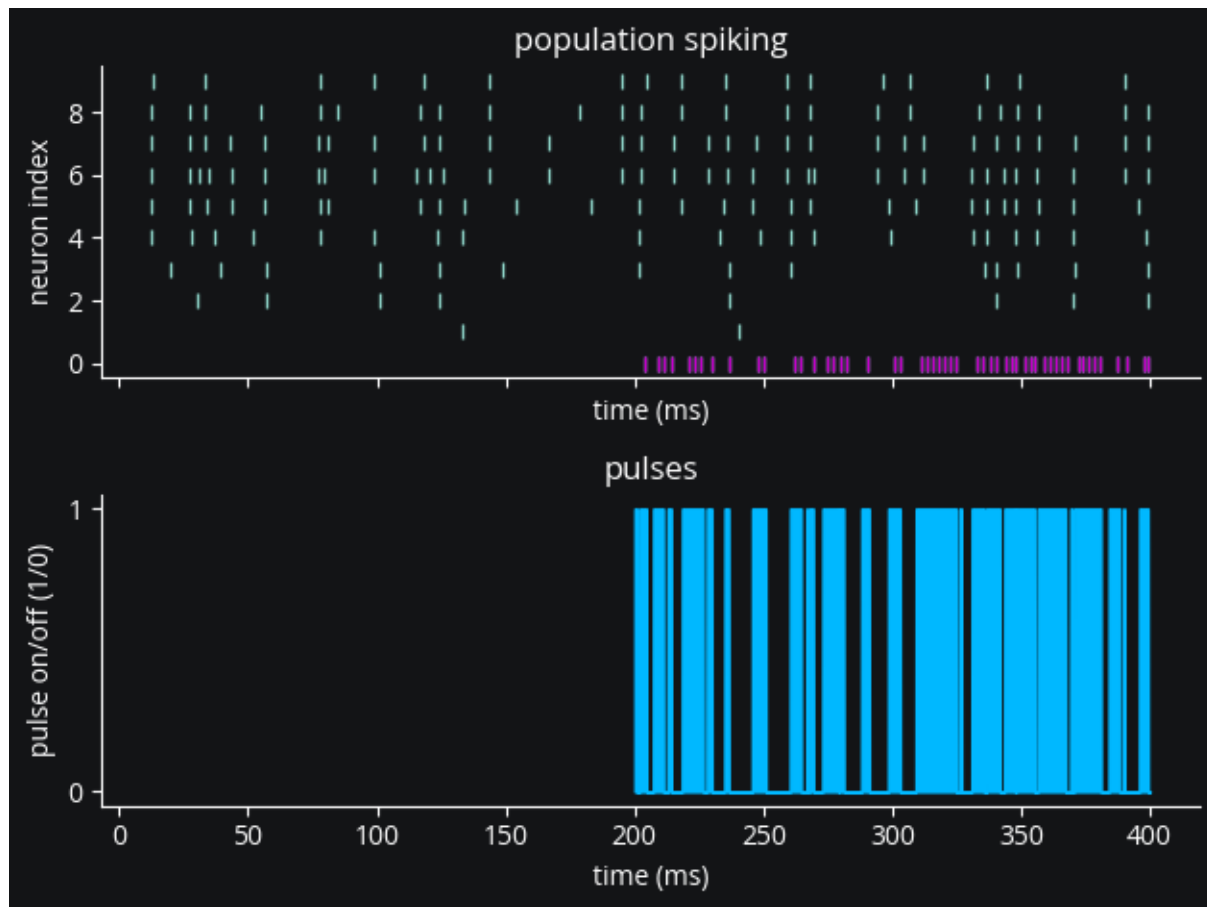
sim.set_io_processor(ReactivePulseIOProcessor(pulse_current=1))
```

```
CLSimulator(io_processor=<__main__.ReactivePulseIOProcessor object at 0x7f57975a1600>,
↳ devices={VoltageRecorder(brian_objects={<StateMonitor, recording ['v'] from
↳ 'neurongroup_subgroup'>}, sim=..., name='rec', voltage_var_name='v', mon=<StateMonitor,
↳ recording ['v'] from 'neurongroup_subgroup'>), StateVariableSetter(brian_
↳ objects=set(), sim=..., name='stim', value=0, default_value=0, save_history=True,
↳ variable_to_ctrl='I', unit=namp, neuron_groups=[<Subgroup 'neurongroup_subgroup_1' of
↳ 'neurongroup' from 0 to 1>])})
```

And run the simulation:

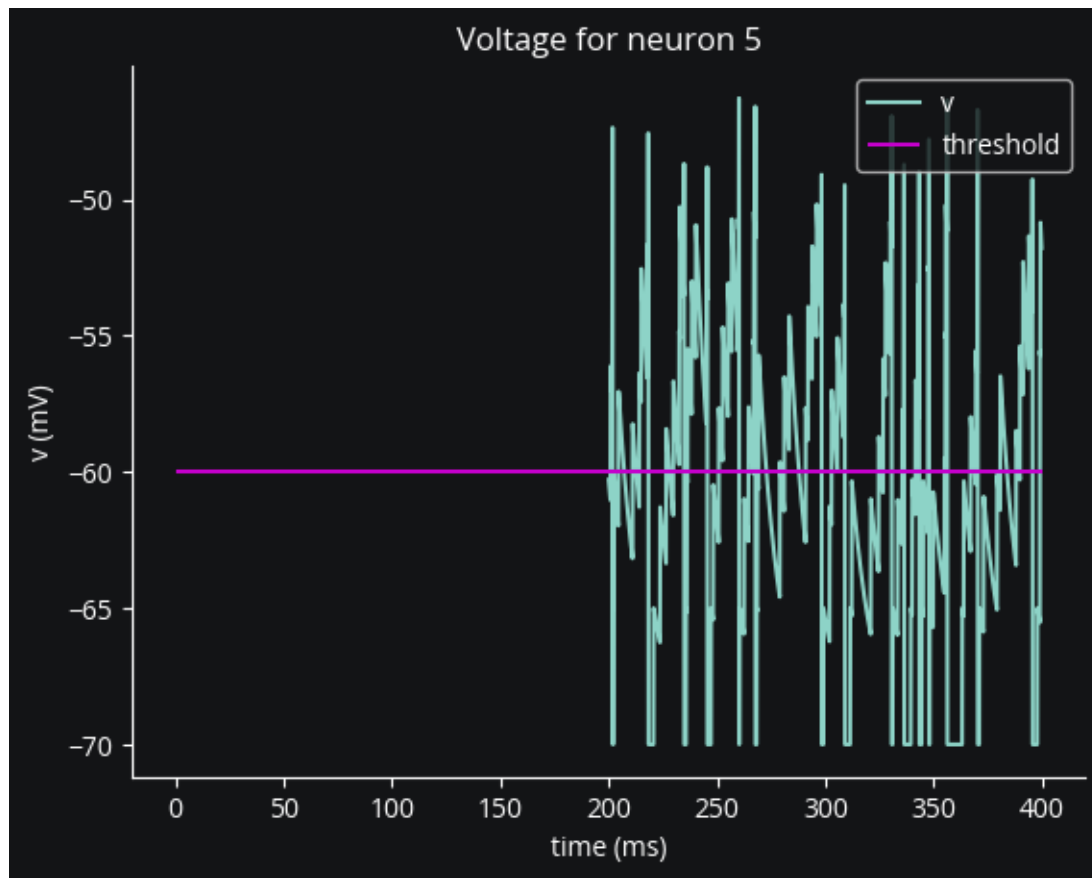
```
sim.run(200*ms)
```

```
fig, (ax1, ax2) = plt.subplots(2, 1, sharex=True)
ax1.plot(pop_mon.t / ms, pop_mon.i[:, "|"])
ax1.plot(
    pop_mon.t[pop_mon.i == i_ctrl] / ms,
    pop_mon.i[pop_mon.i == i_ctrl],
    "|",
    c="#C500CC",
)
ax1.set(title="population spiking", ylabel="neuron index", xlabel="time (ms)")
ax2.fill_between(
    v_rec.mon.t / ms, (v_rec.mon.v.T < trigger_threshold)[:], 0, color=(0.0, 0.72, 1.0)
)
ax2.set(title="pulses", xlabel="time (ms)", ylabel="pulse on/off (1/0)", yticks=[0, 1])
plt.tight_layout()
```



Yes, we see the IO processor triggering pulses as expected. And here's a plot of neuron 5's voltage to confirm that those pulses are indeed where we expect them to be, whenever the voltage is below -60 mV.

```
fig, ax = plt.subplots()
ax.set(title=f"Voltage for neuron {i_rec}", ylabel="v (mV)", xlabel='time (ms)')
ax.plot(v_rec.mon.t/ms, v_rec.mon.v.T / mV);
ax.hlines(-60, 0, 400, color='#c500cc');
ax.legend(['v', 'threshold'], loc='upper right');
```



## Conclusion

In this tutorial we've seen the basics of configuring an `IOProcessor` to implement a closed-loop intervention on a Brian network simulation.

### 6.2.5 PI control

In this tutorial we'll introduce

1. PI control, a commonly used model-free control method,
2. the concept of decomposing the `IOProcessor`'s computation into `ProcessingBlocks`, and
3. modeling computation delays on those blocks to reflect hardware and algorithmic speed limitations present in a real experiment.

Preamble:

```
from brian2 import *
import matplotlib.pyplot as plt
from cleo import *

utilities.style_plots_for_docs()

np.random.seed(7000)
```

(continues on next page)

(continued from previous page)

```
# the default cython compilation target isn't worth it for
# this trivial example
prefs.codegen.target = "numpy"
```

## Create the Brian network

We'll create a population of 10 LIF neurons mainly driven by feedforward input but with some recurrent connections as well.

```
n = 10
population = NeuronGroup(n, '''
    dv/dt = (-v - 70*mV + Rm*I) / tau : volt
    tau: second
    Rm: ohm
    I: amp''',
    threshold='v>-50*mV',
    reset='v=-70*mV'
)
population.tau = 10*ms
population.Rm = 100*Mohm
population.I = 0*mA
population.v = -70*mV

input_group = PoissonGroup(n, np.linspace(20, 200, n)*Hz)

S = Synapses(input_group, population, on_pre='v+=5*mV')
S.connect(condition=f'abs(i-j)<={3}')
S2 = Synapses(population, population, on_pre='v+=2*mV')
S2.connect(p=0.2)

pop_mon = SpikeMonitor(population)

net = Network(population, input_group, S, S2, pop_mon)
population.equations
```

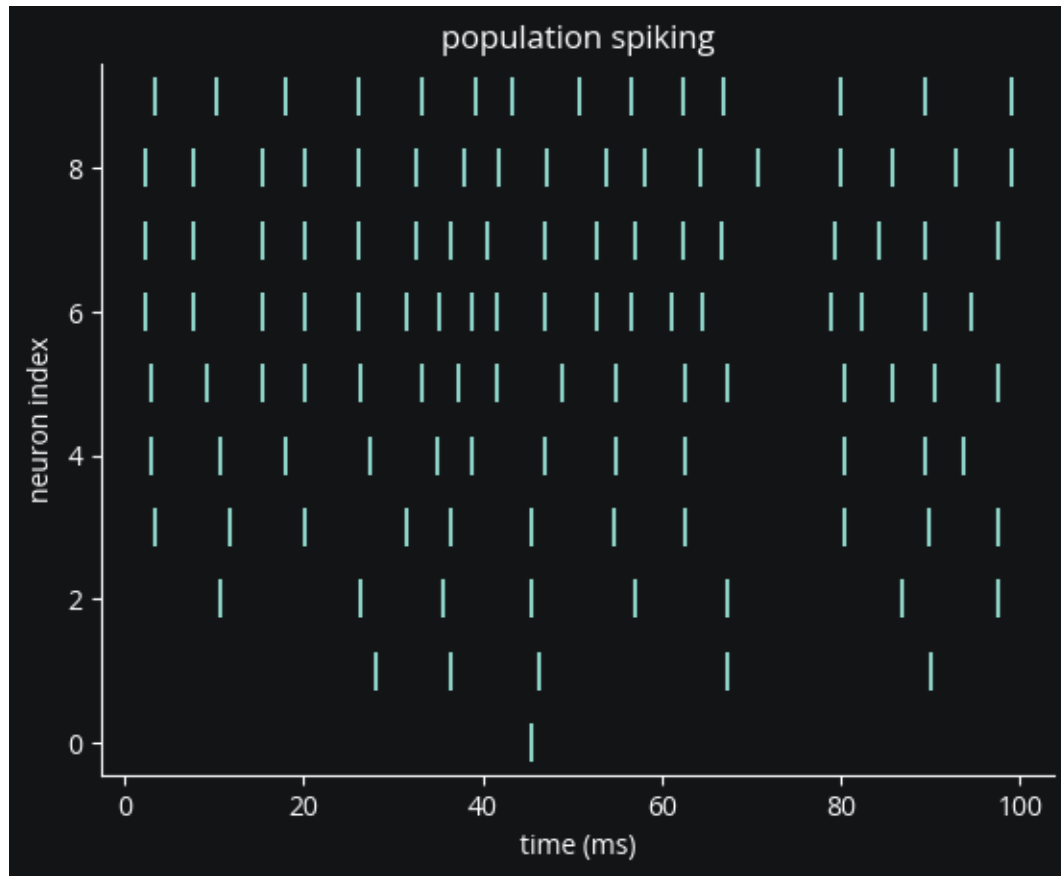
$$\frac{dv}{dt} = \frac{IRm - 70mV - v}{\tau} \quad \begin{array}{l} \text{(unit of } v: \text{ V)} \\ \text{(unit: A)} \\ \text{(unit: ohm)} \\ \text{(unit: s)} \end{array}$$

### Run simulation without control:

```
net.run(100*ms)
```

```
INFO      No numerical integration method specified for group 'neurongroup', using
↳method 'exact' (took 0.06s). [brian2.stateupdaters.base.method_choice]
```

```
fig, ax = plt.subplots()
ax.scatter(pop_mon.t / ms, pop_mon.i, marker='|', s=200);
ax.set(title='population spiking', ylabel='neuron index', xlabel='time (ms)');
```



### Constructing a closed-loop simulation

We will use the popular model-free PI control to control a single neuron's firing rate. PI stands for proportional-integral, referring to a feedback gain *proportional* to the instantaneous error as well as the *integrated* error over time.

First we construct a CLSimulator from the network:

```
from cleo import CLSimulator
sim = CLSimulator(net)
```

Then, to control neuron  $i$ , we need to:

1. capture spiking using a GroundTruthSpikeRecorder

```

from cleo.recorders import GroundTruthSpikeRecorder
i = 0 # neuron to control
rec = GroundTruthSpikeRecorder(name='spike_rec')
sim.inject(rec, population[i])

```

```

CLSimulator(io_processor=None, devices={GroundTruthSpikeRecorder(brian_objects={
    <SpikeMonitor, recording from 'spikemonitor_1'>}, sim=..., name='spike_rec', _mon=
    <SpikeMonitor, recording from 'spikemonitor_1'>, _num_spikes_seen=0, neuron_group=
    <Subgroup 'neurongroup_subgroup' of 'neurongroup' from 0 to 1>))})

```

2. define the firing rate trajectory we want our target neuron to follow

```

# the target firing rate trajectory, as a function of time
def target_Hz(t_ms):
    if t_ms < 250: # constant target at first
        return 400
    else: # sinusoidal afterwards
        a = 200
        t_s = t_ms / 1000
        return a + a * np.sin(2 * np.pi * 20 * t_s)

```

2. estimating its firing rate from incoming spikes using a FiringRateEstimator
3. compute the stimulus intensity with a PIController
4. output that value for a StateVariableSetter stimulator to use

Here we initialize blocks when the IOProcessor is created and define how to process network output and set the control signal in the process function.

```

from cleo.ioproc import (
    LatencyIOProcessor,
    FiringRateEstimator,
    ConstantDelay,
    PIController,
)

class PIRateIOProcessor(LatencyIOProcessor):
    delta = 1 # ms

    def __init__(self):
        super().__init__(sample_period_ms=self.delta, processing="parallel")
        self.rate_estimator = FiringRateEstimator(
            tau_ms=15,
            sample_period_ms=self.delta,
            delay=ConstantDelay(4.1), # latency in ms
            save_history=True, # lets us plot later
        )

        # using hand-tuned gains that seem reasonable
        self.pi_controller = PIController(
            target_Hz,
            Kp=0.005,
            Ki=0.04,

```

(continues on next page)

(continued from previous page)

```

        sample_period_ms=self.delta,
        delay=ConstantDelay(2.87), # latency in ms
        save_history=True, # lets us plot later
    )

    def process(self, state_dict, sample_time_ms):
        spikes = state_dict["spike_rec"]
        # feed output and out_time through each block
        out, time_ms = self.rate_estimator.process(
            spikes, sample_time_ms, sample_time_ms=sample_time_ms
        )
        out, time_ms = self.pi_controller.process(
            out, time_ms, sample_time_ms=sample_time_ms
        )
        # this dictionary output format allows for the flexibility
        # of controlling multiple stimulators
        if out < 0: # limit to positive current
            out = 0
        out_dict = {"I_stim": out}
        # time_ms at the end reflects the delays added by each block
        return out_dict, time_ms

io_processor = PIRateIOProcessor()
sim.set_io_processor(io_processor)

```

```

CLSimulator(io_processor=<__main__.PIRateIOProcessor object at 0x7f6be05fdb70>, devices=
→ {GroundTruthSpikeRecorder(brian_objects={<SpikeMonitor, recording from 'spikemonitor_1'
→ '>}, sim=..., name='spike_rec', _mon=<SpikeMonitor, recording from 'spikemonitor_1'>, _
→ num_spikes_seen=0, neuron_group=<Subgroup 'neurongroup_subgroup' of 'neurongroup' from
→ 0 to 1>}))

```

Note that we can set delays for individual ProcessingBlocks in the IO processor to better approximate the experiment. We use simple constant delays here, but a GaussianDelay class is also available and others could be easily implemented.

Now we inject the stimulator:

```

from cleo.stimulators import StateVariableSetter
sim.inject(
    StateVariableSetter(
        name='I_stim', variable_to_ctrl='I', unit=nA),
    population[i]
)

```

```

CLSimulator(io_processor=<__main__.PIRateIOProcessor object at 0x7f6be05fdb70>, devices=
→ {StateVariableSetter(brian_objects=set(), sim=..., name='I_stim', value=0, default_
→ value=0, save_history=True, variable_to_ctrl='I', unit=namp, neuron_groups=[<Subgroup
→ 'neurongroup_subgroup_1' of 'neurongroup' from 0 to 1>]),
→ GroundTruthSpikeRecorder(brian_objects={<SpikeMonitor, recording from 'spikemonitor_1'>
→ }, sim=..., name='spike_rec', _mon=<SpikeMonitor, recording from 'spikemonitor_1'>, _
→ num_spikes_seen=0, neuron_group=<Subgroup 'neurongroup_subgroup' of 'neurongroup' from
→ 0 to 1>}))

```



## Run the simulation

```
sim.run(300*ms)
```

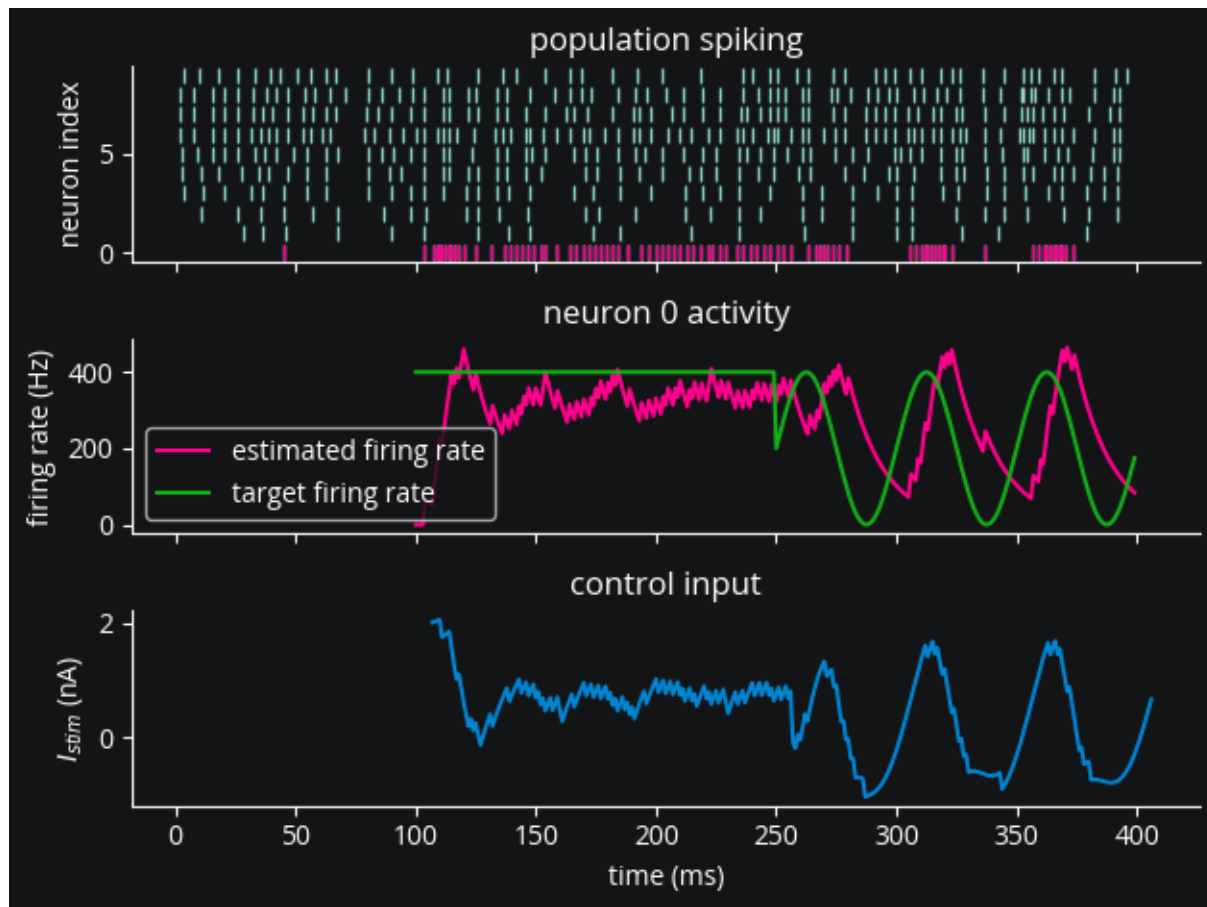
```
fig, (ax1, ax2, ax3) = plt.subplots(3, 1, sharex=True);
ax1.plot(pop_mon.t / ms, pop_mon.i[:], '|');
ax1.plot(pop_mon.t[pop_mon.i == i]/ms, pop_mon.i[pop_mon.i==i], '|', c='xkcd:hot pink')
ax1.set(title='population spiking', ylabel='neuron index')

ax2.plot(io_processor.rate_estimator.t_in_ms, io_processor.rate_estimator.values, c=
↳ 'xkcd:hot pink');
ax2.plot(io_processor.rate_estimator.t_in_ms, [target_Hz(t) for t in io_processor.rate_
↳ estimator.t_in_ms],\
        c='xkcd:green');
ax2.set(ylabel='firing rate (Hz)', title=f'neuron {i} activity');
ax2.legend(['estimated firing rate', 'target firing rate']);

ax3.plot(io_processor.pi_controller.t_out_ms, io_processor.pi_controller.values, c=
↳ 'xkcd:cerulean')
ax3.set(title='control input', ylabel='$I_{stim}$ (nA)', xlabel='time (ms)')

fig.tight_layout()
fig.show()
```

```
WARNING /tmp/ipykernel_28526/1609733203.py:16: UserWarning: Matplotlib is currently_
↳ using module://matplotlib_inline.backend_inline, which is a non-GUI backend, so cannot_
↳ show the figure.
    fig.show()
[py.warnings]
```



Note the lag in keeping up with the target firing rate, which can be directly attributed to the  $\sim 7$  ms delay we coded in to the IO processor.

## Conclusion

In this tutorial, we've learned how to

- use PI control to interact with a Brian simulation,
- decompose processing steps into blocks, and
- assign delays to processing blocks to model real-life latency.

## 6.2.6 LQR optimal control using `ldsctrllest`

This tutorial will be more comprehensive than the others, bringing together all of `cleo`'s main capabilities—electrode recording, optogenetics, and latency modeling—as well as introducing more sophisticated model-based feedback control. To achieve the latter, we will use the `ldsctrllest` Python bindings to the `ldsCtrlEst` C++ library.

Preamble:

```
from brian2 import *
import matplotlib.pyplot as plt
import cleo
```

(continues on next page)

(continued from previous page)

```
cleo.utilities.style_plots_for_docs()

# numpy faster than cython for lightweight example
prefs.codegen.target = 'numpy'
np.random.seed(1856)
```

## Network setup

As in the optogenetics tutorial, we'll use a trivial network of a small neuron group biased by Poisson input spikes. We'll use the exponential integrate-and-fire neuron model, which maintains simplicity while modeling an upward membrane potential swing when spiking.

```
n = 2
ng = NeuronGroup(
    n,
    """
    dv/dt = (-(v - E_L) + Delta_T*exp((v-theta)/Delta_T) + Rm*I) / tau_m : volt
    I : amp
    """,
    threshold="v>30*mV",
    reset="v=-55*mV",
    namespace={
        "tau_m": 20 * ms,
        "Rm": 500 * Mohm,
        "theta": -50 * mV,
        "Delta_T": 2 * mV,
        "E_L": -70 * mV,
    },
)
ng.v = -70 * mV

input_group = PoissonInput(ng, "v", 10, 100 * Hz, 2.5 * mV)

net = Network(ng, input_group)
```

## Coordinates, stimulation, and recording

Here we assign coordinates to the neurons and configure the optogenetic intervention and recording setup:

```
from cleo.coords import assign_coords_rand_rect_prism
from cleo.opto import *
from cleo.light import Light, fiber473nm
from cleo.ephys import Probe, SortedSpiking

hor_lim = 0.05
assign_coords_rand_rect_prism(
    ng, xlim=(-hor_lim, hor_lim), ylim=(-hor_lim, hor_lim), zlim=(0.4, 0.6)
)

fiber = Light(
```

(continues on next page)

(continued from previous page)

```
name="fiber",
light_model=fiber473nm(),
coords=(0, 0, 0.3) * mm,
)

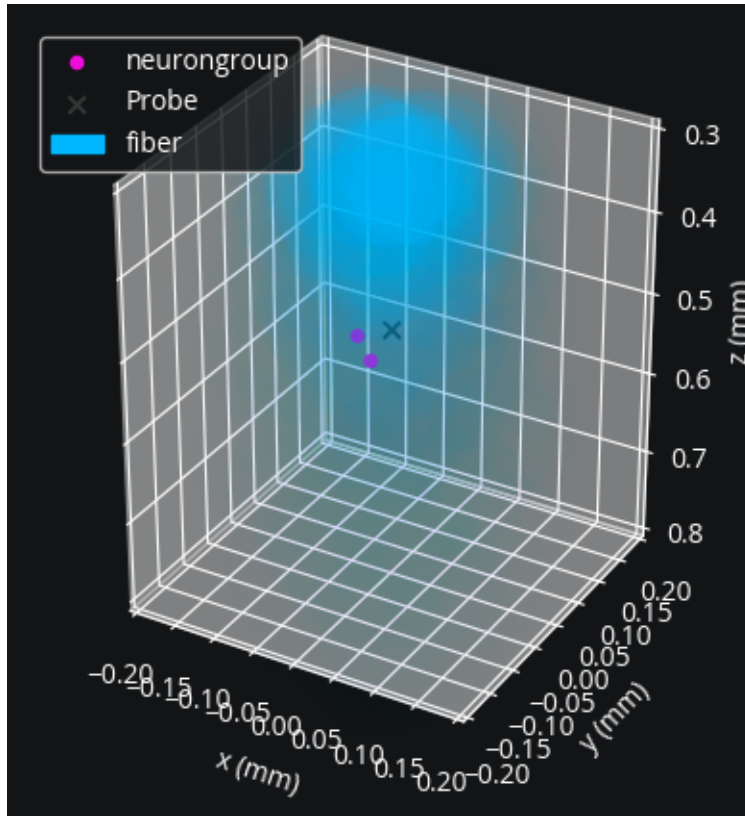
opsin = chr2_4s()

spikes = SortedSpiking(
    name="spikes",
    r_perfect_detection=40 * umeter,
    r_half_detection=80 * umeter,
)

probe = Probe(
    coords=[0, 0, 0.5] * mm,
    signals=[spikes],
    save_history=True,
)

cleo.viz.plot(
    ng,
    colors=["xkcd:fuchsia"],
    xlim=(-0.2, 0.2),
    ylim=(-0.2, 0.2),
    zlim=(0.3, 0.8),
    devices=[probe, fiber],
    scatterargs={"alpha": 1},
    axis_scale_unit=mm,
)
```

```
(<Figure size 640x480 with 1 Axes>,
 <Axes3D: xlabel='x (mm)', ylabel='y (mm)', zlabel='z (mm)'>)
```



Looks right. Let's set up the simulation and inject the devices:

```
sim = cleo.CLSimulator(net)
sim.inject(fiber, ng)
sim.inject(opsin, ng, Iopto_var_name='I')
sim.inject(probe, ng)
```

```
CLSimulator(io_processor=None, devices={Probe(sim=..., name='Probe', save_history=True,
→ signals=[SortedSpiking(name='spikes', brian_objects={<SpikeMonitor, recording from
→ 'spikemonitor'>}, probe=..., r_perfect_detection=40. * umetre, r_half_detection=80. *
→ umetre, cutoff_probability=0.01)], probe=NOTHING), FourStateOpsin(sim=..., name='ChR2',
→ save_history=True, on_pre='', spectrum=[(400, 0.34), (422, 0.65), (460, 0.96), (470,
→ 1), (473, 1), (500, 0.57), (520, 0.22), (540, 0.06), (560, 0.01)], required_vars=[(
→ 'Iopto', amp), ('v', volt)], g0=114. * nsiemens, gamma=0.00742, phim=2.33e+23 *
→ (second ** -1) / (meter ** 2), k1=4.15 * khertz, k2=0.868 * khertz, p=0.833, Gf0=37.3
→ * hertz, kf=58.1 * hertz, Gb0=16.1 * hertz, kb=63. * hertz, q=1.94, Gd1=105. * hertz,
→ Gd2=13.8 * hertz, Gr0=0.33 * hertz, E=0. * volt, v0=43. * mvolt, v1=17.1 * mvolt,
→ model="\n          dC1/dt = Gd1*O1 + Gr0*C2 - Ga1*C1 : 1 (clock-driven)\n          dO1/dt
→ = Ga1*C1 + Gb*O2 - (Gd1+Gf)*O1 : 1 (clock-driven)\n          dO2/dt = Ga2*C2 + Gf*O1 -
→ (Gd2+Gb)*O2 : 1 (clock-driven)\n          C2 = 1 - C1 - O1 - O2 : 1\n\n          Theta =
→ int(phi_pre > 0*phi_pre) : 1\n          Hp = Theta * phi_pre**p/(phi_pre**p + phim**p) :
→ 1\n          Ga1 = k1*Hp : hertz\n          Ga2 = k2*Hp : hertz\n          Hq = Theta * phi_
→ pre**q/(phi_pre**q + phim**q) : 1\n          Gf = kf*Hq + Gf0 : hertz\n          Gb =
→ kb*Hq + Gb0 : hertz\n\n          fphi = O1 + gamma*O2 : 1\n          # v1/v0 when v-E == 0
→ via l'Hopital's rule\n          fv = f_unless_x0(\n          (1 - exp(-(V_VAR_NAME_
→ post-E)/v0)) / ((V_VAR_NAME_post-E)/v1),\n          V_VAR_NAME_post - E,\n
→ v1/v0\n          ) : 1\n\n          IOPTO_VAR_NAME_post = -g0*fphi*fV*(V_VAR_NAME_post-
```

(continues on next page)

(continued from previous page)

```

→E)*rho_rel : ampere (summed)\n          rho_rel : 1", extra_namespace={'f_unless_x0':
→<brian2.core.functions.Function object at 0x7f5971fba6e0>}), Light(sim=..., name='fiber
→', save_history=True, value=array([0.]), light_model=OpticFiber(R0=100. * umetre,
→NAfib=0.37, K=125. * metre ** -1, S=7370. * metre ** -1, ntis=1.36), wavelength=0.473,
→* umetre, direction=array([0., 0., 1.]), max_Irr0_mW_per_mm2=None, max_Irr0_mW_per_mm2_
→viz=None)})

```

## Prepare controller

Our goal will be to control two neuron's firing rates simultaneously. To do this, we will use the LQR technique explained in Bolus et al., 2021 ("State-space optimal feedback control of optogenetically driven neural activity").

## Fit model

Our controller needs a model of the system's dynamics, which we can obtain by fitting to training data. We will generate training data using Gaussian random walk inputs. `ldsCtrlEst` is designed for data coming from an experiment, organized into trials, so we will run the simulation repeatedly, resetting after each run. Here  $u$  represents the input and  $z$  the spike output.

We will intentionally use very little training data so the importance of adaptive control will become apparent later on.

```

n_trials = 5
n_samp = 100
u = []
z = []
n_u = 1 # 1-dimensional input (just one optogenetic actuator)
n_z = 2 # we'll be controlling two neurons
for trial in range(n_trials):
    # one-sided normally distributed training data, stdev of 10 mW/mm2
    u_trial = 10*np.abs(np.random.randn(n_u, n_samp))
    u.append(u_trial)
    z.append(np.zeros((n_z, n_samp)))

```

The IO processor is simple enough here that we won't bother separating steps using `:class:~cleo.ioproc.ProcessingBlock` objects, which is recommended for more complex scenarios where modularity is more important.

```

from cleo.ioproc import LatencyIOProcessor

class TrainingStimIOP(LatencyIOProcessor):
    i_samp = 0
    i_trial = 0

    # here we just feed in the training inputs and record the outputs
    def process(self, state_dict, sample_time_ms):
        i, t, z_t = state_dict['Probe']['spikes']
        z[self.i_trial][:, self.i_samp] = z_t[:n_z] # just first two neurons
        out = {'fiber': u[self.i_trial][:, self.i_samp]}
        self.i_samp += 1
        return out, sample_time_ms

training_stim_iop = TrainingStimIOP(sample_period_ms=1)

```

(continues on next page)

(continued from previous page)

```
sim.set_io_processor(training_stim_iop)

for i_trial in range(n_trials):
    training_stim_iop.i_trial = i_trial
    training_stim_iop.i_samp = 0
    sim.run(n_samp*ms)
    sim.reset()
```

```
INFO      No numerical integration method specified for group 'neurongroup', using
↳method 'euler' (took 0.02s, trying other methods took 0.07s). [brian2.stateupdaters.
↳base.method_choice]
```

```
INFO      No numerical integration method specified for group 'syn_ChR2_neurongroup',
↳using method 'euler' (took 0.02s, trying other methods took 0.09s). [brian2.
↳stateupdaters.base.method_choice]
```

Now we have  $u$  and  $z$  in the form we need for `ldsctrlest`'s fitting functions:  $n\_trial$ -length lists of  $n$  by  $n\_samp$  arrays. We will now fit Gaussian linear dynamical systems using the SSID algorithm. See [the documentation](#) for more detailed explanations.

```
import ldsctrlest as lds
import ldsctrlest.gaussian as gls
n_x_fit = 2 # latent dimensionality of system
n_h = 50 # size of block Hankel data matrix
dt = 0.001 # timestep (in seconds)
u_train = lds.UniformMatrixList(u, free_dim=2)
z_train = lds.UniformMatrixList(z, free_dim=2)
ssid = gls.FitSSID(n_x_fit, n_h, dt, u_train, z_train)
fit, sing_vals = ssid.Run(lds.SSIDWt.kMOESP)
```

## Design controller

### LQR optimal control

We now use the fit parameters to create the controller system and set additional parameters. The feedback gain,  $K_c$ , is especially important, determining how the controller responds to the current “error”—the difference between where the system is (estimated to be) now and where we want it to be. The field of optimal control deals with how to design the controller so as to minimize a cost function reflecting what we care about.

With a linear system (obtained from the fitting procedure above) and quadratic per-timestep cost function  $L$  penalizing distance from the reference  $x^*$  and the input  $u$

$$L = \frac{1}{2} (x - x^*)^T Q (x - x^*) + \frac{1}{2} u^T R u$$

we can use the closed-form optimal solution called the Linear Quadratic Regulator (LQR).

$$K = (R + B^T P B)^{-1} (B^T P A) \quad u = -Kx$$

The  $P$  matrix is obtained by numerically solving the discrete algebraic Riccati equation:

$$P = A^T \{ P A - (A^T P B) (R + B^T P B)^{-1} (B^T P A) + Q \}$$

```

fit_sys = glds.System(fit)
# upper and lower bounds on control signal (optic fiber light intensity)
u_lb = 0 # mW/mm2
u_ub = 30 # mW/mm2
controller = glds.Controller(fit_sys, u_lb, u_ub)
# careful not to use this anymore since controller made a copy
del fit_sys

from scipy.linalg import solve_discrete_are
# cost matrices
# Q reflects how much we care about state error
# we use C'C since we really care about output error, not latent state
Q_cost = controller.sys.C.T @ controller.sys.C
R_cost = 1e-4 * np.eye(n_u) # reflects how much we care about minimizing the stimulus
A, B = controller.sys.A, controller.sys.B
P = solve_discrete_are(A, B, Q_cost, R_cost)
controller.Kc = np.linalg.inv(R_cost + B.T @ P @ B) @ (B.T @ P @ A)
controller.Print()

```

```

***** SYSTEM *****
x:
    0
    0

P:
    1.0000e-06      0
      0      1.0000e-06

A:
    0.5877    0.7122
   -0.7679    0.4800

B:
   -0.0152
   -0.0124

g:
    1.0000

m:
    0
    0

Q:
    0.0783    0.0510
    0.0510    0.1465

Q_m:
    1.0000e-06      0
      0      1.0000e-06

d:

```

(continues on next page)



(continued from previous page)

```

    0
    0

C:
-0.1074    0.0030
-0.0810    0.1761

y:
    0
    0

R:
    0.0943    0.0164
    0.0164    0.1574

g_design :    1.0000

u_lb : 0
u_ub : 30

```

We now configure the IOProcessor to use our controller:

```

class CtrlLoop(LatencyIOProcessor):
    def __init__(self, samp_period_ms, controller, y_ref: callable):
        super().__init__(samp_period_ms)
        self.controller = controller
        self.sys = controller.sys
        self.y_ref = y_ref
        self.do_control = False # allows us to turn on and off control

        # for post hoc visualization/analysis:
        self.u = np.empty((n_u, 0))
        self.x_hat = np.empty((n_x_fit, 0))
        self.y_hat = np.empty((n_z, 0))
        self.z = np.empty((n_z, 0))

    def process(self, state_dict, sample_time_ms):
        i, t, z_t = state_dict["Probe"]["spikes"]
        z_t = z_t[:n_z].reshape((-1, 1)) # just first n_z neurons
        self.controller.y_ref = self.y_ref(sample_time_ms)

        u_t = self.controller.ControlOutputReference(z_t, do_control=self.do_control)
        out = {fiber.name: u_t.squeeze()}

        # record variables from this timestep
        self.u = np.hstack([self.u, u_t])
        self.y_hat = np.hstack([self.y_hat, self.sys.y])
        self.x_hat = np.hstack([self.x_hat, self.sys.x])
        self.z = np.hstack([self.z, z_t])

        return out, sample_time_ms + 3 # 3 ms delay

```

(continues on next page)

(continued from previous page)

```

y_ref = 200 * dt # target rate in Hz
ctrl_loop = CtrlLoop(
    samp_period_ms=1, controller=controller, y_ref=lambda t: np.ones((n_z, 1)) * y_ref
)

```

## Run the experiment

We'll now run the simulation with and without control to compare.

```

sim.set_io_processor(ctrl_loop)
T0 = 100
sim.run(T0*ms)

ctrl_loop.do_control = True
T1 = 350
sim.run(T1*ms)

```

```

WARNING      'dt' is an internal variable of group 'syn_ChR2_neurongroup', but also exists
↳ in the run namespace with the value 0.001. The internal variable will be used. [brian2.
↳ groups.group.Group.resolve.resolution_conflict]

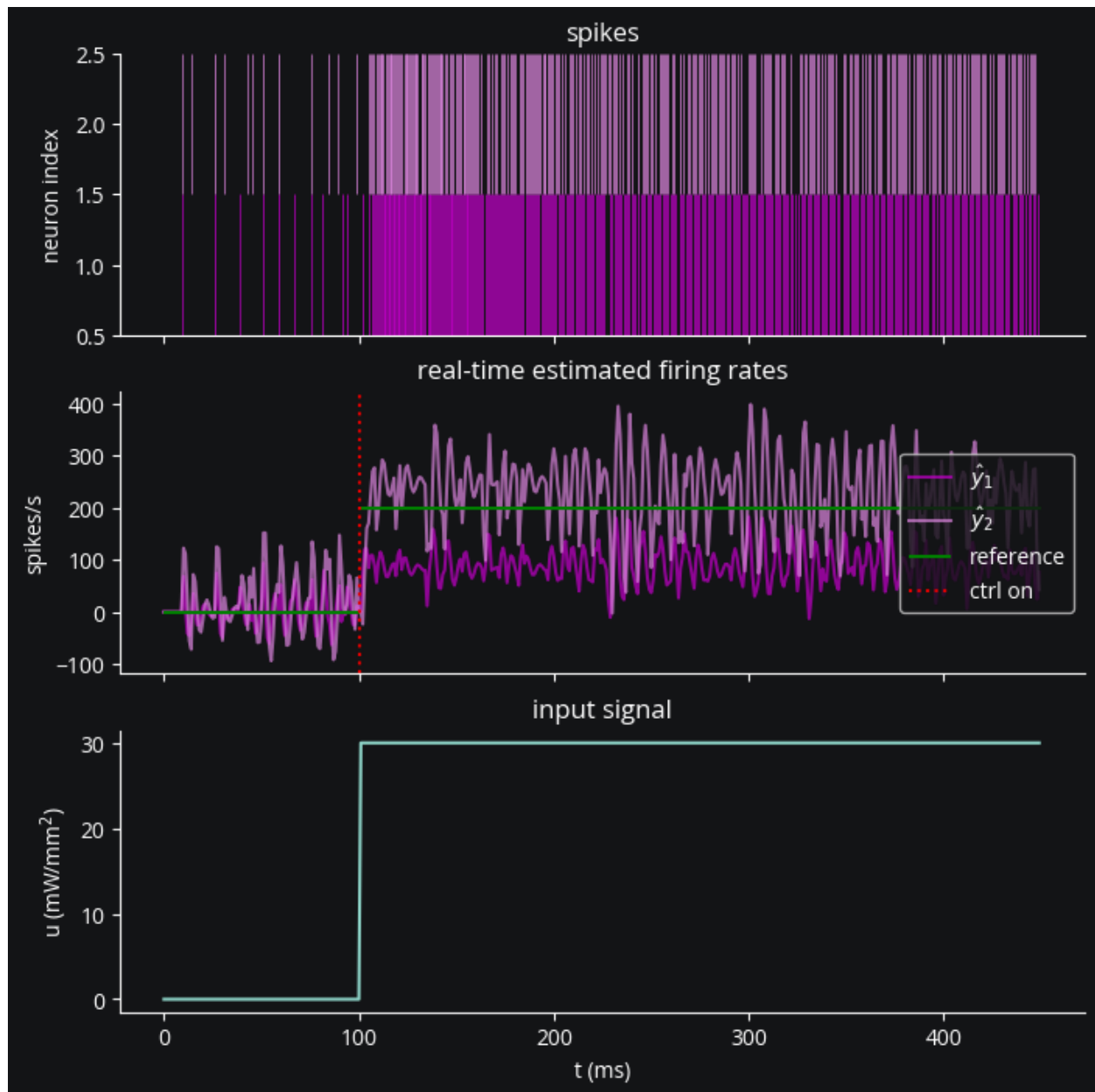
```

Now we plot the results to see how well the controller was able to match the desired firing rate:

```

fig, (ax1, ax2, ax3) = plt.subplots(3, 1, sharex=True, figsize=(8,8))
c1 = "#C500CC"
c2 = "#df87e1"
spikes1 = spikes.t_ms[spikes.i == 0]
spikes2 = spikes.t_ms[spikes.i == 1]
ax1.eventplot([spikes1, spikes2], lineoffsets=[1, 2], colors=[c1, c2], lw=0.5)
ax1.set(ylabel='neuron index', ylim=(0.5, 2.5), title='spikes')
ax2.set(ylabel='spikes/s', title='real-time estimated firing rates')
ax2.plot(ctrl_loop.y_hat[0]/dt, c=c1, alpha=0.7, label='$\hat{y}_1$')
ax2.plot(ctrl_loop.y_hat[1]/dt, c=c2, alpha=0.7, label='$\hat{y}_2$')
ax2.hlines(y_ref/dt, 100, T0+T1, color='green', label='reference')
ax2.hlines(0, 0, 100, color='green')
ax2.axvline(T0, c='xkcd:red', linestyle=':', label='ctrl on')
ax2.legend(loc="right")
ax3.plot(range(T0+T1), ctrl_loop.u.T)
ax3.set(xlabel='t (ms)', ylabel='u (mW/mm$^2$)', title='input signal');

```



Looks all right, but in addition to the system's estimated firing rate let's count the spikes over the control period to see how well we hit the target on average:

```
print("Results (spikes/second):")
print('baseline =', np.sum(ctrl_loop.z[:, :T0], axis=1)/(T0/1000))
print('target =', [y_ref*1000, y_ref*1000])
print('lqr achieved =', (np.sum(ctrl_loop.z[:, T0:T0+T1], axis=1)/(T1/1000)).round(1))
```

```
Results (spikes/second):
baseline = [100. 120.]
target = [200.0, 200.0]
lqr achieved = [928.6 748.6]
```

We can see that the system consistently underestimates the true firing rate. And as we could expect, we weren't able

to maintain the target firing rate with both neurons simultaneously since one was exposed to more light than the other. However, the controller was able to achieve something. See the appendix for how we can avoid overshooting with both neurons, which should be avoidable.

## Conclusion

As a recap, in this tutorial we've seen how to:

- inject optogenetic stimulation into an existing Brian network
- inject an electrode into an existing Brian network to record spikes
- generate training data and fit a Gaussian linear dynamical system to the spiking output using `ldsctrllest`
- configure an `ldsctrllest` LQR controller based on that linear system and design optimal gains
- use that controller in running a complete simulated feedback control experiment

## Appendix

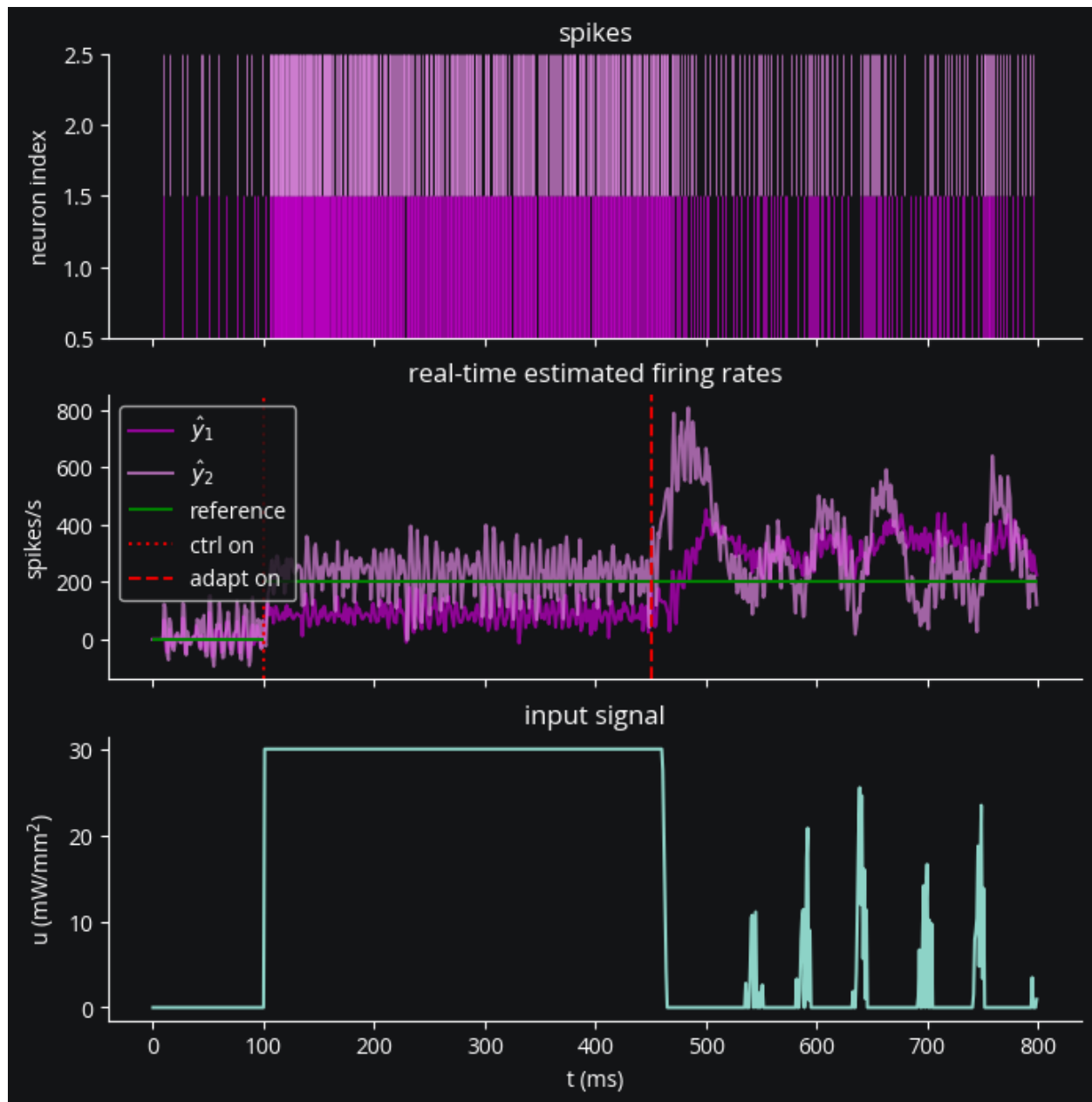
### Adaptive control

`ldsCtrlEst` also provides an *adaptive* variation on LQR, capable of inferring state beyond our static, linear model and thus able to account for unmodeled disturbances and noise. Let's see how it compares:

```
controller.sys.do_adapt_m = True # enable adaptive disturbance estimation
# set covariance for the disturbance state
# larger values mean the system more readily ascribes changes to unmodeled disturbance
controller.sys.Q_m = 1e-2 * np.eye(n_x_fit)
controller.control_type = lds.kControlTypeAdaptM # enable adaptive control
```

```
ctrl_loop.sys.do_adapt_m = True
T2 = 350
sim.run(T2*ms)
T = T0 + T1 + T2
```

```
fig, (ax1, ax2, ax3) = plt.subplots(3, 1, sharex=True, figsize=(8,8))
spikes1 = spikes.t_ms[spikes.i == 0]
spikes2 = spikes.t_ms[spikes.i == 1]
ax1.eventplot([spikes1, spikes2], lineoffsets=[1, 2], colors=[c1, c2], lw=0.5)
ax1.set(ylabel='neuron index', ylim=(.5, 2.5), title='spikes')
ax2.set(ylabel='spikes/s', title='real-time estimated firing rates')
ax2.plot(ctrl_loop.y_hat[0]/dt, c=c1, alpha=0.7, label='$\hat{y}_1$')
ax2.plot(ctrl_loop.y_hat[1]/dt, c=c2, alpha=0.7, label='$\hat{y}_2$')
ax2.hlines(y_ref/dt, 100, T, color='green', label='reference')
ax2.hlines(0, 0, 100, color='green')
ax2.axvline(T0, c='xkcd:red', linestyle=':', label='ctrl on')
ax2.axvline(T0+T1, c='xkcd:red', linestyle='--', label='adapt on')
ax2.legend(loc="upper left")
ax3.plot(range(T), ctrl_loop.u.T)
ax3.set(xlabel='t (ms)', ylabel='u (mW/mm$^2$)', title='input signal');
```



We can see the effect most easily in the input signal, which has much more variation now. Let's confirm that the firing rates were better balanced around the target:

```
print("Results (spikes/second):")
print('baseline =', np.sum(ctrl_loop.z[:, :T0], axis=1)/(T0/1000))
print('target =', [y_ref*1000, y_ref*1000])
print('static achieved =', (np.sum(ctrl_loop.z[:, T0:T0+T1], axis=1)/(T1/1000)).round(1))
print('adaptive achieved =', (np.sum(ctrl_loop.z[:, T0+T1:T], axis=1)/(T2/1000)).
↪round(1))
```

```
Results (spikes/second):
baseline = [100. 120.]
target = [200.0, 200.0]
```

(continues on next page)

(continued from previous page)

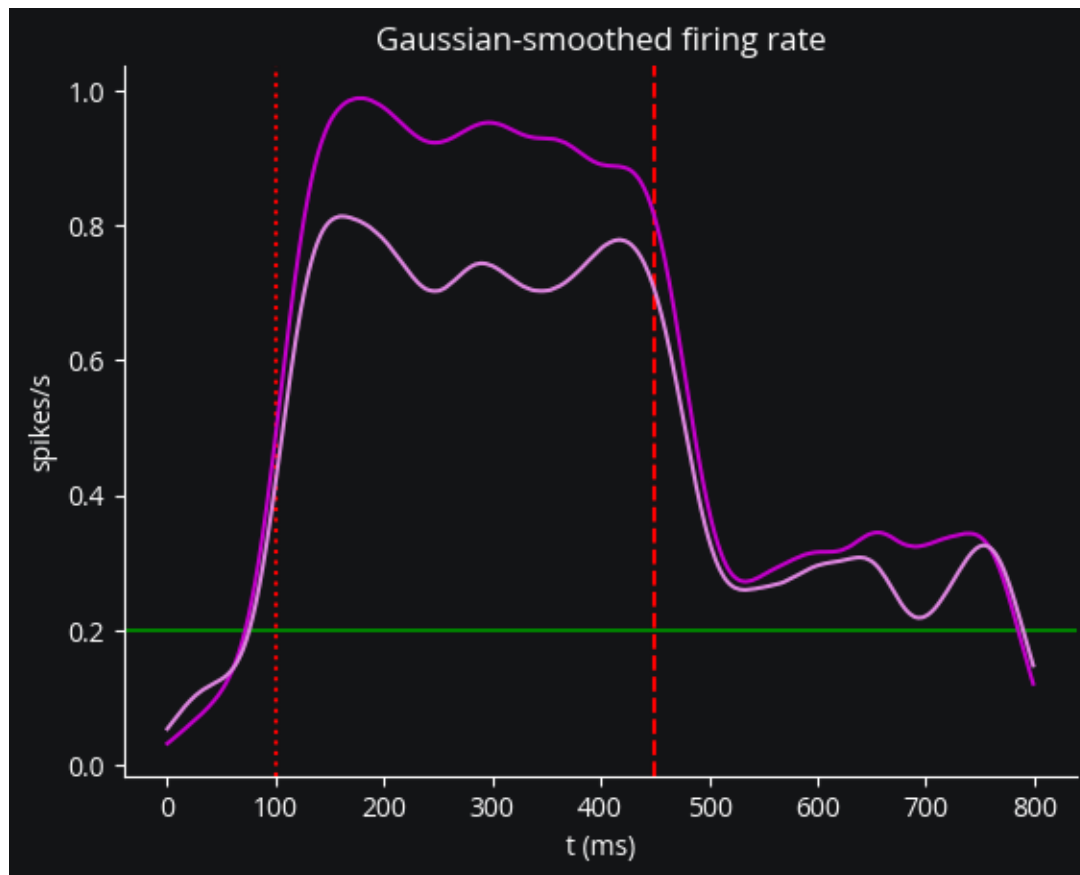
```
static achieved = [928.6 748.6]
adaptive achieved = [354.3 317.1]
```

That looks better. Adaptive control achieves a balance between the two neurons, as we would expect.

### Post-hoc firing rate estimate

To see if the system's online estimation of firing rates is reasonable, we compute a Gaussian-smoothed version with a 25-ms standard deviation:

```
from scipy.stats import norm
kernel = norm.pdf(np.linspace(-75, 75, 151), scale=25) # 25-ms Gaussian window
smoothed1 = np.convolve(ctrl_loop.z[0, :], kernel, mode='same')
smoothed2 = np.convolve(ctrl_loop.z[1, :], kernel, mode='same')
plt.axhline(y_ref, c='g')
plt.axvline(T0, c='r', ls=':')
plt.axvline(T0+T1, c='r', ls='--')
plt.xlabel("t (ms)")
plt.ylabel("spikes/s")
plt.title("Gaussian-smoothed firing rate")
plt.plot(smoothed1, c=c1)
plt.plot(smoothed2, c=c2);
```



## 6.2.7 All-optical control

In this tutorial we'll see how to

- configure and inject a microscope for two-photon imaging and stimulation
- inject a calcium indicator for neurons visible to the microscope
- set up two-photon optogenetics by
  - injecting an opsin with an extended action spectrum and
  - injecting a 2P light source targeting neurons imaged by the microscope

### Preamble

```
import brian2 as b2
from brian2 import np
import cleo
from cleo import opto, imaging, light
import matplotlib.pyplot as plt

# for reproducibility
rng = np.random.default_rng(92)
np.random.seed(92)

cleo.utilities.style_plots_for_docs()
```

### Brian network setup

All we need are LIF neurons that will spike in response to photostimulation for us to see the resulting fluorescence traces.

```
ng = b2.NeuronGroup(
    200,
    """dv/dt = -(v - E_L) + Rm*Iopto) / tau_m : volt
    Iopto : amp""",
    threshold="v > -50*mV",
    reset="v=E_L",
    namespace={
        "tau_m": 20 * b2.ms,
        "Rm": 500 * b2.Mohm,
        "E_L": -70 * b2.mV,
    },
)
ng.v = -70 * b2.mV
cleo.coords.assign_coords_rand_rect_prism(ng, [-0.2, 0.2], [-0.2, 0.2], [0, 0.4])
sim = cleo.CLSimulator(b2.Network(ng))
```

## Microscope configuration

By default a scope selects neurons based on focus depth and assigns them a signal-to-noise ratio (SNR) based on soma size. (Larger cells' SNR decays more slowly with distance from the focal plane.) `focus_depth` and `soma_radius` are taken from scope parameters but can be overridden on injection.

We'll use `GCaMP6f` as our indicator and leave the default parameters, though those can be changed in the function call `gcamp6f()`.

```
scope = imaging.Scope(
    focus_depth=100 * b2.um,
    img_width=500 * b2.um,
    sensor=imaging.gcamp6f(),
)
```

Heterogeneous expression is simulated by providing a `rho_rel_generator` function that takes the number of neurons as an argument and returns a vector of relative expression levels. This is done on scope injection (as opposed to sensor injection afterward) because the scope needs this information to select ROIs with sufficiently high SNR.

```
expr_level_gen = lambda n: rng.lognormal(0, 0.2, n)
sim.inject(scope, ng, rho_rel_generator=expr_level_gen) # uses scope's parameters
# optional overrides (we'll use a small soma to intentionally get few targets)
sim.inject(scope, ng, focus_depth=200 * b2.um, soma_radius=5 * b2.um, rho_rel_
generator=expr_level_gen)
```

```
CLSimulator(io_processor=None, devices={Scope(sim=..., name='Scope', save_history=True,
sensor=GECI(sim=None, name='gcamp6f', save_history=True, model='\n          dCa/dt = -
gamma * (Ca - Ca_rest) / (1 + kappa_S + kappa_B) : mmolar (clock-driven)\n
kappa_B = B_T * K_d / (Ca + K_d)**2 : 1\n\n          CaB_active = Ca_rest + b :
mmolar # add tiny bit to avoid /0\n          db/dt = beta : mmolar (clock-driven)\n
lam = 1/tau_off + 1/tau_on : 1/second\n          kap = 1/tau_off : 1/
second\n          dbeta/dt = (          # should be M/s/s\n
A * (lam - kap) * (Ca - Ca_rest) # M/s/s\n          - (kap + lam) * beta
# M/s/s\n          - kap * lam * b # M/s/s\n          ) : mmolar/second
(clock-driven)\n          \nexc_factor = 1 : 1\n\n          dFF_baseline = 1 / (1
+ (K_d / Ca_rest) ** n_H) : 1\n          dFF = exc_factor * rho_rel * dFF_max * (\n
1 / (1 + (K_d / CaB_active) ** n_H)\n          - dFF_baseline\n
) : 1\n          rho_rel : 1\n          ', on_pre='Ca += dCa_T / (1 + kappa_S
+ kappa_B)', sigma_noise=0.03748181818181818, dFF_1AP=0.097755000000000001, location=
'cytoplasm', cal_model=DynamicCalcium(on_pre='Ca += dCa_T / (1 + kappa_S + kappa_B)',
model='\n          dCa/dt = -gamma * (Ca - Ca_rest) / (1 + kappa_S + kappa_B) :
mmolar (clock-driven)\n          kappa_B = B_T * K_d / (Ca + K_d)**2 : 1', Ca_
rest=50. * nmolar, gamma=292.3 * hertz, B_T=200. * umolar, kappa_S=110, dCa_T=7.6 *
umolar), bind_act_model=DoubExpCalBindingActivation(model='\n          CaB_active =
Ca_rest + b : mmolar # add tiny bit to avoid /0\n          db/dt = beta : mmolar
(clock-driven)\n          lam = 1/tau_off + 1/tau_on : 1/second\n          kap = 1/
tau_off : 1/second\n          dbeta/dt = (          # should be M/s/s\n
A * (lam - kap) * (Ca - Ca_rest) # M/s/s\n          - (kap + lam) *
beta # M/s/s\n          - kap * lam * b # M/s/s\n          ) :
mmolar/second (clock-driven)\n          ', A=7.61251 * khertz, tau_on=1.17164616 *
second, tau_off=10.14020867 * msecond, Ca_rest=50. * nmolar), exc_
model=NULLExcitation(model='exc_factor = 1 : 1'), fluor_model='\n          dFF_
baseline = 1 / (1 + (K_d / Ca_rest) ** n_H) : 1\n          dFF = exc_factor * rho_
rel * dFF_max * (\n          1 / (1 + (K_d / CaB_active) ** n_H)\n
```

(continues on next page)



(continued from previous page)

```

→ - dFF_baseline\n                ) : 1\n                rho_rel : 1\n                ', K_d=290. * \n
→ nmolar, n_H=2.7, dFF_max=25.2), img_width=0.5 * mmetre, focus_depth=100. * umetre, \n
→ location=array([0., 0., 0.]) * metre, direction=array([0., 0., 1.]), soma_radius=10. * \n
→ umetre, snr_cutoff=1))

```

Signal strength (F/F for 1 spike) per ROI is thus defined as `dFF_1AP` \* `rho_rel`.

While Cleo adjusts noise levels according to distance from the focal plane automatically, other adjustments can be made manually by calling `target_neurons_in_plane()` to reflect factors such as cell heterogeneity or indicator expression or increased noise with depth.

Neurons with SNR (defined as `dFF_1AP` / `sigma_noise`) below the scope's `snr_cutoff` parameter are discarded.

```

i_targets, noise_focus_factor, focus_coords = scope.target_neurons_in_plane(ng, focus_
→ depth=300 * b2.um, soma_radius=5 * b2.um)
# scale noise std randomly to simulate biological variability
std_noise = scope.sensor.sigma_noise * noise_focus_factor * rng.normal(1, .2, len(i_
→ targets))
sim.inject(scope, ng, focus_depth=None, i_targets=i_targets, sigma_noise=std_noise, rho_
→ rel_generator=expr_level_gen)

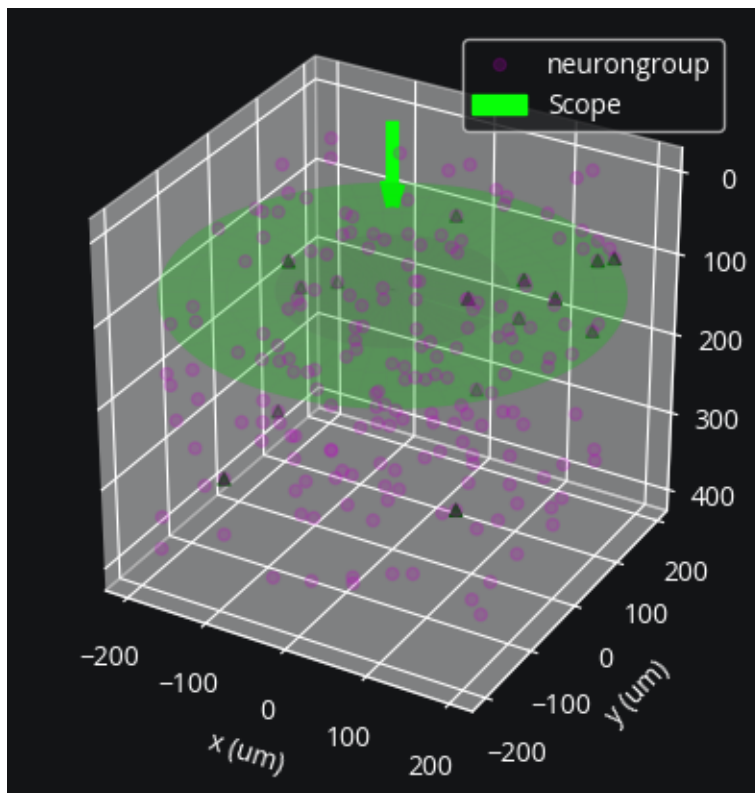
cleo.viz.plot(ng, colors=['#c500cc'], sim=sim)

```

```

(<Figure size 640x480 with 1 Axes>,
 <Axes3D: xlabel='x (um)', ylabel='y (um)', zlabel='z (um)'>)

```



We can see targets off the visualized plane, resulting from our injections at 200 and 300  $\mu\text{m}$  depths, resembling a multi-plane imaging experiment.

When `focus_depth` is set to `None`, corresponding to [sculpted holographic imaging](#), the scope will select all neurons in the volume, or the user can specify a list of neurons to select via `i_targets` on injection.

After all targets are specified, the sensor protein must also be injected. Doing it this way allows for efficiency: the sensor protein is only simulated for the imaged neurons. Expression levels are as sampled previously on scope injection.

```
scope.inject_sensor_for_targets()
```

## 2p stimulation configuration

Future work will address how to better quantify the relationship between 2P and the 1P action spectra, but for now we'll pretend that Vf-Chrimson activation at 1060 nm is 1/100th that of peak 1P activation for the same power density.

```
opsin = opto.vfchrimson_4s()
opsin.spectrum.append((1060, .01))
sim.inject(opsin, ng)
```

```
CLSimulator(io_processor=None, devices={GECI(sim=..., name='gcamp6f', save_history=True,
→model='\n          dCa/dt = -gamma * (Ca - Ca_rest) / (1 + kappa_S + kappa_B) :
→mmolar (clock-driven)\n          kappa_B = B_T * K_d / (Ca + K_d)**2 : 1\n\n
→CaB_active = Ca_rest + b : mmolar # add tiny bit to avoid /0\n          db/dt =
→beta : mmolar (clock-driven)\n          lam = 1/tau_off + 1/tau_on : 1/second\n
→kap = 1/tau_off : 1/second\n          dbeta/dt = ( # should
→be M/s/s\n          A * (lam - kap) * (Ca - Ca_rest) # M/s/s\n
→(kap + lam) * beta # M/s/s\n          - kap * lam * b # M/s/s\n
→) : mmolar/second (clock-driven)\n          \nexc_factor = 1 : 1\n
→dFF_baseline = 1 / (1 + (K_d / Ca_rest) ** n_H) : 1\n          dFF = exc_factor *
→rho_rel * dFF_max * (\n          1 / (1 + (K_d / CaB_active) ** n_H)\n
→- dFF_baseline\n          ) : 1\n          rho_rel : 1\n          ', on_pre=
→'Ca += dCa_T / (1 + kappa_S + kappa_B)', sigma_noise=0.03748181818181818, dFF_1AP=0.
→097755000000000001, location='cytoplasm', cal_model=DynamicCalcium(on_pre='Ca += dCa_T /
→(1 + kappa_S + kappa_B)', model='\n          dCa/dt = -gamma * (Ca - Ca_rest) / (1
→+ kappa_S + kappa_B) : mmolar (clock-driven)\n          kappa_B = B_T * K_d / (Ca +
→K_d)**2 : 1', Ca_rest=50. * nmolar, gamma=292.3 * hertz, B_T=200. * umolar, kappa_
→S=110, dCa_T=7.6 * umolar), bind_act_model=DoubExpCalBindingActivation(model='\n
→CaB_active = Ca_rest + b : mmolar # add tiny bit to avoid /0\n          db/dt
→= beta : mmolar (clock-driven)\n          lam = 1/tau_off + 1/tau_on : 1/second\n
→kap = 1/tau_off : 1/second\n          dbeta/dt = ( #
→should be M/s/s\n          A * (lam - kap) * (Ca - Ca_rest) # M/s/s\n
→- (kap + lam) * beta # M/s/s\n          - kap * lam * b # M/s/s\n
→) : mmolar/second (clock-driven)\n          ', A=7.61251 * khertz, tau_
→on=1.17164616 * second, tau_off=10.14020867 * msecond, Ca_rest=50. * nmolar), exc_
→model=NULLExcitation(model='exc_factor = 1 : 1'), fluor_model='\n          dFF_
→baseline = 1 / (1 + (K_d / Ca_rest) ** n_H) : 1\n          dFF = exc_factor * rho_
→rel * dFF_max * (\n          1 / (1 + (K_d / CaB_active) ** n_H)\n
→- dFF_baseline\n          ) : 1\n          rho_rel : 1\n          ', K_d=290. *
→nmolar, n_H=2.7, dFF_max=25.2), Scope(sim=..., name='Scope', save_history=True,
→sensor=GECI(sim=..., name='gcamp6f', save_history=True, model='\n          dCa/dt = -
→gamma * (Ca - Ca_rest) / (1 + kappa_S + kappa_B) : mmolar (clock-driven)\n
→kappa_B = B_T * K_d / (Ca + K_d)**2 : 1\n\n          CaB_active = Ca_rest + b :
→mmolar # add tiny bit to avoid /0\n          db/dt = beta : mmolar (clock-driven)\n
→lam = 1/tau_off + 1/tau_on : 1/second\n          kap = 1/tau_off : 1/
→second\n          dbeta/dt = ( # should be M/s/s\n
```

(continues on next page)

(continued from previous page)

```

→ A * (lam - kap) * (Ca - Ca_rest) # M/s/s\n          - (kap + lam) * beta
→ # M/s/s\n          - kap * lam * b # M/s/s\n          ) : mmolar/second\n
→ (clock-driven)\n          \nexc_factor = 1 : 1\n          dFF_baseline = 1 / (1\n
→ + (K_d / Ca_rest) ** n_H) : 1\n          dFF = exc_factor * rho_rel * dFF_max * (\n
→          1 / (1 + (K_d / CaB_active) ** n_H)\n          - dFF_baseline\n
→          ) : 1\n          rho_rel : 1\n          ', on_pre='Ca += dCa_T / (1 + kappa_S\n
→ + kappa_B)', sigma_noise=0.03748181818181818, dFF_1AP=0.097755000000000001, location=\n
→ 'cytoplasm', cal_model=DynamicCalcium(on_pre='Ca += dCa_T / (1 + kappa_S + kappa_B)',\n
→ model='\n          dCa/dt = -gamma * (Ca - Ca_rest) / (1 + kappa_S + kappa_B) :\n
→ mmolar (clock-driven)\n          kappa_B = B_T * K_d / (Ca + K_d)**2 : 1', Ca_\n
→ rest=50. * nmolar, gamma=292.3 * hertz, B_T=200. * umolar, kappa_S=110, dCa_T=7.6 *\n
→ umolar), bind_act_model=DoubExpCalBindingActivation(model='\n          CaB_active =\n
→ Ca_rest + b : mmolar # add tiny bit to avoid /0\n          db/dt = beta : mmolar\n
→ (clock-driven)\n          lam = 1/tau_off + 1/tau_on : 1/second\n          kap = 1/\n
→ tau_off : 1/second\n          dbeta/dt = (          # should be M/s/s\n
→          A * (lam - kap) * (Ca - Ca_rest) # M/s/s\n          - (kap + lam) *\n
→ beta # M/s/s\n          - kap * lam * b # M/s/s\n          ) :\n
→ mmolar/second (clock-driven)\n          ', A=7.61251 * khertz, tau_on=1.17164616 *\n
→ second, tau_off=10.14020867 * msecond, Ca_rest=50. * nmolar), exc_\n
→ model=NullExcitation(model='exc_factor = 1 : 1'), fluor_model='\n          dFF_\n
→ baseline = 1 / (1 + (K_d / Ca_rest) ** n_H) : 1\n          dFF = exc_factor * rho_\n
→ rel * dFF_max * (\n          1 / (1 + (K_d / CaB_active) ** n_H)\n
→ - dFF_baseline\n          ) : 1\n          rho_rel : 1\n          ', K_d=290. *\n
→ nmolar, n_H=2.7, dFF_max=25.2), img_width=0.5 * mmetre, focus_depth=100. * umetre,\n
→ location=array([0., 0., 0.]) * metre, direction=array([0., 0., 1.]), soma_radius=10. *\n
→ umetre, snr_cutoff=1), BansalFourStateOpsin(sim=..., name='VfChrimson', save_\n
→ history=True, on_pre='', spectrum=[(470.0, 0.4123404255319149), (490.0, 0.\n
→ 593265306122449), (510.0, 0.7935294117647058), (530.0, 0.8066037735849055), (550.0, 0.\n
→ 8912727272727272), (570.0, 1.0), (590.0, 0.9661016949152542), (610.0, 0.\n
→ 7475409836065574), (630.0, 0.4342857142857143), (1060, 0.01)], required_vars=[('Iopto',\n
→ amp), ('v', volt)], Gd1=0.37 * khertz, Gd2=175. * hertz, Gr0=0.667 * mhertz, g0=17.5\n
→ * nsiemens, phim=1.5e+22 * (second ** -1) / (meter ** 2), k1=3. * khertz, k2=200. *\n
→ hertz, Gf0=20. * hertz, Gb0=3.2 * hertz, kf=10. * hertz, kb=10. * hertz, gamma=0.05,\n
→ p=1, q=1, E=0. * volt, model='\n          dC1/dt = Gd1*O1 + Gr0*C2 - Ga1*C1 : 1 (clock-\n
→ driven)\n          dO1/dt = Ga1*C1 + Gb*O2 - (Gd1+Gf)*O1 : 1 (clock-driven)\n          dO2/\n
→ dt = Ga2*C2 + Gf*O1 - (Gd2+Gb)*O2 : 1 (clock-driven)\n          C2 = 1 - C1 - O1 - O2 :\n
→ 1\n\n          Theta = int(phi_pre > 0*phi_pre) : 1\n          Hp = Theta * phi_pre**p/\n
→ (phi_pre**p + phim**p) : 1\n          Ga1 = k1*Hp : hertz\n          Ga2 = k2*Hp : hertz\n
→          Hq = Theta * phi_pre**q/(phi_pre**q + phim**q) : 1\n          Gf = kf*Hq + Gf0 :\n
→ hertz\n          Gb = kb*Hq + Gb0 : hertz\n          fphi = O1 + gamma*O2 : 1\n\n\n
→ IOPTO_VAR_NAME_post = -g0*fphi*(V_VAR_NAME_post-E)*rho_rel : ampere (summed)\n
→ rho_rel : 1'}}

```

Now, to target these neurons with 2P laser power, we use `tp_light_from_scope()` to create a Light object with a 2P laser *GaussianEllipsoid* light profile centered on each of the scope's targets:

```

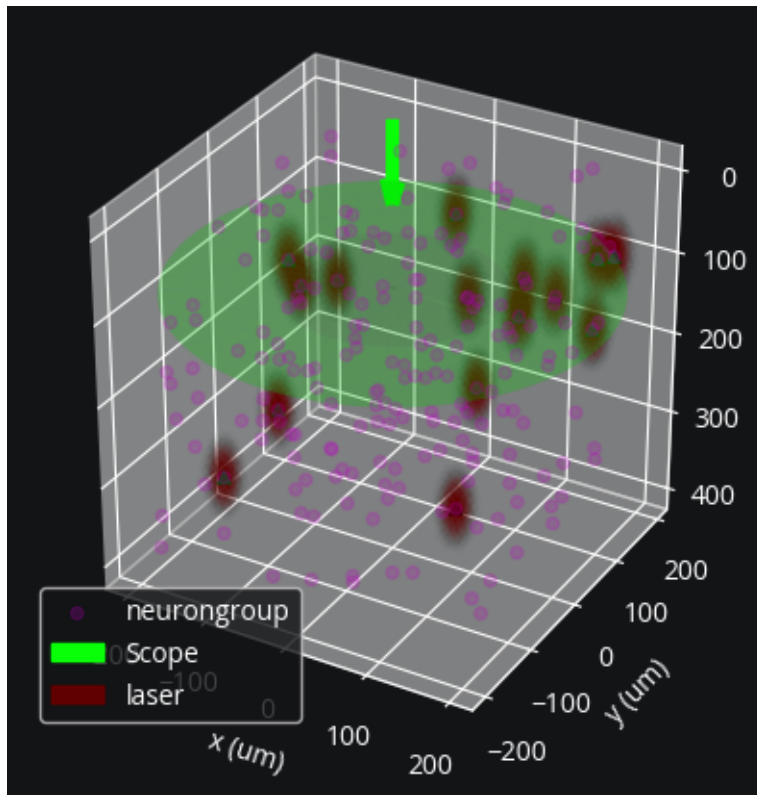
laser = light.tp_light_from_scope(scope, wavelength=1060*b2.nmeter, name='laser')
sim.inject(laser, ng)
cleo.viz.plot(ng, colors=['#c500cc'], sim=sim)

```

```

(<Figure size 640x480 with 1 Axes>,
 <Axes3D: xlabel='x (um)', ylabel='y (um)', zlabel='z (um)'>)

```



## Simulating an all-optical experiment

We'll stimulate each neuron with a different number of laser pulses and see the resulting calcium traces.

```
from cleo.ioproc import LatencyIOProcessor

# for seeing ground-truth spikes
i_all_targets = scope.i_targets_for_neuron_group(ng)
smon = b2.SpikeMonitor(ng, record=i_all_targets)
sim.network.add(smon)

amplitude_mW = 2.5
pulse_width_ms = 2
interpulse_ms = 20

num_pulses = np.arange(1, scope.n + 1)
train_ends_ms = interpulse_ms * num_pulses

class IOProc(LatencyIOProcessor):
    def process(self, state_dict, time_ms):
        t_in_cycle = time_ms % interpulse_ms
        on = (0 <= t_in_cycle) & (t_in_cycle < pulse_width_ms) & (time_ms < (interpulse_
        ms * num_pulses))

        return {"laser": on * amplitude_mW}, time_ms
```

(continues on next page)

(continued from previous page)

```
sim.set_io_processor(IOProc(1))

sim.reset()
tmax = 1000
sim.run(tmax * b2.ms)
```

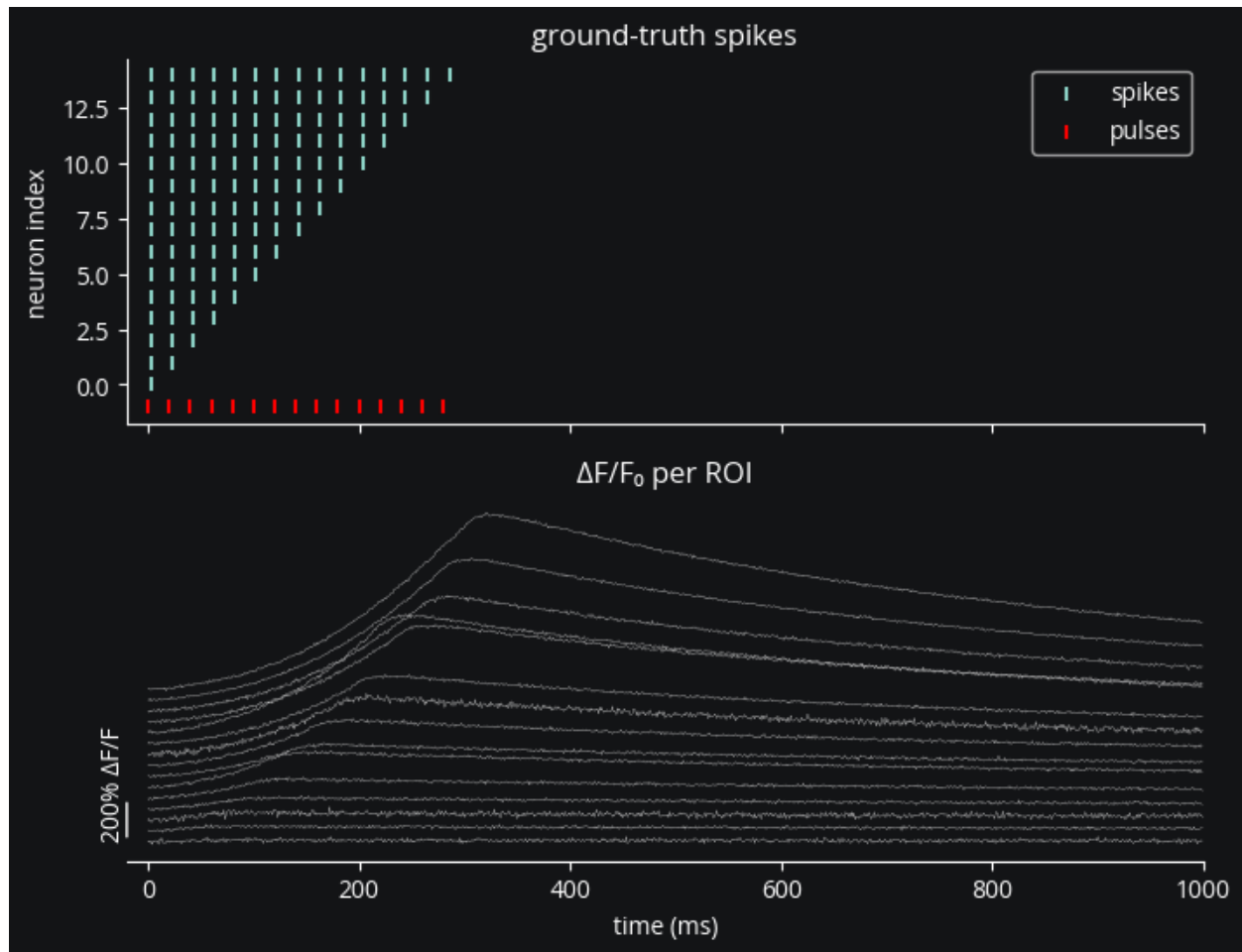
```
INFO      No numerical integration method specified for group 'neurongroup', using
↳ method 'exact' (took 0.18s). [brian2.stateupdaters.base.method_choice]
INFO      No numerical integration method specified for group 'syn_VfChrimson_
↳ neurongroup', using method 'euler' (took 0.02s, trying other methods took 0.06s).
↳ [brian2.stateupdaters.base.method_choice]
INFO      No numerical integration method specified for group 'syn_gcamp6f_neurongroup',
↳ using method 'euler' (took 0.02s, trying other methods took 0.05s). [brian2.
↳ stateupdaters.base.method_choice]
```

Now we'll plot the result:

```
import matplotlib.pyplot as plt
sep = .5
n2plot = len(i_all_targets)
fig, (ax1, ax2) = plt.subplots(2, 1, figsize=(8, 6), sharex=True)

spt = np.empty((0,))
spi = np.empty((0,))
for i_target, i_ng in enumerate(i_all_targets):
    single_spt = smon.t[smon.i == i_ng]
    spt = np.concatenate((spt, single_spt))
    spi = np.concatenate((spi, np.full_like(single_spt, i_target)))
ax1.scatter(spt / b2.ms, spi, marker='|', label='spikes')
t_pulse_ms = np.arange(0, num_pulses.max() * interpulse_ms, interpulse_ms)
ax1.scatter(t_pulse_ms, np.full_like(t_pulse_ms, -1), marker='|', color='r', label=
↳ 'pulses')
ax1.set(ylabel='neuron index', title='ground-truth spikes', xlim=(0, tmax))
ax1.legend()

ax2.plot(scope.t_ms[:,], np.array(scope.dFF)[:, :n2plot] + sep*np.arange(n2plot), lw=.2,
↳ c='w')
y_scale = 2
y0 = 0
x0 = -20
ax2.set(xlabel='time (ms)', title='F/F0 per ROI', xlim=(x0, tmax))
ax2.annotate('', xy=(x0, y0), xytext=(x0, y0 + y_scale), arrowprops=dict(arrowstyle='-'))
ax2.annotate(f'{y_scale * 100}% F/F', xy=(x0, y0), va='bottom', ha='right', rotation=90)
ax2.yaxis.set_visible(False)
ax2.spines['left'].set_visible(False)
```



Results are as expected:

- We see one spike for every pulse, though for longer trains we might start to see missed spikes due to the opsin activation plateauing.
- If neurons were closer together, we'd expect to see some off-target effects; if there are any here, they are apparently subthreshold.
- Signal strength is proportional to the number of spikes, as expected, but varies somewhat due to heterogeneous expression.
- We also see that noise levels vary from ROI to ROI due to varying distances from the focal plane.

## 6.2.8 Video visualization

In this tutorial we'll see how to inject a video visualizer into a simulation.

Preamble:

```
import brian2.only as b2
from brian2 import np
import matplotlib.pyplot as plt

import cleo
```

(continues on next page)

(continued from previous page)

```

cleo.utilities.style_plots_for_docs()

# numpy faster than cython for lightweight example
b2.prefs.codegen.target = 'numpy'
# for reproducibility
np.random.seed(1866)

c_exc = 'xkcd:tomato'
c_inh = 'xkcd:cerulean blue'

```

```

WARNING      /home/kyle/miniforge3/envs/cleo/lib/python3.12/site-packages/attr/_make.
→py:918: RuntimeWarning: Running interpreter doesn't sufficiently support code object_
→introspection. Some features like bare super() or accessing __class__ will not work_
→with slotted classes.
      set_closure_cell(cell, cls)
[py.warnings]

```

## Set up the simulation

### Network

We'll use excitatory and inhibitory populations of [exponential integrate-and-fire neurons](#).

```

n_e = 400
n_i = n_e // 4
def eif(n, name):
    ng = b2.NeuronGroup(
        n,
        """
        dv/dt =  $-(v - E_L) + \Delta_T \exp((v - \theta) / \Delta_T) + R_m I$  / tau_m : volt
        I : amp
        """,
        threshold="v>30*mV",
        reset="v=-55*mV",
        namespace={
            "tau_m": 20 * b2.ms,
            "Rm": 500 * b2.Mohm,
            "theta": -50 * b2.mV,
            "Delta_T": 2 * b2.mV,
            "E_L": -70 * b2.mV,
        },
        name=name,
    )
    ng.v = -70 * b2.mV
    return ng

exc = eif(n_e, "exc")
inh = eif(n_i, "inh")
W = 250

```

(continues on next page)

(continued from previous page)

```

p_S = 0.3
S_ei = b2.Synapses(exc, inh, on_pre="v_post+=W*mV/n_e")
S_ei.connect(p=p_S)
S_ie = b2.Synapses(inh, exc, on_pre="v_post-=W*mV/n_i")
S_ie.connect(p=p_S)
S_ee = b2.Synapses(exc, exc, on_pre="v_post+=W*mV/n_e")
S_ee.connect(condition='abs(i-j)<=20')

mon_e = b2.SpikeMonitor(exc)
mon_i = b2.SpikeMonitor(inh)

net = b2.Network(exc, inh, S_ei, S_ie, S_ee, mon_e, mon_i)

```

## Coordinates and optogenetics

Here we configure the coordinates and optogenetic stimulation. For more details, see the *“Optogenetic stimulation” tutorial*. Note that we save the arguments used in the plotting function for reuse later on when generating the video.

```

from cleo.coords import assign_coords_uniform_cylinder
from cleo.viz import plot

r = 1
assign_coords_uniform_cylinder(
    exc, xyz_start=(0, 0, 0.3), xyz_end=(0, 0, 0.4), radius=r
)
assign_coords_uniform_cylinder(
    inh, xyz_start=(0, 0, 0.3), xyz_end=(0, 0, 0.4), radius=r
)

```

```

from cleo.opto import chr2_4s
from cleo.light import fiber473nm, Light

opsin = chr2_4s()
fibers = Light(
    coords=[[0, 0, 0], [700, 0, 0]] * b2.um,
    light_model=fiber473nm(),
    max_Irr0_mW_per_mm2=30,
    save_history=True,
)

plotargs = {
    "colors": [c_exc, c_inh],
    "zlim": (0, 600),
    "scatterargs": {"s": 20}, # to adjust neuron marker size
    "axis_scale_unit": b2.um,
}

plot(
    exc,
    inh,

```

(continues on next page)



(continued from previous page)

```

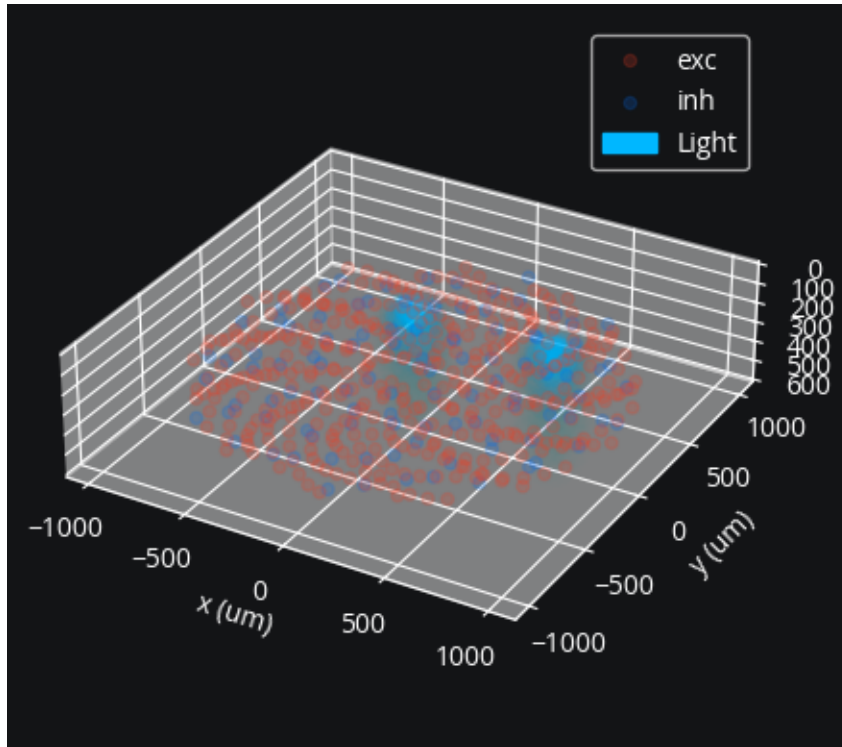
**plotargs,
devices=[fibers],
)

```

```

(<Figure size 640x480 with 1 Axes>,
 <Axes3D: xlabel='x (um)', ylabel='y (um)', zlabel='z (um)'>)

```



## Simulator, optogenetics injection

Here we create the simulator and inject the OptogeneticIntervention.

```

sim = cleo.CLSimulator(net)
sim.inject(opsin, exc, Iopto_var_name='I')
sim.inject(fibers, exc)

```

```

CLSimulator(io_processor=None, devices={Light(name='Light', save_history=True,
→ value=array([0., 0.]), light_model=OpticFiber(R0=100. * umetre, NAfib=0.37, K=125. *
→ metre ** -1, S=7370. * metre ** -1, ntis=1.36), wavelength=0.473 * umetre,
→ direction=array([0., 0., 1.]), max_Irr0_mW_per_mm2=30, max_Irr0_mW_per_mm2_viz=None),
→ FourStateOpsin(name='Chr2', save_history=True, on_pre='', spectrum=[(400, 0.34), (422,
→ 0.65), (460, 0.96), (470, 1), (473, 1), (500, 0.57), (520, 0.22), (540, 0.06), (560, 0.
→ 01)], required_vars=[('Iopto', amp), ('v', volt)], g0=114. * nsiemens, gamma=0.00742,
→ phim=2.33e+23 * (second ** -1) / (meter ** 2), k1=4.15 * khertz, k2=0.868 * khertz,
→ p=0.833, Gf0=37.3 * hertz, kf=58.1 * hertz, Gb0=16.1 * hertz, kb=63. * hertz, q=1.94,
→ Gd1=105. * hertz, Gd2=13.8 * hertz, Gr0=0.33 * hertz, E=0. * volt, v0=43. * mvolt,
→ v1=17.1 * mvolt, model="\n          dC1/dt = Gd1*O1 + Gr0*C2 - Ga1*C1 : 1 (clock-driven)\n

```

(continues on next page)

(continued from previous page)

```

→n      d01/dt = Ga1*C1 + Gb*02 - (Gd1+Gf)*01 : 1 (clock-driven)\n      d02/dt =
→Ga2*C2 + Gf*01 - (Gd2+Gb)*02 : 1 (clock-driven)\n      C2 = 1 - C1 - 01 - 02 : 1\n\n
→      Theta = int(phi_pre > 0*phi_pre) : 1\n      Hp = Theta * phi_pre**p/(phi_
→pre**p + phim**p) : 1\n      Ga1 = k1*Hp : hertz\n      Ga2 = k2*Hp : hertz\n
→      Hq = Theta * phi_pre**q/(phi_pre**q + phim**q) : 1\n      Gf = kf*Hq + Gf0 : hertz\n
→n      Gb = kb*Hq + Gb0 : hertz\n\n      fphi = 01 + gamma*02 : 1\n      # v1/v0
→when v-E == 0 via l'Hopital's rule\n      fv = f_unless_x0(\n      (1 - exp(-
→(V_VAR_NAME_post-E)/v0)) / ((V_VAR_NAME_post-E)/v1),\n      V_VAR_NAME_post - E,\n
→n      v1/v0\n      ) : 1\n\n      IOPTO_VAR_NAME_post = -g0*fphi*fv*(V_VAR_
→NAME_post-E)*rho_rel : ampere (summed)\n      rho_rel : 1", extra_namespace={'f_
→unless_x0': <brian2.core.functions.Function object at 0x7f4f3c391c40>}}))

```

## Processor

And we set up open-loop optogenetic stimulation:

```

fibers.update([5, 0])
class OpenLoopOpto(cleo.ioproc.LatencyIOProcessor):
    def process(self, state_dict, time_ms):
        # random walk stimulation
        fiber_intensities = fibers.value + np.random.randn(2)*.5
        fiber_intensities[fiber_intensities < 0] = 0
        if time_ms > 50:
            fiber_intensities = [0, 5]
        return ({"Light": fiber_intensities}, time_ms)

sim.set_io_processor(OpenLoopOpto(sample_period_ms=1))

```

```

CLSimulator(io_processor=OpenLoopOpto(sample_period_ms=1, sampling='fixed', processing=
→'parallel'), devices={Light(name='Light', save_history=True, value=array([5, 0]),
→light_model=OpticFiber(R0=100. * umetre, NAfib=0.37, K=125. * metre ** -1, S=7370. *
→metre ** -1, ntis=1.36), wavelength=0.473 * umetre, direction=array([0., 0., 1.]), max_
→Irr0_mW_per_mm2=30, max_Irr0_mW_per_mm2_viz=None), FourStateOpsin(name='ChR2', save_
→history=True, on_pre='', spectrum=[(400, 0.34), (422, 0.65), (460, 0.96), (470, 1),
→(473, 1), (500, 0.57), (520, 0.22), (540, 0.06), (560, 0.01)], required_vars=[('Iopto',
→amp), ('v', volt)], g0=114. * nsiemens, gamma=0.00742, phim=2.33e+23 * (second ** -1)
→/ (meter ** 2), k1=4.15 * khertz, k2=0.868 * khertz, p=0.833, Gf0=37.3 * hertz, kf=58.
→1 * hertz, Gb0=16.1 * hertz, kb=63. * hertz, q=1.94, Gd1=105. * hertz, Gd2=13.8 *
→hertz, Gr0=0.33 * hertz, E=0. * volt, v0=43. * mvolt, v1=17.1 * mvolt, model="\n
→dC1/dt = Gd1*01 + Gr0*C2 - Ga1*C1 : 1 (clock-driven)\n      d01/dt = Ga1*C1 + Gb*02
→- (Gd1+Gf)*01 : 1 (clock-driven)\n      d02/dt = Ga2*C2 + Gf*01 - (Gd2+Gb)*02 : 1
→(clock-driven)\n      C2 = 1 - C1 - 01 - 02 : 1\n\n      Theta = int(phi_pre >
→0*phi_pre) : 1\n      Hp = Theta * phi_pre**p/(phi_pre**p + phim**p) : 1\n
→Ga1 = k1*Hp : hertz\n      Ga2 = k2*Hp : hertz\n      Hq = Theta * phi_pre**q/(phi_
→pre**q + phim**q) : 1\n      Gf = kf*Hq + Gf0 : hertz\n      Gb = kb*Hq + Gb0 :
→hertz\n\n      fphi = 01 + gamma*02 : 1\n      # v1/v0 when v-E == 0 via l'Hopital
→'s rule\n      fv = f_unless_x0(\n      (1 - exp(-(V_VAR_NAME_post-E)/v0)) /
→((V_VAR_NAME_post-E)/v1),\n      V_VAR_NAME_post - E,\n      v1/v0\n
→) : 1\n\n      IOPTO_VAR_NAME_post = -g0*fphi*fv*(V_VAR_NAME_post-E)*rho_rel :
→ampere (summed)\n      rho_rel : 1", extra_namespace={'f_unless_x0': <brian2.core.
→functions.Function object at 0x7f4f3c391c40>}}))

```

## Inject VideoVisualizer

A VideoVisualizer is an InterfaceDevice like recorders and stimulators and needs to be injected in order to properly interact with the Brian network. Keep in mind the following:

- It must be injected *after* all other devices for the `devices='all'` argument to work as expected.
- Similarly to recording and stimulation, you must specify the target neuron groups (to display, in this case) on injection
- The `dt` argument makes a huge difference on the amount of time it takes to generate the video. You may want to keep this high while experimenting and only lower it when you are ready to generate a high-quality video since the process is so slow.

```
vv = cleo.viz.VideoVisualizer(dt=1 * b2.ms, devices="all")
sim.inject(vv, exc, inh)
```

```
CLSimulator(io_processor=OpenLoopOpto(sample_period_ms=1, sampling='fixed', processing=
→ 'parallel'), devices={VideoVisualizer(name='VideoVisualizer', save_history=True,
→ devices=[Light(name='Light', save_history=True, value=array([5, 0]), light_
→ model=OpticFiber(R0=100. * umetre, NAfib=0.37, K=125. * metre ** -1, S=7370. * metre_
→ ** -1, ntis=1.36), wavelength=0.473 * umetre, direction=array([0., 0., 1.]), max_Irr0_
→ mW_per_mm2=30, max_Irr0_mW_per_mm2_viz=None)], dt=1. * msecond, fig=None, ax=None,
→ neuron_groups=[NeuronGroup(clock=Clock(dt=100. * usecond, name='defaultclock'),
→ when=start, order=0, name='exc'), NeuronGroup(clock=Clock(dt=100. * usecond, name=
→ 'defaultclock'), when=start, order=0, name='inh')], _spike_mons=[<SpikeMonitor,
→ recording from 'spikemonitor_2'>, <SpikeMonitor, recording from 'spikemonitor_3'>], _
→ num_old_spikes=[0, 0], _value_per_device_per_frame=[], _i_spikes_per_ng_per_frame=[]),
→ Light(name='Light', save_history=True, value=array([5, 0]), light_
→ model=OpticFiber(R0=100. * umetre, NAfib=0.37, K=125. * metre ** -1, S=7370. * metre_
→ ** -1, ntis=1.36), wavelength=0.473 * umetre, direction=array([0., 0., 1.]), max_Irr0_
→ mW_per_mm2=30, max_Irr0_mW_per_mm2_viz=None), FourStateOpsin(name='Chr2', save_
→ history=True, on_pre='', spectrum=[(400, 0.34), (422, 0.65), (460, 0.96), (470, 1),
→ (473, 1), (500, 0.57), (520, 0.22), (540, 0.06), (560, 0.01)], required_vars=[('Iopto',
→ amp), ('v', volt)], g0=114. * nsiemens, gamma=0.00742, phim=2.33e+23 * (second ** -1),
→ / (meter ** 2), k1=4.15 * khertz, k2=0.868 * khertz, p=0.833, Gf0=37.3 * hertz, kf=58.
→ 1 * hertz, Gb0=16.1 * hertz, kb=63. * hertz, q=1.94, Gd1=105. * hertz, Gd2=13.8 *
→ hertz, Gr0=0.33 * hertz, E=0. * volt, v0=43. * mvolt, v1=17.1 * mvolt, model="\n
→ dC1/dt = Gd1*O1 + Gr0*C2 - Ga1*C1 : 1 (clock-driven)\n          dO1/dt = Ga1*C1 + Gb*O2
→ - (Gd1+Gf)*O1 : 1 (clock-driven)\n          dO2/dt = Ga2*C2 + Gf*O1 - (Gd2+Gb)*O2 : 1
→ (clock-driven)\n          C2 = 1 - C1 - O1 - O2 : 1\n\n          Theta = int(phi_pre >
→ 0*phi_pre) : 1\n          Hp = Theta * phi_pre**p/(phi_pre**p + phim**p) : 1\n
→ Ga1 = k1*Hp : hertz\n          Ga2 = k2*Hp : hertz\n          Hq = Theta * phi_pre**q/(phi_
→ pre**q + phim**q) : 1\n          Gf = kf*Hq + Gf0 : hertz\n          Gb = kb*Hq + Gb0 :
→ hertz\n\n          fphi = O1 + gamma*O2 : 1\n          # v1/v0 when v-E == 0 via l'Hopital
→ 's rule\n          fv = f_unless_x0(\n          (1 - exp(-(V_VAR_NAME_post-E)/v0)) /
→ ((V_VAR_NAME_post-E)/v1),\n          V_VAR_NAME_post - E,\n          v1/v0\n
→ ) : 1\n\n          IOPTO_VAR_NAME_post = -g0*fphi*f_v*(V_VAR_NAME_post-E)*rho_rel :
→ ampere (summed)\n          rho_rel : 1", extra_namespace={'f_unless_x0': <brian2.core.
→ functions.Function object at 0x7f4f3c391c40>}}))
```

## Run simulation and visualize

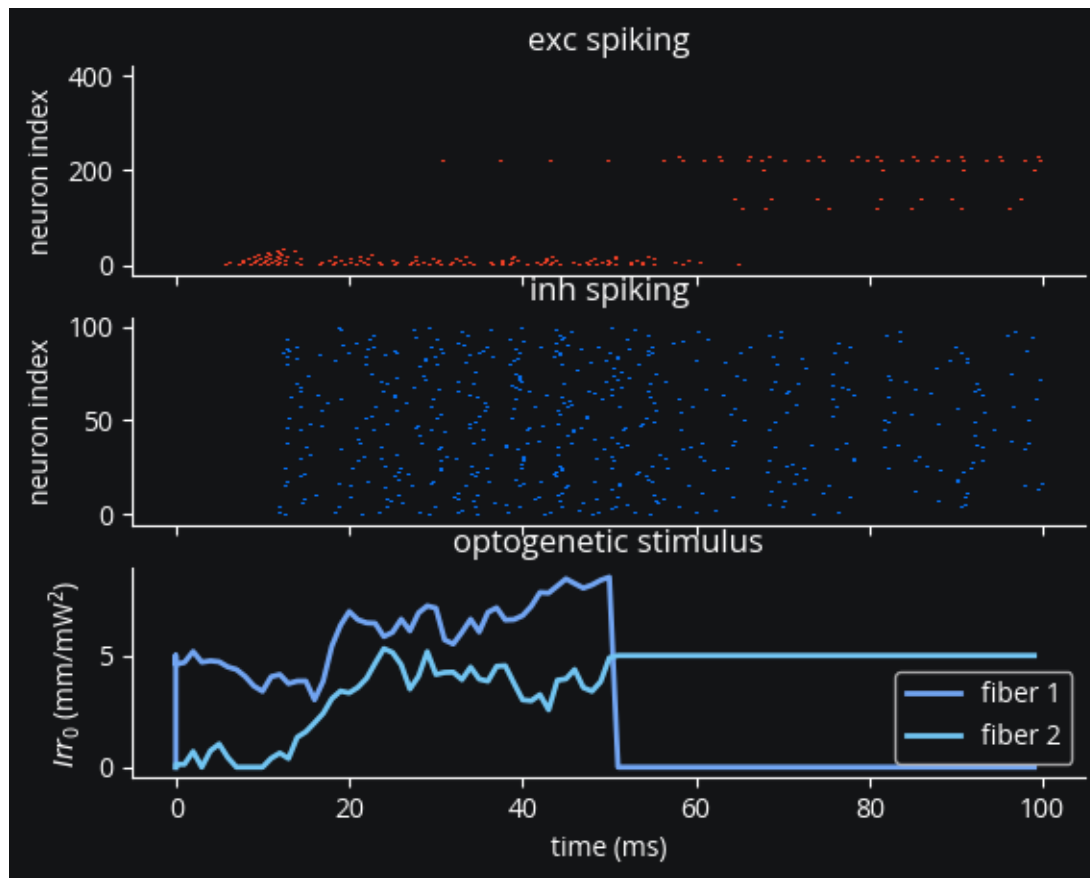
Here we display a quick plot before generating the video:

```
T = 100
sim.run(T * b2.ms)
```

```
WARNING      'T' is an internal variable of group 'light_prop_ChR2_exc', but also exists
↳ in the run namespace with the value 100. The internal variable will be used. [brian2.
↳ groups.group.Group.resolve.resolution_conflict]
INFO         No numerical integration method specified for group 'exc', using method 'euler
↳ ' (took 0.01s, trying other methods took 0.06s). [brian2.stateupdaters.base.method_
↳ choice]
INFO         No numerical integration method specified for group 'inh', using method 'euler
↳ ' (took 0.01s, trying other methods took 0.01s). [brian2.stateupdaters.base.method_
↳ choice]
```

```
INFO         No numerical integration method specified for group 'syn_ChR2_exc', using
↳ method 'euler' (took 0.02s, trying other methods took 0.15s). [brian2.stateupdaters.
↳ base.method_choice]
```

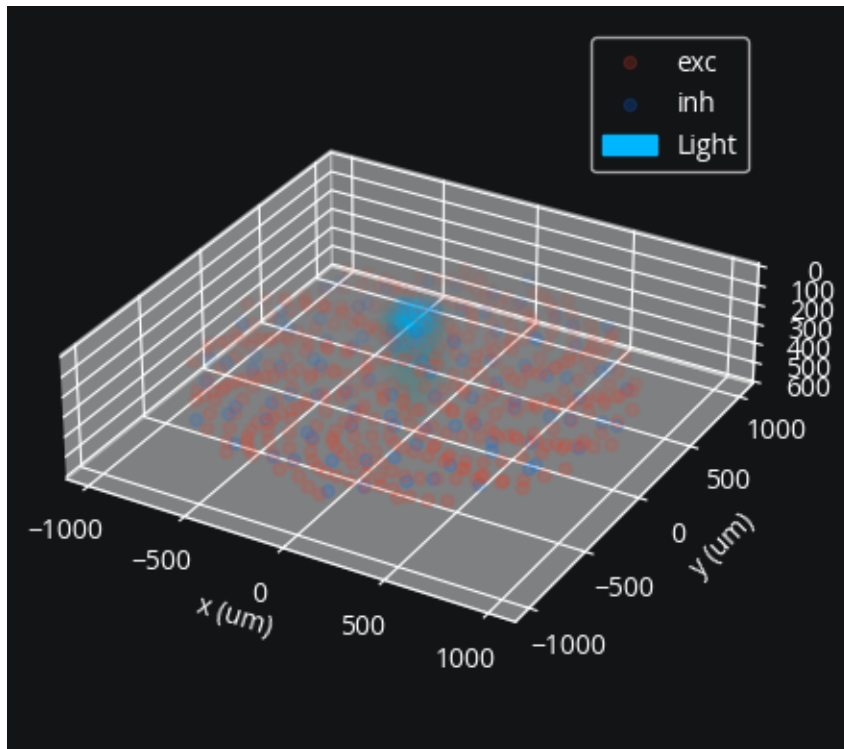
```
fig, (ax1, ax2, ax3) = plt.subplots(3, 1, sharex=True)
stim_vals = np.array(fibers.values)
sptexc = mon_e.spike_trains()
ax1.eventplot([t/b2.ms for t in sptexc.values()], lineoffsets=list(sptexc.keys()),
↳ color=c_exc)
ax1.set(ylabel="neuron index", title="exc spiking")
sptinh = mon_i.spike_trains()
ax2.eventplot([t/b2.ms for t in sptinh.values()], lineoffsets=list(sptinh.keys()),
↳ color=c_inh)
ax2.set(ylabel="neuron index", title="inh spiking")
ax3.plot(fibers.t_ms, stim_vals[:, 0], c="#72a5f2", lw=2, label='fiber 1')
ax3.plot(fibers.t_ms, stim_vals[:, 1], c="#72c5f2", lw=2, label='fiber 2')
ax3.legend()
ax3.set(ylabel=r"$I_{rr,0}$ (mm/mW$^2$)", title="optogenetic stimulus", xlabel="time (ms)");
```



The `VideoVisualizer` stores the data it needs during the simulation, but hasn't yet produced any visual output. We first use the `generate_Animation()`, plugging in the arguments we used for the original plot.

Also, we set the `max_Irr0_mW_per_mm2_viz` attribute of the optogenetic intervention. This effectively scales how bright the light appears in the visualization. That is, a high maximum irradiance makes the stimulus values small in comparison and produces a faint light, while a low ceiling makes the values relatively large and produces a bright light in the resulting video.

```
fibers.max_Irr0_mW_per_mm2_viz = np.max(stim_vals)
ani = vv.generate_Animation(plotargs, slowdown_factor=10)
```



The `generate_Animation()` function returns a matplotlib `FuncAnimation` object, which you can then use however you want. You will probably want to [save a video](#).

Note that at this point the video still hasn't been rendered; that happens when you try and save or visualize the animation. This step takes a while if your temporal resolution is high, so we suggest you do this only after your experiment is finalized and after you've experimented with low framerate videos to finalize video parameters.

Here we embed the video using HTML so you can see the output:

```
from matplotlib import rc
rc('animation', html='jshtml')

ani
```

```
<matplotlib.animation.FuncAnimation at 0x7f4f33cad250>
```

## 6.2.9 Neo export

Cleo allows you to export data from the whole *CLSimulator* or from individual *InterfaceDevices* objects to *Neo* objects. This facilitates data analysis and visualization with packages that take Neo as an input format, as well as export to various *open* and *proprietary* ephys data file formats, which could be useful for integration with existing pipelines developed for experimental data.

### Setup

```
import brian2 as b2
from brian2 import np
import cleo
from cleo import opto, ephys, light
import neo

# numpy faster than cython for lightweight example
b2.prefs.codegen.target = 'numpy'
# for reproducibility
np.random.seed(33)

cleo.utilities.style_plots_for_docs()
```

We'll set up a basic simulation to see how export works for different devices and the whole simulation.

```
ng = b2.NeuronGroup(
    100,
    """dv/dt = ((-70*mV - v) + (500*Mohm)*Iopto) / (20*ms) : volt
    Iopto : amp""",
    threshold="v > -50*mV",
    reset="v = -70*mV",
)
ng.v = -70 * b2.mV
syn = b2.Synapses(ng, on_pre="v += 3*mV")
cleo.coords.assign_coords_rand_rect_prism(ng, (-.2, .2), (-.2, .2), (0, .4))

light = light.Light(
    coords=[[-0.1, 0, 0], [0.1, 0, 0]] * b2.mm, light_model=light.fiber473nm()
)
probe = ephys.Probe(
    coords=[[0, 0, 50], [0, 0, 150], [0, 0, 250]] * b2.um,
    signals=[
        ephys.TKLFPSignal(),
        ephys.MultiUnitSpiking(
            r_perfect_detection=50 * b2.um, r_half_detection=100 * b2.um
        ),
        ephys.SortedSpiking(
            r_perfect_detection=50 * b2.um, r_half_detection=100 * b2.um
        ),
    ],
)
sim = cleo.CLSimulator(b2.Network(ng))
```

(continues on next page)

(continued from previous page)

```

class IOProc(cleo.ioproc.LatencyIOProcessor):
    def process(self, state_dict, t_samp_ms):
        return {'Light': .5 * np.random.rand(light.n)}, t_samp_ms
sim.set_io_processor(IOProc(1))
sim.inject(light, ng).inject(probe, ng, tklf_type="exc")
sim.inject(opto.chr2_4s(), ng)

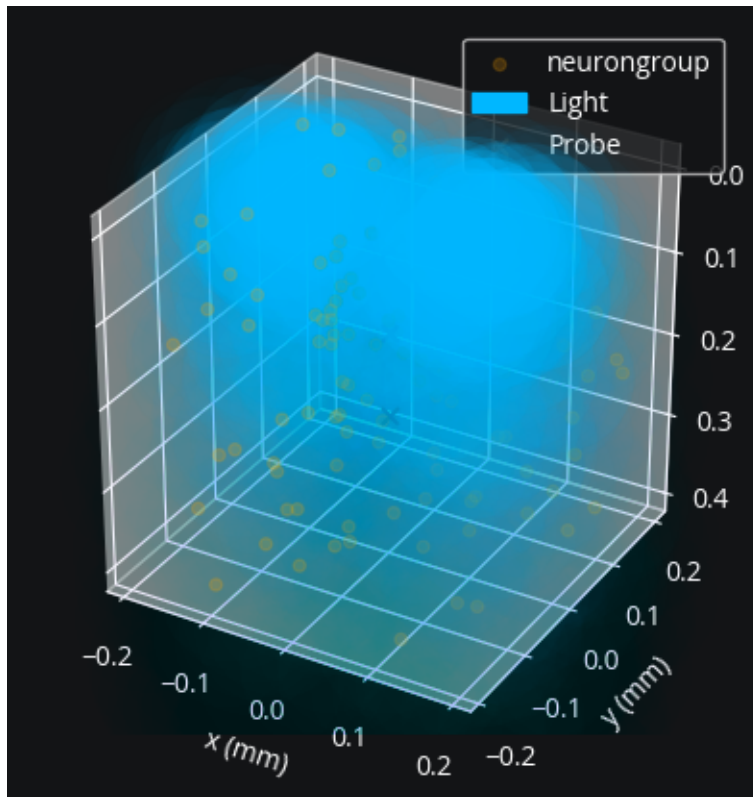
cleo.viz.plot(ng, colors=['orange'], sim=sim)

```

```

(<Figure size 640x480 with 1 Axes>,
 <Axes3D: xlabel='x (mm)', ylabel='y (mm)', zlabel='z (mm)'>)

```



```
sim.run(50 * b2.ms)
```

```

INFO      No numerical integration method specified for group 'neurongroup', using
→method 'exact' (took 0.23s). [brian2.stateupdaters.base.method_choice]
INFO      No numerical integration method specified for group 'syn_ChR2_neurongroup',
→using method 'euler' (took 0.02s, trying other methods took 0.06s). [brian2.
→stateupdaters.base.method_choice]

```



## Exporting individual devices

Depending on whether time values are regularly spaced, data is exported to either `AnalogSignal` or `IrregularlySampledSignal` objects, with appropriate annotations and per-channel annotations.

```
light.to_neo()
```

```
AnalogSignal with 2 channels of length 50; units mW/mm**2; datatype float64
name: 'Light'
description: 'Exported from Cleo Light device'
annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 181618)}
sampling rate: 1.0 1/ms
time: 0.0 ms to 50.0 ms
```

```
light.to_neo().array_annotations
```

```
{'x': array([-0.1,  0.1]) * mm,
 'y': array([0.,  0.]) * mm,
 'z': array([0.,  0.]) * mm,
 'direction_x': array([0.,  0.]),
 'direction_y': array([0.,  0.]),
 'direction_z': array([1.,  1.]),
 'i_channel': array([0, 1])}
```

Different signals from the same device are grouped together:

```
probe_neo = probe.to_neo()
probe_neo
```

```
Group with 2 groups, 1 analogsignals
name: 'Probe'
description: 'Exported from Cleo Probe device'
```

```
probe_neo.children
```

```
(AnalogSignal with 3 channels of length 50; units uV; datatype float64
name: 'Probe.TKLFPSignal'
description: 'Exported from Cleo TKLFPSignal object'
annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 239616)}
sampling rate: 1.0 1/ms
time: 0.0 ms to 50.0 ms,
Group with 3 spiketrains
name: 'Probe.MultiUnitSpiking'
description: 'Exported from Cleo MultiUnitSpiking object'
annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 241341)}},
Group with 55 spiketrains
name: 'Probe.SortedSpiking'
description: 'Exported from Cleo SortedSpiking object'
annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 265177)})
```

## Whole-simulation export

Exporting the whole *CLSimulator* object generates a *Block* object with a single *Segment* object containing all the data from the simulation.

```
sim.to_neo()
```

```
Block with 1 segments, 1 groups
description: 'Exported from Cleo simulation'
annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 320150)}
# segments (N=1)
0: Segment with 2 analogsignals, 58 spiketrains
  # analogsignals (N=2)
    0: AnalogSignal with 2 channels of length 50; units mW/mm**2; datatype float64
      name: 'Light'
      description: 'Exported from Cleo Light device'
      annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 320674)}
      sampling rate: 1.0 1/ms
      time: 0.0 ms to 50.0 ms
    1: AnalogSignal with 3 channels of length 50; units uV; datatype float64
      name: 'Probe.TKLFPSignal'
      description: 'Exported from Cleo TKLFPSignal object'
      annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 321221)}
      sampling rate: 1.0 1/ms
      time: 0.0 ms to 50.0 ms
```

## Trial structure

Using the convention that a *Segment* represents a single “trial,” there are a couple ways we can generate trial-structured Neo data:

1. Resetting the simulation and exporting the data from each trial individually, merging them into a single *Block* object.
2. Structuring multiple trials into a single call to *run()* and segmenting the data on export. This is not implemented, but could be in the future if there is sufficient demand.

Let’s try the first of these options:

```
block1 = sim.to_neo()
block1.segments[0].name = 'trial 1'
sim.reset()
sim.run(50 * b2.ms)
block2 = sim.to_neo()
block2.segments[0].name = 'trial 2'

block = neo.Block()
block.segments.extend(block1.segments)
block.segments.extend(block2.segments)
block.groups.extend(block1.groups)
block.groups.extend(block2.groups)
block
```

```

Block with 2 segments, 2 groups
# segments (N=2)
0: Segment with 2 analogsignals, 58 spiketrains
  name: 'trial 1'
  # analogsignals (N=2)
  0: AnalogSignal with 2 channels of length 50; units mW/mm**2; datatype float64
    name: 'Light'
    description: 'Exported from Cleo Light device'
    annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 375717)}
    sampling rate: 1.0 1/ms
    time: 0.0 ms to 50.0 ms
  1: AnalogSignal with 3 channels of length 50; units uV; datatype float64
    name: 'Probe.TKLFPSignal'
    description: 'Exported from Cleo TKLFPSignal object'
    annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 33, 376227)}
    sampling rate: 1.0 1/ms
    time: 0.0 ms to 50.0 ms
1: Segment with 2 analogsignals, 63 spiketrains
  name: 'trial 2'
  # analogsignals (N=2)
  0: AnalogSignal with 2 channels of length 50; units mW/mm**2; datatype float64
    name: 'Light'
    description: 'Exported from Cleo Light device'
    annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 34, 138750)}
    sampling rate: 1.0 1/ms
    time: 0.0 ms to 50.0 ms
  1: AnalogSignal with 3 channels of length 50; units uV; datatype float64
    name: 'Probe.TKLFPSignal'
    description: 'Exported from Cleo TKLFPSignal object'
    annotations: {'export_datetime': datetime.datetime(2023, 9, 6, 9, 59, 34, 139149)}
    sampling rate: 1.0 1/ms
    time: 0.0 ms to 50.0 ms

```

The more attractive approach suggested by Neo's API of `block1.merge(block2)` does not work, but could potentially be fixed in the future.

## Example analysis use case

Let's try `elephant`, following their [statistics tutorial](#):

```

import matplotlib.pyplot as plt
from elephant.statistics import instantaneous_rate, time_histogram, mean_firing_rate
import quantities as pq

fig, ax = plt.subplots()
st1 = block.segments[0].spiketrains[0]
histogram_rate = time_histogram([st1], 5*pq.ms, output='rate')
inst_rate = instantaneous_rate(st1, sampling_period=1*pq.ms)

# plotting the original spiketrain
ax.plot(
    st1,

```

(continues on next page)

(continued from previous page)

```

    [0] * len(st1),
    "r",
    marker=2,
    ms=25,
    markeredgewidth=2,
    lw=0,
    label="poisson spike times",
)

# mean firing rate
ax.hlines(
    mean_firing_rate(st1),
    xmin=st1.t_start,
    xmax=st1.t_stop,
    linestyle="--",
    label="mean firing rate",
)

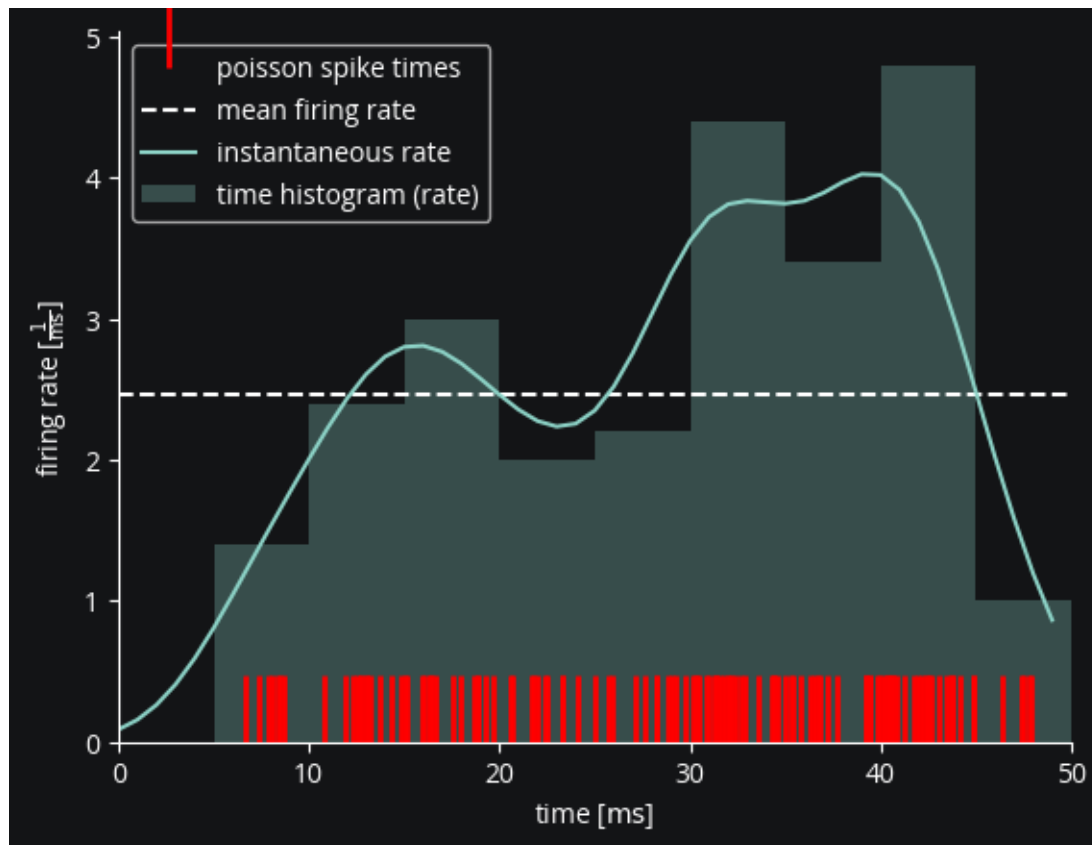
# time histogram
ax.bar(
    histogram_rate.times,
    histogram_rate.magnitude.flatten(),
    width=histogram_rate.sampling_period,
    align="edge",
    alpha=0.3,
    label="time histogram (rate)",
)

# instantaneous rate
ax.plot(
    inst_rate.times.rescale(pq.ms),
    inst_rate.rescale(histogram_rate.dimensionality).magnitude.flatten(),
    label="instantaneous rate",
)

# axis labels and legend
ax.set(
    xlabel=f"time [{st1.times.dimensionality.latex}]",
    ylabel=f"firing rate [{histogram_rate.dimensionality.latex}]",
    xlim=(st1.t_start, st1.t_stop),
)
ax.legend()

```

```
<matplotlib.legend.Legend at 0x7f527c923e20>
```



### 6.2.10 Advanced LFP

Here we will demonstrate the diverse ways the different LFP proxies can be computed and compare them to each other.

```
# preamble:
import brian2.only as b2
from brian2 import np
import matplotlib.pyplot as plt
import cleo
from cleo import ephys
import cleo.utilities

# the default cython compilation target isn't worth it for
# this trivial example
b2.prefs.codegen.target = "numpy"
b2.seed(18010601)
np.random.seed(18010601)
rng = np.random.default_rng(18010601)

cleo.utilities.style_plots_for_docs()

# colors
c = {
    "light": "#df87e1",
    "main": "#C500CC",
```

(continues on next page)

(continued from previous page)

```

    "dark": "#8000B4",
    "exc": "#d6755e",
    "inh": "#056eee",
    "accent": "#36827F",
}

```

## Network setup

First we need a point neuron simulation to approximate the LFP for. Here we adapt a balanced E/I network implementation from the [Neuronal Dynamics](#) textbook, using some parameters from [Mazzoni, Lindén et al., 2015](#).

```

n_exc = 800
n_inh = None # None = N_excit / 4
n_ext = 100
connection_probability = 0.2
w0 = 0.07 * b2.nA
g = 4
synaptic_delay = 1 * b2.ms
poisson_input_rate = 220 * b2.Hz
w_ext = 0.091 * b2.nA
v_rest = -70 * b2.mV
v_reset = -60 * b2.mV
firing_threshold = -50 * b2.mV
membrane_time_scale = 20 * b2.ms
Rm = 100 * b2.Mohm
abs_refractory_period = 2 * b2.ms

if n_inh is None:
    n_inh = int(n_exc / 4)
N_tot = n_exc + n_inh
if n_ext is None:
    n_ext = int(n_exc * connection_probability)
if w_ext is None:
    w_ext = w0

J_excit = w0
J_inhib = -g * w0

lif_dynamics = """
    dv/dt = (-(v-v_rest) + Rm*(I_exc + I_ext + I_gaba)) / membrane_time_scale : volt,
↪(unless refractory)
    I_gaba : amp
    I_exc : amp
    I_ext : amp
"""

neurons = b2.NeuronGroup(
    N_tot,
    model=lif_dynamics,
    threshold="v>firing_threshold",
    reset="v=v_reset",

```

(continues on next page)

(continued from previous page)

```

    refractory=abs_refractory_period,
    method="linear",
)
neurons.v = (
    np.random.uniform(
        v_rest / b2.mV, high=firing_threshold / b2.mV, size=(n_exc + n_inh)
    )
    * b2.mV
)
cleo.coords.assign_coords_rand_cylinder(
    neurons, (0, 0, 700), (0, 0, 900), 250, unit=b2.um
)

exc = neurons[:n_exc]
inh = neurons[n_exc:]

syn_eqs = """
    dI_syn_syn/dt = (s - I_syn_syn)/tau_dsyn : amp (clock-driven)
    I_TYPE_post = I_syn_syn : amp (summed)
    ds/dt = -s/tau_rsyn : amp (clock-driven)
"""

exc_synapses = b2.Synapses(
    exc,
    target=neurons,
    model=syn_eqs.replace("TYPE", "exc"),
    on_pre="s += J_excit",
    delay=synaptic_delay,
    namespace={"tau_rsyn": 0.4 * b2.ms, "tau_dsyn": 2 * b2.ms},
)
exc_synapses.connect(p=connection_probability)

inh_synapses = b2.Synapses(
    inh,
    target=neurons,
    model=syn_eqs.replace("TYPE", "gaba"),
    on_pre="s += J_inhib",
    delay=synaptic_delay,
    namespace={"tau_rsyn": 0.25 * b2.ms, "tau_dsyn": 5 * b2.ms},
)
inh_synapses.connect(p=connection_probability)

ext_input = b2.PoissonGroup(n_ext, poisson_input_rate, name="ext_input")
ext_synapses = b2.Synapses(
    ext_input,
    target=neurons,
    model=syn_eqs.replace("TYPE", "ext"),
    on_pre="s += w_ext",
    delay=synaptic_delay,
    namespace={"tau_rsyn": 0.4 * b2.ms, "tau_dsyn": 2 * b2.ms},
)
ext_synapses.connect(p=connection_probability)

```

(continues on next page)

(continued from previous page)

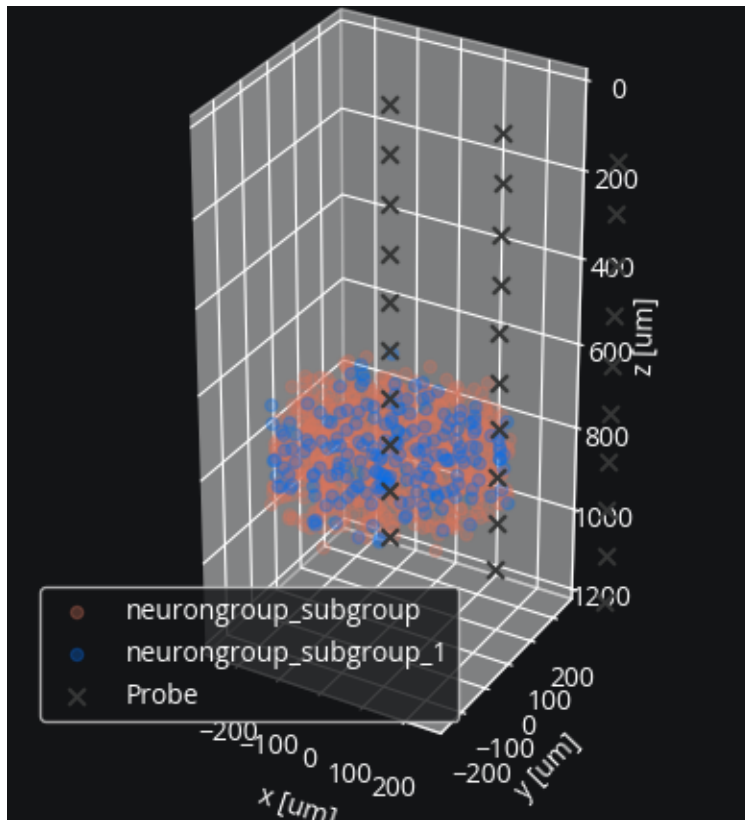
```
net = b2.Network(  
    neurons,  
    exc_synapses,  
    inh_synapses,  
    ext_input,  
    ext_synapses,  
)  
sim = cleo.CLSimulator(net)
```

## Electrode setup

```
elec_coords = cleo.ephys.linear_shank_coords(1 * b2.mm, channel_count=10)  
elec_coords = cleo.ephys.tile_coords(  
    elec_coords, num_tiles=3, tile_vector=(500, 0, 0) * b2.um  
)  
probe = cleo.ephys.Probe(elec_coords)
```

```
cleo.viz.plot(  
    exc,  
    inh,  
    colors=[c["exc"], c["inh"]],  
    zlim=(0, 1200),  
    devices=[probe],  
    scatterargs={"alpha": 0.3},  
);
```





```
mua = ephys.MultiUnitSpiking(
    r_perfect_detection=0.05 * b2.mm,
    r_half_detection=0.1 * b2.mm,
)
ss = ephys.SortedSpiking(0.05 * b2.mm, 0.1 * b2.mm)
tklfp = ephys.TKLFPSignal()
```

There are a few important variations on how to record RWSLFP:

- Currents can be summed over the population, so that a postsynaptic current (PSC) in one location has the same effect on LFP as one on the other side of the population. The main advantage to this approach is it saves some memory storing currents. To use this, set `pop_aggregate` to True. You'll also want to set `amp_func` to `wslfp.mazzoni15_pop` to get the population amplitude profile from Mazzoni et al., 2015. The default is to not sum over the population, and use `wslfp.mazzoni15_nrn` to get per-neuron contributions to LFP instead.
- The LFP can be computed from PSCs if your model simulates them or from spikes (after synthesizing PSCs from them). Use `RWSLFPSignalFromSpikes` or `RWSLFPSignalFromPSCs` accordingly. In this example, we are simulating synaptic dynamics in the form of biexponential currents, which happens to be the form used to generate synthetic PSCs.

`RWSLFPSignalFromSpikes` needs to know about all spikes transmitted to the population being recorded from, so `ampa_syns` and `gaba_syns` must be passed on injection. To account for the relative impact of incoming spikes on synaptic currents, Cleo needs to be able to find the weight as well. It looks for a variable or parameter named `w` in the synapses by default, but you can pass in an alternate name or a value on injection instead.

`RWSLFPSignalFromSpikes` has sensible defaults, but can be overridden with the exact parameters used in our model. These are used in the synthetic current generation process. These parameters then serve as the default for the signal, which can be overridden on a per-injection basis.

```

import wslfp

rwsلفp_spk_nrn = ephys.RWSLFPSignalFromSpikes(
    tau1_ampa=exc_synapses.namespace["tau_dsyn"],
    tau2_ampa=exc_synapses.namespace["tau_rsyn"],
    tau1_gaba=inh_synapses.namespace["tau_dsyn"],
    tau2_gaba=inh_synapses.namespace["tau_rsyn"],
    syn_delay=synaptic_delay,
    name="per-neuron RWSLFP from spikes",
)

rwsلفp_spk_pop = ephys.RWSLFPSignalFromSpikes(
    pop_aggregate=True,
    amp_func=wslfp.mazzoni15_pop,
    name="population-aggregated RWSLFP from spikes",
)

```

All that's needed for *RWSLFPSignalFromPSCs* is *Iampa\_var\_names* and *Igaba\_var\_names* on injection, which are lists of the variables representing AMPA and GABA currents.

```

rwsلفp_psc_nrn = ephys.RWSLFPSignalFromPSCs(name="per-neuron RWSLFP from PSCs")
rwsلفp_psc_pop = ephys.RWSLFPSignalFromPSCs(
    pop_aggregate=True,
    amp_func=wslfp.mazzoni15_pop,
    name="population-aggregated RWSLFP from PSCs",
)

```

All signals are grouped together on the probe, but we can avoid RWSLFP being recorded from interneurons by omitting *ampa\_syns*, *gaba\_syns*, *Iampa\_var\_names*, and *Igaba\_var\_names* from the injection.

```

probe.add_signals(
    mua,
    ss,
    tkلفp,
    rwsلفp_spk_nrn,
    rwsلفp_spk_pop,
    rwsلفp_psc_nrn,
    rwsلفp_psc_pop,
)

sim.set_io_processor(cleo.ioproc.RecordOnlyProcessor(sample_period_ms=1))
sim.inject(
    probe,
    exc,
    # for TKLFPSignal:
    tkلفp_type="exc",
    # for RWSLFPSignalFromSpikes:
    synaptic_delay=synaptic_delay, # can override for all synapses
    ampa_syns=[ # or per synapse by with (syn, kwargs) tuples:
        # want only synapses onto pyramidal cells
        (exc_synapses[f"j < {n_exc}"], {"weight": J_excit}),
        (ext_synapses[f"j < {n_exc}"], {"weight": w_ext}),
    ],
)

```

(continues on next page)

(continued from previous page)

```

gaba_syns=[(inh_synapses[f"j < {n_exc}"], {"weight": J_inhib})],
# for RWSLFPSignalFromPSCs:
Iampa_var_names=["I_exc", "I_ext"],
Igaba_var_names=["I_gaba"],
)
# we don't include ampa_syns, gaba_syns, Iampa_var_name, or Igaba_var_name since RWSLFP
# is only recorded from pyramidal cells
sim.inject(probe, inh, tklf_type="inh")

```

```

CLSimulator(io_processor=RecordOnlyProcessor(sample_period_ms=1, sampling='fixed',
→processing='parallel'), devices={Probe(name='Probe', save_history=True,
→signals=[MultiUnitSpiking(name='MultiUnitSpiking', brian_objects={<SpikeMonitor,
→recording from 'spikemonitor'>, <SpikeMonitor, recording from 'spikemonitor_9'>},
→probe=..., r_perfect_detection=50. * umetre, r_half_detection=100. * umetre, cutoff_
→probability=0.01), SortedSpiking(name='SortedSpiking', brian_objects={<SpikeMonitor,
→recording from 'spikemonitor_1'>, <SpikeMonitor, recording from 'spikemonitor_10'>},
→probe=..., r_perfect_detection=50. * umetre, r_half_detection=100. * umetre, cutoff_
→probability=0.01), TKLFPSignal(name='TKLFPSignal', brian_objects={<SpikeMonitor,
→recording from 'spikemonitor_11'>, <SpikeMonitor, recording from 'spikemonitor_2'>},
→probe=..., uLFP_threshold_uV=0.001, _lfp_unit=uvolt), RWSLFPSignalFromSpikes(name='per-
→neuron RWSLFP from spikes', brian_objects={<SpikeMonitor, recording from 'spikemonitor_
→4'>, <SpikeMonitor, recording from 'spikemonitor_5'>, <SpikeMonitor, recording from
→'spikemonitor_3'>}, probe=..., amp_func=<function mazzoni15_nrn at 0x7f80f62c99e0>,
→pop_aggregate=False, _lfp_unit=1, tau1_ampa=2. * msecond, tau2_ampa=0.4 * msecond,
→tau1_gaba=5. * msecond, tau2_gaba=250. * usecond, syn_delay=1. * msecond, I_
→threshold=0.001, weight='w'), RWSLFPSignalFromSpikes(name='population-aggregated
→RWSLFP from spikes', brian_objects={<SpikeMonitor, recording from 'spikemonitor_7'>,
→<SpikeMonitor, recording from 'spikemonitor_8'>, <SpikeMonitor, recording from
→'spikemonitor_6'>}, probe=..., amp_func=<function mazzoni15_pop at 0x7f80f62c9940>,
→pop_aggregate=True, _lfp_unit=1, tau1_ampa=2. * msecond, tau2_ampa=0.4 * msecond, tau1_
→gaba=5. * msecond, tau2_gaba=250. * usecond, syn_delay=1. * msecond, I_threshold=0.001,
→weight='w'), RWSLFPSignalFromPSCs(name='per-neuron RWSLFP from PSCs', brian_
→objects=set(), probe=..., amp_func=<function mazzoni15_nrn at 0x7f80f62c99e0>, pop_
→aggregate=False, _lfp_unit=1), RWSLFPSignalFromPSCs(name='population-aggregated RWSLFP
→from PSCs', brian_objects=set(), probe=..., amp_func=<function mazzoni15_pop at
→0x7f80f62c9940>, pop_aggregate=True, _lfp_unit=1)], probe=NOTHING)})

```

```

sim.reset()
sim.run(500 * b2.ms)

```

```

INFO      No numerical integration method specified for group 'synapses_1', using_
→method 'exact' (took 0.21s). [brian2.stateupdaters.base.method_choice]
INFO      No numerical integration method specified for group 'synapses_2', using_
→method 'exact' (took 0.10s). [brian2.stateupdaters.base.method_choice]
INFO      No numerical integration method specified for group 'synapses', using method
→'exact' (took 0.00s). [brian2.stateupdaters.base.method_choice]

```

```

from matplotlib.colors import ListedColormap

fig, axs = plt.subplots(2, 1, sharex=True)

```

(continues on next page)

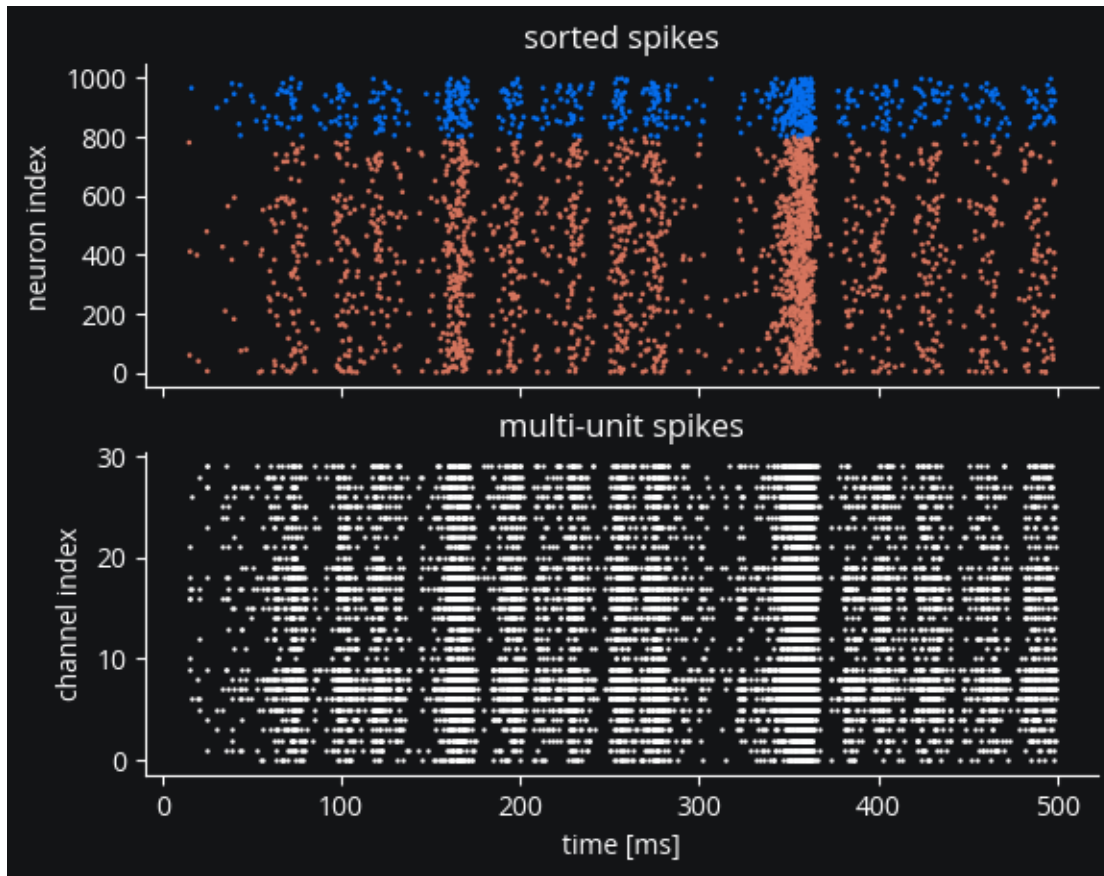
(continued from previous page)

```

# assuming all neurons are detectable for c=ss.i >= n_e to work
# in practice this will often not be the case and we'd have to map
# from probe index to neuron group index using ss.i_probe_by_i_ng.inverse
exc_inh_cmap = ListedColormap([c["exc"], c["inh"]])
axs[0].scatter(ss.t_ms, ss.i, marker=".", c=ss.i >= n_exc, cmap=exc_inh_cmap, s=3)
axs[0].set(title="sorted spikes", ylabel="neuron index")

axs[1].scatter(mua.t_ms, mua.i, marker=".", s=2, c="white")
axs[1].set(title="multi-unit spikes", ylabel="channel index", xlabel="time [ms]");

```



```

def plot_lfp(t_ms, lfp, title=None):
    n_shanks = 3
    n_contacts_per_shank = 10
    fig, axs = plt.subplots(1, n_shanks, sharey=True, figsize=(8, 4))
    for i, color, r_rec, ax in zip(
        range(n_shanks), [c["light"], c["main"], c["dark"]], [0, 250, 500], axs
    ):
        lfp_for_shank = lfp[
            :, i * n_contacts_per_shank : (i + 1) * n_contacts_per_shank
        ]
        ax.plot(
            t_ms,
            lfp_for_shank + np.arange(n_contacts_per_shank) * 1.1 * np.abs(lfp.max()),
            c=color,

```

(continues on next page)

(continued from previous page)

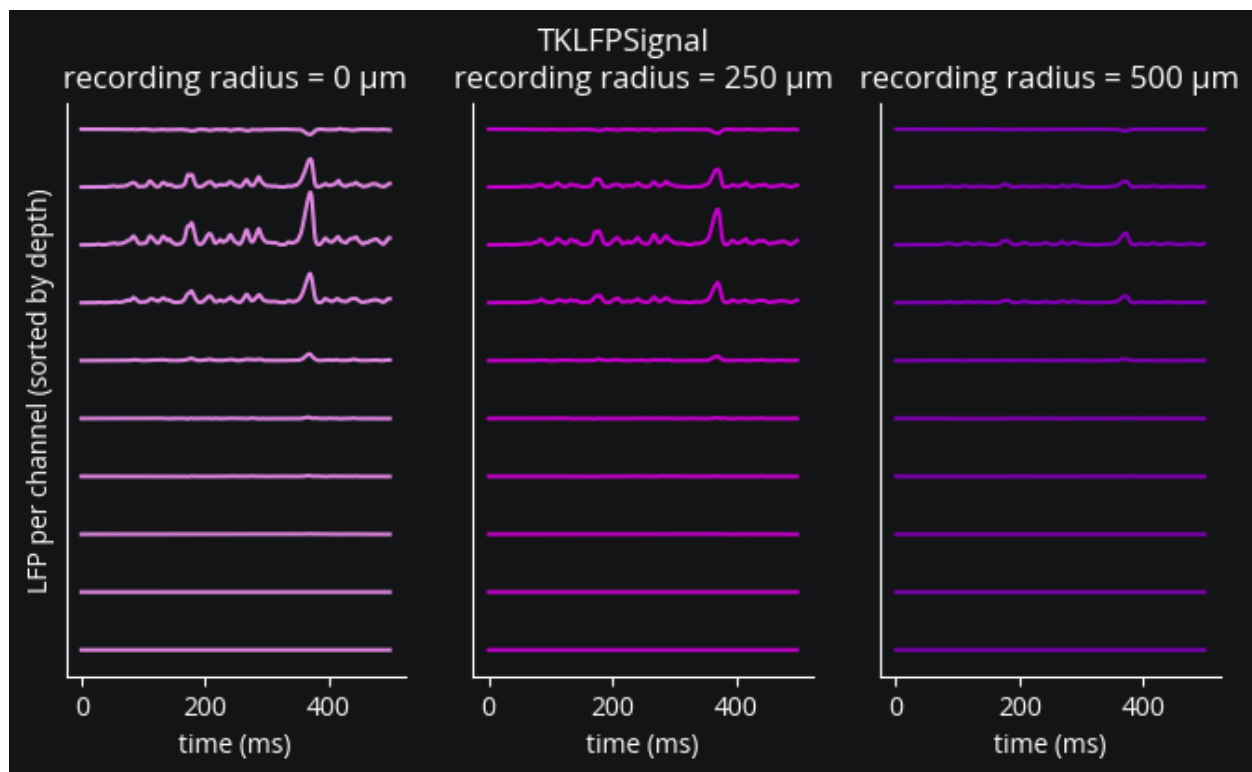
```

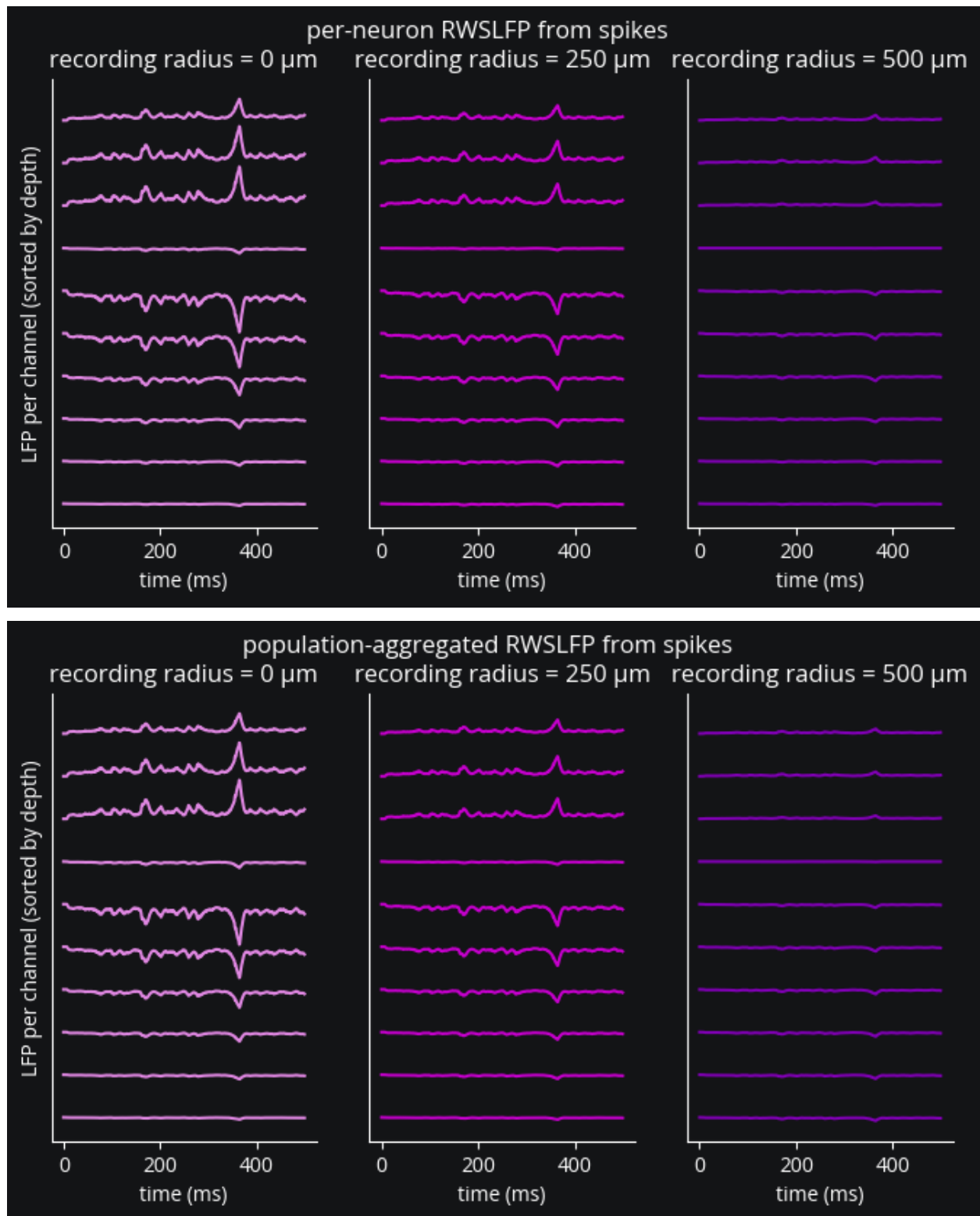
    )
    ax.set(xlabel="time (ms)", yticks=[], title=f"recording radius = {r_rec}  $\mu\text{m}$ ")

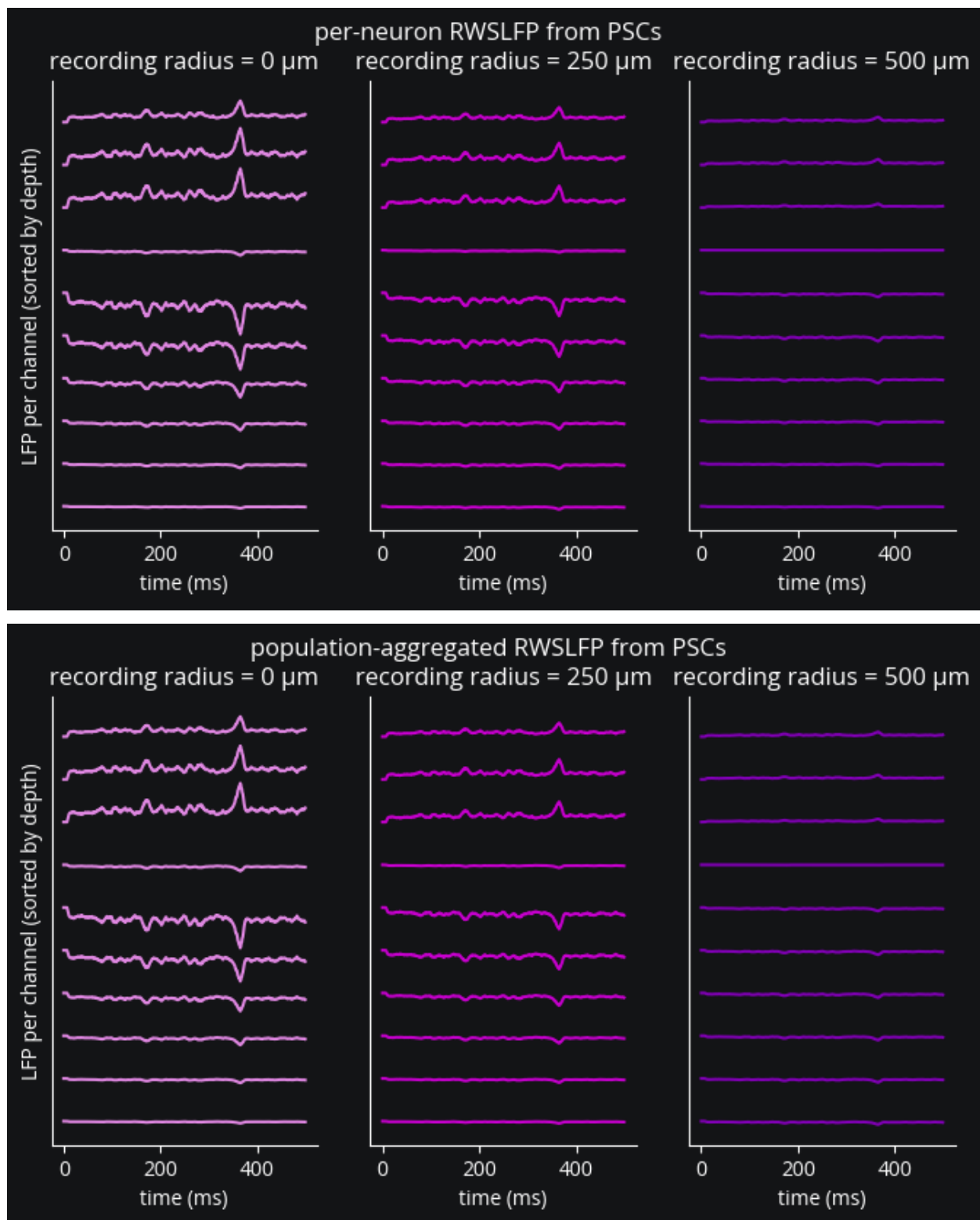
    axs[0].set(ylabel="LFP per channel (sorted by depth)")
    if title:
        fig.suptitle(title)

for signal in probe.signals:
    if isinstance(
        signal,
        (ephys.TKLFPSignal, ephys.RWSLFPSignalFromPSCs, ephys.RWSLFPSignalFromSpikes),
    ):
        lfp = signal.lfp
    else:
        continue
    plot_lfp(signal.t_ms, lfp, title=signal.name)

```







Despite using all the same parameters for the postsynaptic current curves, the `FromSpikes` and `FromPSCs` signals look different. In fact, what we see above matches the [WSLFP demo](#), where the spike convolution signal is somewhat spikier (i.e., with higher peaks the jump at the beginning looks smaller).



## 6.3 Reference

### 6.3.1 cleo module

Contains core classes and functions for the Cleo package.

**class** `cleo.CLSimulator`(*network*: *Network*)

Bases: *NeoExportable*

The centerpiece of cleo. Integrates simulation components and runs.

Method generated by attrs for class CLSimulator.

**devices:** `set[InterfaceDevice]`

**get\_state()** → dict

Return current recorder measurements.

**Returns**

A dictionary of *name: state* pairs for all recorders in the simulator.

**Return type**

dict

**inject**(*device*: *InterfaceDevice*, *\*neuron\_groups*: *NeuronGroup*, *\*\*kwargs*: *Any*) → *CLSimulator*

Inject InterfaceDevice into the network, connecting to specified neurons.

Calls `connect_to_neuron_group()` for each group with *kwargs* and adds the device's *brian\_objects* to the simulator's *network*.

**Parameters**

**device** (*InterfaceDevice*) – Device to inject

**Returns**

self

**Return type**

*CLSimulator*

**io\_processor:** *IOProcessor*

**network:** *Network*

The Brian network forming the core model

**recorders:** `dict[str, Recorder]`

**reset**(*\*\*kwargs*)

Reset the simulator to a neutral state

Restores the Brian Network to where it was when the CLSimulator was last modified (last injection, IO-Processor change). Calls `reset()` on *devices* and *io\_processor*.

**run**(*duration*: *Quantity*, *\*\*kwargs*) → None

Run simulation.

**Parameters**

- **duration** (*brian2 temporal Quantity*) – Length of simulation
- **\*\*kwargs** (*additional arguments passed to brian2.run()*) – level has a default value of 1



**set\_io\_processor**(*io\_processor*: [IOProcessor](#), *communication\_period*=None) → [CLSimulator](#)

Set simulator IO processor

Will replace any previous IOProcessor so there is only one at a time. A Brian NetworkOperation is created to govern communication between the Network and the IOProcessor.

**Parameters**

**io\_processor** ([IOProcessor](#)) –

**Returns**

self

**Return type**

[CLSimulator](#)

**stimulators**: dict[str, [Stimulator](#)]

**to\_neo**() → [Block](#)

Exports simulator data to a Neo Block

**Returns**

Neo Block containing signals representing each device's data

**Return type**

[neo.core.Block](#)

**update\_stimulators**(*stim\_values*: dict[str, Any]) → None

Update stimulators with output from the [IOProcessor](#)

**Parameters**

**stim\_values** (dict) – {*stimulator\_name*: *stim\_value*} dictionary with values to update each stimulator.

**class** [cleo.IOProcessor](#)(*sample\_period\_ms*: float = 1)

Bases: ABC

Abstract class for implementing sampling, signal processing and control

This must be implemented by the user with their desired closed-loop use case, though most users will find the [LatencyIOProcessor\(\)](#) class more useful, since delay handling is already defined.

Method generated by attrs for class IOProcessor.

**abstract** **get\_ctrl\_signals**(*query\_time\_ms*: float) → dict

Get per-stimulator control signal from the [IOProcessor](#).

**Parameters**

**query\_time\_ms** (float) – Current simulation time.

**Returns**

A {'stimulator\_name': ctrl\_signal} dictionary for updating stimulators.

**Return type**

dict

**get\_intersample\_ctrl\_signal**(*query\_time\_ms*: float) → dict

Get per-stimulator control signal between samples. I.e., for implementing a time-varying waveform based on parameters from the last sample. Such parameters will need to be stored in the [IOProcessor](#).

**get\_stim\_values**(*query\_time\_ms*: float) → dict

**abstract is\_sampling\_now**(*time*) → bool

Determines whether the processor will take a sample at this timestep.

**Parameters**

**time** (*Brian 2 temporal Unit*) – Current timestep.

**Return type**

bool

**latest\_ctrl\_signal:** dict

The most recent control signal returned by [get\\_ctrl\\_signals\(\)](#)

**preprocess\_ctrl\_signals**(*latest\_ctrl\_signals: dict, query\_time\_ms: float*) → dict

Preprocess control signals as needed to control stimulator waveforms between samples.

I.e., if a control signal defines the frequency of a periodic light stimulus, this function computes the current intensity given the latest frequency and the current time. This is called immediately after [get\\_ctrl\\_signals\(\)](#) and on every timestep to update the stimulator waveform between samples.

This only needs to be implemented when a stimulus that varies between samples is desired. Otherwise, the control signal returned by [get\\_ctrl\\_signals\(\)](#) is used directly. If not all stimulators need this functionality, only return a dict for those that do. The original, unprocessed control signal is used for the others.

**Parameters**

**query\_time\_ms** (*float*) – Current simulation time.

**Returns**

A {'stimulator\_name': value} dictionary for updating stimulators.

**Return type**

dict

**abstract put\_state**(*state\_dict: dict, sample\_time\_ms: float*) → None

Deliver network state to the [IOProcessor](#).

**Parameters**

- **state\_dict** (*dict*) – A dictionary of recorder measurements, as returned by [get\\_state\(\)](#)
- **sample\_time\_ms** (*float*) – The current simulation timestep. Essential for simulating control latency and for time-varying control.

**reset**(*\*\*kwargs*) → None

**sample\_period\_ms:** float

Determines how frequently the processor takes samples

**class** cleo.**InterfaceDevice**(*\*, name: str = \_Nothing.NOTHING, save\_history: bool = True*)

Bases: ABC

Base class for devices to be injected into the network

Method generated by attrs for class InterfaceDevice.

**add\_self\_to\_plot**(*ax: Axes3D, axis\_scale\_unit: Unit, \*\*kwargs*) → list[Artist]

Add device to an existing plot

Should only be called by [plot\(\)](#).

**Parameters**

- **ax** (*Axes3D*) – The existing matplotlib Axes object

- **axis\_scale\_unit** (*Unit*) – The unit used to label axes and define chart limits
- **\*\*kwargs** (*optional*) –

**Returns**

A list of artists used to render the device. Needed for use in conjunction with [VideoVisualizer](#).

**Return type**

list[Artist]

**brian\_objects: set**

All the Brian objects added to the network by this device. Must be kept up-to-date in [connect\\_to\\_neuron\\_group\(\)](#) and other functions so that those objects can be automatically added to the network when the device is injected.

**abstract connect\_to\_neuron\_group**(*neuron\_group: NeuronGroup*, *\*\*kwargs*) → None

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a NeuronGroup, Synapses, or Monitor, make sure to add these to [brian\\_objects](#).

**Parameters**

- **neuron\_group** (*NeuronGroup*) –
- **\*\*kwargs** (*optional*) – Passed from *inject*

**init\_for\_simulator**(*simulator: CLSimulator*) → None

Initialize device for simulator on initial injection.

This function is called only the first time a device is injected into a simulator and performs any operations that are independent of the individual neuron groups it is connected to.

**Parameters**

**simulator** (*CLSimulator*) – simulator being injected into

**name: str**

Identifier for device, used in sampling, plotting, etc. Name of the class by default. Must be unique among recorders and stimulators

**reset**(*\*\*kwargs*) → None

Reset the device to a neutral state

**save\_history: bool**

Determines whether times and inputs/outputs are recorded. True by default.

For stimulators, this is when [update\(\)](#) is called. For recorders, it is when [get\\_state\(\)](#) is called.

**sim: CLSimulator**

The simulator the device is injected into

**update\_artists**(*artists: list[Artist]*, *\*args*, *\*\*kwargs*) → list[Artist]

Update the artists used to render the device

Used to set the artists' state at every frame of a video visualization. The current state would be passed in *\*args* or *\*\*kwargs*

**Parameters**

**artists** (*list[Artist]*) – the artists used to render the device originally, i.e., which were returned from the first [add\\_self\\_to\\_plot\(\)](#) call.

**Returns**

The artists that were actually updated. Needed for efficient blit rendering, where only updated artists are re-rendered.

**Return type**

list[Artist]

**class cleo.NeoExportable**

Bases: ABC

Mixin class for classes that can be exported to Neo objects

**abstract to\_neo()** → neo.core.BaseNeo

Return a Neo signal object with the device's data

**Returns**

Neo object representing exported data

**Return type**

neo.core.BaseNeo

**class cleo.Recorder**(\*, name: str = \_Nothing.NOTHING, save\_history: bool = True)

Bases: [InterfaceDevice](#)

Device for taking measurements of the network.

Method generated by attrs for class Recorder.

**abstract get\_state()** → Any

Return current measurement.

**class cleo.Stimulator**(default\_value: Any = 0, \*, name: str = \_Nothing.NOTHING, save\_history: bool = True)

Bases: [InterfaceDevice](#), [NeoExportable](#)

Device for manipulating the network

Method generated by attrs for class Stimulator.

**default\_value: Any**

The default value of the device—used on initialization and on [reset\(\)](#)

**reset(\*\*kwargs)** → None

Reset the stimulator device to a neutral state

**t\_ms: list[float]**

Times stimulator was updated, stored if [save\\_history](#)

**to\_neo()**

Return a Neo signal object with the device's data

**Returns**

Neo object representing exported data

**Return type**

neo.core.BaseNeo

**update(ctrl\_signal)** → None

Set the stimulator value.

By default this sets [value](#) to ctrl\_signal and updates saved times and values. You will want to implement this method if your stimulator requires additional logic. Use `super.update(self, value)` to preserve the `self.value` and `save_history` logic

**Parameters**

**ctrl\_signal** (*any*) – The value the stimulator is to take.

**value:** **Any**

The current value of the stimulator device

**values:** **list[Any]**

Values taken by the stimulator at each [update\(\)](#) call, stored if [save\\_history](#)

**class** cleo.SynapseDevice(*extra\_namespace: dict = \_Nothing.NOTHING, \*, name: str = \_Nothing.NOTHING, save\_history: bool = True*)

Bases: [InterfaceDevice](#)

Base class for devices that record from/stimulate neurons via a Synapses object with device-specific model. Used for opsin and indicator classes

Method generated by attrs for class SynapseDevice.

**connect\_to\_neuron\_group**(*neuron\_group: NeuronGroup, \*\*kwparams*) → None

Transfect neuron group with device.

**Parameters**

**neuron\_group** (*NeuronGroup*) – The neuron group to transform

**Keyword Arguments**

- **p\_expression** (*float*) – Probability ( $0 \leq p \leq 1$ ) that a given neuron in the group will express the protein. 1 by default.
- **i\_targets** (*array-like*) – Indices of neurons in the group to transfect. recommended for efficiency when stimulating or imaging a small subset of the group. Incompatible with **p\_expression**.
- **rho\_rel** (*float*) – The expression level, relative to the standard model fit, of the protein. 1 by default. For heterogeneous expression, this would have to be modified in the light-dependent synapse post-injection, e.g., `opsin.syns["neuron_group_name"].rho_rel = ...`
- **[default\_name]\_var\_name** (*str*) – See [required\\_vars](#). Allows for custom variable names.

**extra\_namespace:** **dict**

Additional items (beyond parameters) to be added to the opto synapse namespace

**init\_syn\_vars**(*syn: Synapses*) → None

Initializes appropriate variables in Synapses implementing the model

Also called on [reset\(\)](#).

**Parameters**

**syn** (*Synapses*) – The synapses object implementing this model

**model:** **str**

Basic Brian model equations string.

Should contain a *rho\_rel* term reflecting relative expression levels. Will likely also contain special NeuronGroup-dependent symbols such as `V_VAR_NAME` to be replaced on injection in [modify\\_model\\_and\\_params\\_for\\_ng\(\)](#).

**modify\_model\_and\_params\_for\_ng**(*neuron\_group*: *NeuronGroup*, *injt\_params*: *dict*) →  
 Tuple[Equations, dict]

Adapt model for given neuron group on injection

This enables the specification of variable names differently for each neuron group, allowing for custom names and avoiding conflicts.

#### Parameters

- **neuron\_group** (*NeuronGroup*) – NeuronGroup this opsin model is being connected to
- **injt\_params** (*dict*) – kwargs passed in on injection, could contain variable names to plug into the model

#### Keyword Arguments

**model** (*str*, *optional*) – Model to start with, by default that defined for the class. This allows for prior string manipulations before it can be parsed as an *Equations* object.

#### Returns

A tuple containing an *Equations* object and a parameter dictionary, constructed from *model* and *params*, respectively, with modified names for use in *synapses*

#### Return type

Equations, dict

**on\_pre:** *str*

Model string for *brian2.synapses.synapses.Synapses* reacting to spikes.

**property params:** *dict*

Returns a dictionary of parameters for the model

**per\_ng\_unit\_replacements:** *list*[*Tuple*[*str*, *str*]]

List of (UNIT\_NAME, neuron\_group\_specific\_unit\_name) tuples to be substituted in the model string on injection and before checking required variables.

**required\_vars:** *list*[*Tuple*[*str*, *Unit*]]

Default names of state variables required in the neuron group, along with units, e.g., [(‘Iopto’, amp)].

It is assumed that non-default values can be passed in on injection as a keyword argument [default\_name]\_var\_name=[non\_default\_name] and that these are found in the model string as [DEFAULT\_NAME]\_VAR\_NAME before replacement.

**reset**(\*\**kwargs*)

Reset the device to a neutral state

**source\_ngs:** *dict*[*str*, *NeuronGroup*]

{target\_ng.name: source\_ng} dict of source neuron groups.

The source is the target itself by default or light aggregator neurons for *LightDependent*.

**synapses:** *dict*[*str*, *Synapses*]

Stores the synapse objects implementing the model, connecting from source (light aggregator neurons or the target group itself) to target neuron groups, of form {target\_ng.name: synapses}.

### 6.3.2 cleo.coords module

Contains functions for assigning neuron coordinates and visualizing

`cleo.coords.assign_coords(neuron_group: NeuronGroup, coords: Quantity)`

Assigns x, y, and z coordinates for neuron group from (n, 3) array

`cleo.coords.assign_coords_grid_rect_prism(neuron_group: NeuronGroup, xlim: Tuple[float, float], ylim: Tuple[float, float], zlim: Tuple[float, float], shape: Tuple[int, int, int], unit: Unit = mmetre) → None`

Assign grid coordinates to neurons in a rectangular grid

#### Parameters

- **neuron\_group** (*NeuronGroup*) – The neuron group to assign coordinates to
- **xlim** (*Tuple*[float, float]) – xmin, xmax, with no unit
- **ylim** (*Tuple*[float, float]) – ymin, ymax, with no unit
- **zlim** (*Tuple*[float, float]) – zmin, zmax with no unit
- **shape** (*Tuple*[int, int, int]) – n\_x, n\_y, n\_z tuple representing the shape of the resulting grid
- **unit** (*Unit*, optional) – Brian unit determining what scale to use for coordinates, by default mm

#### Raises

**ValueError** – When the shape is incompatible with the number of neurons in the group

`cleo.coords.assign_coords_rand_cylinder(neuron_group: NeuronGroup, xyz_start: Tuple[float, float, float], xyz_end: Tuple[float, float, float], radius: float, unit: Unit = mmetre) → None`

Assign random coordinates within a cylinder.

#### Parameters

- **neuron\_group** (*NeuronGroup*) – neurons to assign coordinates to
- **xyz\_start** (*Tuple*[float, float, float]) – starting position of cylinder without unit
- **xyz\_end** (*Tuple*[float, float, float]) – ending position of cylinder without unit
- **radius** (float) – radius of cylinder without unit
- **unit** (*Unit*, optional) – Brian unit to scale other params, by default mm

`cleo.coords.assign_coords_rand_rect_prism(neuron_group: NeuronGroup, xlim: Tuple[float, float], ylim: Tuple[float, float], zlim: Tuple[float, float], unit: Unit = mmetre) → None`

Assign random coordinates to neurons within a rectangular prism

#### Parameters

- **neuron\_group** (*NeuronGroup*) – neurons to assign coordinates to
- **xlim** (*Tuple*[float, float]) – xmin, xmax without unit
- **ylim** (*Tuple*[float, float]) – ymin, ymax without unit
- **zlim** (*Tuple*[float, float]) – zmin, zmax without unit
- **unit** (*Unit*, optional) – Brian unit to specify scale implied in limits, by default mm

`cleo.coords.assign_coords_uniform_cylinder(neuron_group: NeuronGroup, xyz_start: Tuple[float, float, float], xyz_end: Tuple[float, float, float], radius: float, unit: Unit = mmetre) → None`

Assign uniformly spaced coordinates within a cylinder.

**Parameters**

- **neuron\_group** (*NeuronGroup*) – neurons to assign coordinates to
- **xyz\_start** (*Tuple*[float, float, float]) – starting position of cylinder without unit
- **xyz\_end** (*Tuple*[float, float, float]) – ending position of cylinder without unit
- **radius** (float) – radius of cylinder without unit
- **unit** (*Unit*, optional) – Brian unit to scale other params, by default mm

`cleo.coords.assign_xyz(neuron_group: NeuronGroup, x: ndarray, y: ndarray, z: ndarray, unit: Unit = mmetre)`

Assign arbitrary coordinates to neuron group.

**Parameters**

- **neuron\_group** (*NeuronGroup*) – neurons to be assigned coordinates
- **x** (*np.ndarray*) – x positions to assign (preferably 1D with no unit)
- **y** (*np.ndarray*) – y positions to assign (preferably 1D with no unit)
- **z** (*np.ndarray*) – z positions to assign (preferably 1D with no unit)
- **unit** (*Unit*, optional) – Brian unit determining what scale to use for coordinates, by default mm

`cleo.coords.concat_coords(*coords: Quantity) → Quantity`

Combine multiple coordinate *Quantity* arrays into one

**Parameters**

- **\*coords** (*Quantity*) – Multiple coordinate n x 3 *Quantity* arrays to combine

**Returns**

A single n x 3 combined *Quantity* array

**Return type**

*Quantity*

`cleo.coords.coords_from_ng(ng: NeuronGroup) → Quantity`

Get (n, 3) coordinate array from *NeuronGroup*.

`cleo.coords.coords_from_xyz(x: Quantity, y: Quantity, z: Quantity) → Quantity`

Create (... , 3) coordinate array from x, y, z arrays (with units).



### 6.3.3 cleo.ephys module

Contains probes, convenience functions for generating electrode array coordinates, signals, spiking, and LFP

**class** cleo.ephys.LFPSignalBase(\*, name: str = \_Nothing.NOTHING)

Bases: [Signal](#), [NeoExportable](#)

Base class for LFP Signals.

**Injection kwargs**

**orientation** (*np.ndarray, optional*) – Array of shape (n\_neurons, 3) representing which way is “up,” that is, towards the surface of the cortex, for each neuron. If a single vector is given, it is taken to be the orientation for all neurons in the group. [0, 0, -1] is the default, meaning the negative z axis is “up.”

Method generated by attrs for class Signal.

**brian\_objects:** set

All Brian objects created by the signal. Must be kept up-to-date for automatic injection into the network

**lfp:** NDArray[Any, Any, Quantity] = \_CountingAttr(counter=155, \_default=NOTHING, repr=False, eq=True, order=True, hash=None, init=False, on\_setattr=None, alias=None, metadata={})

Approximated LFP from every call to get\_state(). Shape is (n\_samples, n\_channels). Stored if [save\\_history](#) on [probe](#)

**name:** str

Unique identifier used to organize probe output. Name of the class by default.

**probe:** [Probe](#)

The probe the signal is configured to record for.

**t\_ms:** NDArray[Any, float] = \_CountingAttr(counter=154, \_default=NOTHING, repr=False, eq=True, order=True, hash=None, init=False, on\_setattr=None, alias=None, metadata={})

Times at which LFP is recorded, in ms, stored if [save\\_history](#) on [probe](#)

**to\_neo()** → [AnalogSignal](#)

Return a Neo signal object with the device’s data

**Returns**

Neo object representing exported data

**Return type**

neo.core.BaseNeo

**class** cleo.ephys.MultiUnitSpiking(*r\_perfect\_detection: Quantity, r\_half\_detection: Quantity, cutoff\_probability: float = 0.01, \*, name: str = \_Nothing.NOTHING*)

Bases: [Spiking](#)

Detects (unsorted) spikes per channel.

Method generated by attrs for class MultiUnitSpiking.

**connect\_to\_neuron\_group**(*neuron\_group: NeuronGroup, \*\*kwparams*) → None

Configure signal to record from specified neuron group

**Parameters**

**neuron\_group** (*NeuronGroup*) – group to record from

**get\_state()** → tuple[NDArray[None, UInt], NDArray[None, Float], NDArray[None, UInt]]

Return spikes since method was last called (i, t\_ms, y)

**Returns**

(i, t\_ms, y) where i is channel (for multi-unit) or neuron (for sorted) spike indices, t\_ms is spike times, and y is a spike count vector suitable for control- theoretic uses—i.e., a 0 for every channel/neuron that hasn’t spiked and a 1 for a single spike.

**Return type**

tuple[NDArray[np.uint], NDArray[float], NDArray[np.uint]]

**to\_neo()** → Group

Return a Neo signal object with the device’s data

**Returns**

Neo object representing exported data

**Return type**

neo.core.BaseNeo

**class** cleo.ephys.Probe(*coords*: Quantity, *signals*: list[Signal] = \_Nothing.NOTHING, \*, *name*: str = \_Nothing.NOTHING, *save\_history*: bool = True)

Bases: Recorder, NeoExportable

Picks up specified signals across an array of electrodes.

**Visualization kwargs**

- **marker** (*str*, *optional*) – The marker used to represent each contact. “x” by default.
- **size** (*float*, *optional*) – The size of each contact marker. 40 by default.
- **color** (*Any*, *optional*) – The color of contact markers. “xkcd:dark gray” by default.

Method generated by attrs for class Probe.

**add\_self\_to\_plot**(*ax*: Axes3D, *axis\_scale\_unit*: Unit, *\*\*kwargs*) → list[Artist]

Add device to an existing plot

Should only be called by *plot()*.

**Parameters**

- **ax** (Axes3D) – The existing matplotlib Axes object
- **axis\_scale\_unit** (Unit) – The unit used to label axes and define chart limits
- **\*\*kwargs** (*optional*) –

**Returns**

A list of artists used to render the device. Needed for use in conjunction with *VideoVisualizer*.

**Return type**

list[Artist]

**add\_signals**(*\*signals*: Signal) → None

Add signals to the probe for recording

**Parameters**

**\*signals** (Signal) – signals to add

**connect\_to\_neuron\_group**(*neuron\_group*: *NeuronGroup*, *\*\*kwargs*: *Any*) → None

Configure probe to record from given neuron group

Will call *Signal.connect\_to\_neuron\_group()* for each signal

**Parameters**

- **neuron\_group** (*NeuronGroup*) – neuron group to connect to, i.e., record from
- **\*\*kwargs** (*Any*) – Passed in to signals' connect functions, needed for some signals

**coords:** *Quantity*

Coordinates of n electrodes. Must be an n x 3 array (with unit) where columns represent x, y, and z

**get\_state()** → dict

Get current state from probe, i.e., all signals

**Returns**

{'signal\_name': value} dict with signal states

**Return type**

dict

**property n**

Number of electrode contacts in the probe

**probe:** *Probe*

**reset**(*\*\*kwargs*)

Reset the probe to a neutral state

Calls reset() on each signal

**signals:** list[*Signal*]

Signals recorded by the probe. Can be added to post-init with *add\_signals()*.

**to\_neo()** → *Group*

Return a Neo signal object with the device's data

**Returns**

Neo object representing exported data

**Return type**

neo.core.BaseNeo

**property xs:** *Quantity*

x coordinates of recording contacts

**Returns**

x coordinates represented as a Brian quantity, that is, including units. Should be like a 1D array.

**Return type**

*Quantity*

**property ys:** *Quantity*

y coordinates of recording contacts

**Returns**

y coordinates represented as a Brian quantity, that is, including units. Should be like a 1D array.

**Return type**

Quantity

**property zs:** [Quantity](#)

z coordinates of recording contacts

**Returns**

z coordinates represented as a Brian quantity, that is, including units. Should be like a 1D array.

**Return type**

Quantity

```
class cleo.ephys.RWSLFPSignalBase(amp_func: callable = <function mazzoni15_nrn>, pop_aggregate: bool
                                = False, lfp_unit: ~brian2.units.fundamentalunits.Unit = 1, *, name: str
                                = _Nothing.NOTHING)
```

Bases: [LFPSignalBase](#)Base class for [RWSLFPSignalFromSpikes](#) and [RWSLFPSignalFromPSCs](#).These signals should only be injected into neurons representing pyramidal cells with standard synaptic structure (see [Mazzoni, Lindén et al., 2015](#)).RWSLFP is computed using the [wslfp](#) package.

amp\_func and pop\_aggregate can be overridden on injection.

Method generated by attrs for class RWSLFPSignalBase.

**amp\_func:** **callable**Function to calculate LFP amplitudes, by default `wslfp.mazzoni15_nrn`.See [wslfp documentation](#) for more info.**get\_state()** → ndarray

Get the signal's current value

**pop\_aggregate:** **bool**

Whether to aggregate currents across the population (as opposed to neurons having differential contributions to LFP depending on their location). False by default.

**reset(\*\*kwargs)** → None

Reset signal to a neutral state

**to\_neo()** → [AnalogSignal](#)

Return a Neo signal object with the device's data

**Returns**

Neo object representing exported data

**Return type**

neo.core.BaseNeo

```
class cleo.ephys.RWSLFPSignalFromPSCs(amp_func: callable = <function mazzoni15_nrn>, pop_aggregate:
                                     bool = False, lfp_unit: ~brian2.units.fundamentalunits.Unit = 1, *,
                                     name: str = _Nothing.NOTHING)
```

Bases: [RWSLFPSignalBase](#)

Computes RWSLFP from the currents onto pyramidal cells.

Use this if your model already simulates synaptic current dynamics. `Iampa_var_names` and `Igaba_var_names` are lists of variable names to include and must be passed in as kwargs on injection or else the target neuron group will not contribute to this signal (desirable for interneurons).

RWSLFP refers to the Reference Weighted Sum of synaptic currents LFP proxy from [Mazzoni, Lindén et al., 2015](#).

#### Injection kwargs

- **Iampa\_var\_names** (*list[str]*) – List of variable names in the neuron group representing AMPA currents.
- **Igaba\_var\_names** (*list[str]*) – List of variable names in the neuron group representing GABA currents.

Method generated by attrs for class `RWSLFPSignalFromPSCs`.

**connect\_to\_neuron\_group**(*neuron\_group*: [NeuronGroup](#), *\*\*kwparams*)

Configure signal to record from specified neuron group

#### Parameters

**neuron\_group** ([NeuronGroup](#)) – group to record from

**reset**(*\*\*kwargs*) → None

Reset signal to a neutral state

```
class cleo.ephys.RWSLFPSignalFromSpikes(amp_func: callable = <function mazzoni15_nrn>,
                                         pop_aggregate: bool = False, lfp_unit:
                                         ~brian2.units.fundamentalunits.Unit = 1, tau1_ampa:
                                         ~brian2.units.fundamentalunits.Quantity = 2. * msecond,
                                         tau2_ampa: ~brian2.units.fundamentalunits.Quantity = 0.4 *
                                         msecond, tau1_gaba: ~brian2.units.fundamentalunits.Quantity
                                         = 5. * msecond, tau2_gaba:
                                         ~brian2.units.fundamentalunits.Quantity = 250. * usecond,
                                         syn_delay: ~brian2.units.fundamentalunits.Quantity = 1. *
                                         msecond, I_threshold: float = 0.001, weight: str = 'w', *, name:
                                         str = _Nothing.NOTHING)
```

Bases: [RWSLFPSignalBase](#)

Computes RWSLFP from the spikes onto pyramidal cell.

Use this if your model does not simulate synaptic current dynamics directly. The parameters of this class are used to synthesize biexponential synaptic currents using `wslfp.spikes_to_biexp_currents()`. `ampa_syms` and `gaba_syms` are lists of `Synapses` or `SynapticSubgroup` objects and must be passed as kwargs on injection, or else this signal will not be recorded for the target neurons (useful for ignoring interneurons). Attributes on the signal object serve as the default, but can be overridden on injection. Also, in the case that parameters (e.g., `tau1_ampa` or `weight`) vary by synapse, these can be overridden by passing a tuple of the `Synapses` or `SynapticSubgroup` object and a dictionary of the parameters to override.

RWSLFP refers to the Reference Weighted Sum of synaptic currents LFP proxy from [Mazzoni, Lindén et al., 2015](#).

#### Injection kwargs

- **ampa\_syms** (*list[Synapses | SynapticSubgroup | tuple[Synapses|SynapticSubgroup, dict]]*) – `Synapses` or `SynapticSubgroup` objects representing AMPA synapses (delivering excitatory currents). Or a tuple of the `Synapses` or `SynapticSubgroup` object and a dictionary of parameters to override.
- **gaba\_syms** (*list[Synapses | SynapticSubgroup | tuple[Synapses|SynapticSubgroup, dict]]*) – `Synapses` or `SynapticSubgroup` objects representing GABA synapses (delivering inhibitory

currents). Or a tuple of the Synapses or SynapticSubgroup object and a dictionary of parameters to override.

- **weight** (*str* | *float*, *optional*) – Name of the weight variable or parameter in the Synapses or SynapticSubgroup objects, or a float in the case of a single weight for all synapses. Default is ‘w’.

Method generated by attrs for class RWSLFPSignalFromSpikes.

**I\_threshold:** **float**

Threshold, as a proportion of the peak current, below which spikes’ contribution to synaptic currents (and thus LFP) is ignored, by default 1e-3.

**connect\_to\_neuron\_group**(*neuron\_group*: *NeuronGroup*, *\*\*kwparams*)

Configure signal to record from specified neuron group

**Parameters**

**neuron\_group** (*NeuronGroup*) – group to record from

**syn\_delay:** **Quantity**

The synaptic transmission delay, i.e., between a spike and the onset of the postsynaptic current. 1 ms by default.

**tau1\_ampa:** **Quantity**

The fall time constant of the biexponential current kernel for AMPA synapses. 2 ms by default.

**tau1\_gaba:** **Quantity**

The fall time constant of the biexponential current kernel for GABA synapses. 5 ms by default.

**tau2\_ampa:** **Quantity**

The time constant of subtracted part of the biexponential current kernel for AMPA synapses. 0.4 ms by default.

**tau2\_gaba:** **Quantity**

The time constant of subtracted part of the biexponential current kernel for GABA synapses. 0.25 ms by default.

**weight:** **str**

Name of the weight variable or parameter in the Synapses or SynapticSubgroup objects. Default is ‘w’.

**class** cleo.ephys.**Signal**(\*, *name*: *str* = *\_Nothing.NOTHING*)

Bases: ABC

Base class representing something an electrode can record

Method generated by attrs for class Signal.

**brian\_objects:** **set**

All Brian objects created by the signal. Must be kept up-to-date for automatic injection into the network

**abstract connect\_to\_neuron\_group**(*neuron\_group*: *NeuronGroup*, *\*\*kwparams*)

Configure signal to record from specified neuron group

**Parameters**

**neuron\_group** (*NeuronGroup*) – group to record from

**abstract get\_state**() → Any

Get the signal’s current value

**init\_for\_probe**(*probe*: [Probe](#)) → None

Called when attached to a probe.

Ensures signal can access probe and is only attached to one

**Parameters**

**probe** ([Probe](#)) – Probe to attach to

**Raises**

**ValueError** – When signal already attached to another probe

**name**: **str**

Unique identifier used to organize probe output. Name of the class by default.

**probe**: [Probe](#)

The probe the signal is configured to record for.

**reset**(\*\**kwargs*) → None

Reset signal to a neutral state

**class** cleo.ephys.**SortedSpiking**(*r\_perfect\_detection*: [Quantity](#), *r\_half\_detection*: [Quantity](#),  
*cutoff\_probability*: float = 0.01, \*, *name*: str = [\\_Nothing.NOTHING](#))

Bases: [Spiking](#)

Detect spikes identified by neuron indices.

The indices used by the probe do not correspond to those coming from neuron groups, since the probe must consider multiple potential groups and within a group ignores those neurons that are too far away to be easily detected.

Method generated by attrs for class SortedSpiking.

**connect\_to\_neuron\_group**(*neuron\_group*: [NeuronGroup](#), \*\**kwparams*) → None

Configure sorted spiking signal to record from given neuron group

**Parameters**

**neuron\_group** ([NeuronGroup](#)) – group to record from

**get\_state**() → tuple[NDArray[None, UInt], NDArray[None, Float], NDArray[None, UInt]]

Return spikes since method was last called (i, t\_ms, y)

**Returns**

(i, t\_ms, y) where i is channel (for multi-unit) or neuron (for sorted) spike indices, t\_ms is spike times, and y is a spike count vector suitable for control-theoretic uses—i.e., a 0 for every channel/neuron that hasn't spiked and a 1 for a single spike.

**Return type**

tuple[NDArray[np.uint], NDArray[float], NDArray[np.uint]]

**class** cleo.ephys.**Spiking**(*r\_perfect\_detection*: [Quantity](#), *r\_half\_detection*: [Quantity](#), *cutoff\_probability*: float  
= 0.01, \*, *name*: str = [\\_Nothing.NOTHING](#))

Bases: [Signal](#), [NeoExportable](#)

Base class for probabilistically detecting spikes

Method generated by attrs for class Spiking.

**connect\_to\_neuron\_group**(*neuron\_group*: [NeuronGroup](#), \*\**kwparams*) → ndarray

Configure signal to record from specified neuron group

**Parameters**

**neuron\_group** ([NeuronGroup](#)) – Neuron group to record from

**Returns**

num\_neurons\_to\_consider x num\_channels array of spike detection probabilities, for use in subclasses

**Return type**

np.ndarray

**cutoff\_probability:** float

Spike detection probability below which neurons will not be considered. For computational efficiency.

**abstract get\_state()** → tuple[NDArray[None, UInt], NDArray[None, Float], NDArray[None, UInt]]

Return spikes since method was last called (i, t\_ms, y)

**Returns**

(i, t\_ms, y) where i is channel (for multi-unit) or neuron (for sorted) spike indices, t\_ms is spike times, and y is a spike count vector suitable for control- theoretic uses—i.e., a 0 for every channel/neuron that hasn't spiked and a 1 for a single spike.

**Return type**

tuple[NDArray[np.uint], NDArray[float], NDArray[np.uint]]

**i:** NDArray[Any, np.uint]

Channel (for multi-unit) or neuron (for sorted) indices of spikes, stored if *save\_history* on *probe*

**i\_probe\_by\_i\_ng:** bidict

(neuron\_group, i\_ng) keys, i\_probe values. bidict for converting between neuron group indices and the indices the probe uses

**r\_half\_detection:** Quantity

Radius (with Brian unit) within which half of all spikes are detected

**r\_perfect\_detection:** Quantity

Radius (with Brian unit) within which all spikes are detected

**reset(\*\*kwargs)** → None

Reset signal to a neutral state

**t\_ms:** NDArray[Any, float]

Spike times in ms, stored if *save\_history* on *probe*

**t\_samp\_ms:** NDArray[Any, float]

Sample times in ms when each spike was recorded, stored if *save\_history* on *probe*

**to\_neo()** → Group

Return a Neo signal object with the device's data

**Returns**

Neo object representing exported data

**Return type**

neo.core.BaseNeo

**class cleo.ephys.TKLFPSignal**(uLFP\_threshold\_uV: float = 0.001, lfp\_unit: Unit = uvolt, \*, name: str = \_Nothing.NOTHING)

Bases: *LFPSignalBase*

Records the Teleñczuk kernel LFP approximation.

Requires tk\_lfp\_type='exc' | 'inh' to specify cell type on injection.

TKLFP is computed from spikes using the *tklfp* package.



**Injection kwargs**

**tklfp\_type** (*str*) – Either ‘exc’ or ‘inh’ to specify the cell type.

Method generated by attrs for class TKLFPSignal.

**connect\_to\_neuron\_group** (*neuron\_group*: *NeuronGroup*, *\*\*kwparams*)

Configure signal to record from specified neuron group

**Parameters**

**neuron\_group** (*NeuronGroup*) – group to record from

**get\_state**() → ndarray

Get the signal’s current value

**reset** (*\*\*kwargs*) → None

Reset signal to a neutral state

**uLFP\_threshold\_uV**: float

Threshold, in microvolts, above which the uLFP for a single spike is guaranteed to be considered, by default 1e-3. This determines the buffer length of past spikes, since the uLFP from a long-past spike becomes negligible and is ignored.

**cleo.ephys.linear\_shank\_coords** (*array\_length*: *Quantity*, *channel\_count*: *int*, *start\_location*: *Quantity* = *array([0., 0., 0.]) \* metre*, *direction*: *Tuple[float, float, float]* = (0, 0, 1)) → *Quantity*

Generate coordinates in a linear pattern

**Parameters**

- **array\_length** (*Quantity*) – Distance from the first to the last contact (with a Brian unit)
- **channel\_count** (*int*) – Number of coordinates to generate, i.e. electrode contacts
- **start\_location** (*Quantity*, *optional*) – x, y, z coordinate (with unit) for the start of the electrode array, by default (0, 0, 0)\*mm
- **direction** (*Tuple[float, float, float]*, *optional*) – x, y, z vector indicating the direction in which the array extends, by default (0, 0, 1), meaning pointing straight down

**Returns**

channel\_count x 3 array of coordinates, where the 3 columns represent x, y, and z

**Return type**

*Quantity*

**cleo.ephys.poly2\_shank\_coords** (*array\_length*: *Quantity*, *channel\_count*: *int*, *intercol\_space*: *Quantity*, *start\_location*: *Quantity* = *array([0., 0., 0.]) \* metre*, *direction*: *Tuple[float, float, float]* = (0, 0, 1)) → *Quantity*

Generate NeuroNexus-style Poly2 array coordinates

Poly2 refers to 2 parallel columns with staggered contacts. See <https://www.neuronexus.com/products/electrode-arrays/up-to-15-mm-depth> for more detail.

**Parameters**

- **array\_length** (*Quantity*) – Length from the beginning to the end of the two-column array, as measured in the center
- **channel\_count** (*int*) – Total (not per-column) number of coordinates (recording contacts) desired
- **intercol\_space** (*Quantity*) – Distance between columns (with Brian unit)

- **start\_location** (*Quantity*, *optional*) – Where to place the beginning of the array, by default (0, 0, 0)\*mm
- **direction** (*Tuple[float, float, float]*, *optional*) – x, y, z vector indicating the direction in which the two columns extend; by default (0, 0, 1), meaning straight down.

**Returns**

channel\_count x 3 array of coordinates, where the 3 columns represent x, y, and z

**Return type**

*Quantity*

```
cleo.ephys.poly3_shank_coords(array_length: Quantity, channel_count: int, intercol_space: Quantity,
                             start_location: Quantity = array([0., 0., 0.]) * metre, direction: Tuple[float, float, float] = (0, 0, 1)) → Quantity
```

Generate NeuroNexus Poly3-style array coordinates

Poly3 refers to three parallel columns of electrodes. The middle column will be longest if the channel count isn't divisible by three and the side columns will be centered vertically with respect to the middle.

**Parameters**

- **array\_length** (*Quantity*) – Length from beginning to end of the array as measured along the center column
- **channel\_count** (*int*) – Total (not per-column) number of coordinates to generate (i.e., electrode contacts)
- **intercol\_space** (*Quantity*) – Spacing between columns, with Brian unit
- **start\_location** (*Quantity*, *optional*) – Location of beginning of the array, that is, the first contact in the center column, by default (0, 0, 0)\*mm
- **direction** (*Tuple[float, float, float]*, *optional*) – x, y, z vector indicating the direction along which the array extends, by default (0, 0, 1), meaning straight down

**Returns**

channel\_count x 3 array of coordinates, where the 3 columns represent x, y, and z

**Return type**

*Quantity*

```
cleo.ephys.tetrode_shank_coords(array_length: Quantity, tetrode_count: int, start_location: Quantity =
                                array([0., 0., 0.]) * metre, direction: Tuple[float, float, float] = (0, 0, 1),
                                tetrode_width: Quantity = 25. * umetre) → Quantity
```

Generate coordinates for a linear array of tetrodes

See <https://www.neuronexus.com/products/electrode-arrays/up-to-15-mm-depth> to visualize NeuroNexus-style arrays.

**Parameters**

- **array\_length** (*Quantity*) – Distance from the center of the first tetrode to the last (with a Brian unit)
- **tetrode\_count** (*int*) – Number of tetrodes desired
- **start\_location** (*Quantity*, *optional*) – Center location of the first tetrode in the array, by default (0, 0, 0)\*mm
- **direction** (*Tuple[float, float, float]*, *optional*) – x, y, z vector determining the direction in which the linear array extends, by default (0, 0, 1), meaning straight down.

- **tetrode\_width** (*Quantity*, *optional*) – Distance between contacts in a single tetrode. Not the diagonal distance, but the length of one side of the square. By default 25\*umeter, as in NeuroNexus probes.

**Returns**

(tetrode\_count\*4) x 3 array of coordinates, where 3 columns represent x, y, and z

**Return type**

Quantity

`cleo.ephys.tile_coords(coords: Quantity, num_tiles: int, tile_vector: Quantity) → Quantity`

Tile (repeat) coordinates to produce multi-shank/matrix arrays

**Parameters**

- **coords** (*Quantity*) – The n x 3 coordinates array to tile
- **num\_tiles** (*int*) – Number of times to tile (repeat) the coordinates. For example, if you are tiling linear shank coordinates to produce multi-shank coordinates, this would be the desired number of shanks
- **tile\_vector** (*Quantity*) – x, y, z array with Brian unit determining both the length and direction of the tiling

**Returns**

(n \* num\_tiles) x 3 array of coordinates, where the 3 columns represent x, y, and z

**Return type**

Quantity

## 6.3.4 cleo.imaging module

Contains Scope and sensors for two-photon microscopy

**class** `cleo.imaging.CalBindingActivationModel(model: str)`

Bases: object

Base class for modeling calcium binding/activation

Method generated by attrs for class CalBindingActivationModel.

**model:** str

**class** `cleo.imaging.CalciumModel(model: str)`

Bases: object

Base class for how GECI computes calcium concentration

Must provide variable Ca (molar) in model.

Method generated by attrs for class CalciumModel.

**model:** str

**on\_pre:** str

**class** `cleo.imaging.DoubExpCalBindingActivation(*, A: float, tau_on: Quantity, tau_off: Quantity, Ca_rest: Quantity)`

Bases: *CalBindingActivationModel*

Double exponential kernel convolution representing CaB binding/activation.

Convolution is implemented via ODEs; see `notebooks/double_exp_conv_as_ode.ipynb` for derivation.

`A`, `tau_on`, and `tau_off` are the versions with proper scale and units of NAOMi's `ca_amp`, `t_on`, and `t_off`.

Some parameters found [here](#). Fitting code [here](#).

Method generated by attrs for class `DoubExpCalBindingActivation`.

**A: float**

amplitude of double exponential kernel

**Ca\_rest: Quantity**

Resting Ca<sup>2+</sup> concentration (molar).

**init\_syn\_vars**(*syn: Synapses*) → None

**model: str**

**tau\_off: Quantity**

CaB unbinding/deactivation time constant (sec)

**tau\_on: Quantity**

CaB binding/activation time constant (sec)

```
class cleo.imaging.DynamicCalcium(*, Ca_rest: Quantity, gamma: Quantity, B_T: Quantity, kappa_S: float,
                                   dCa_T: Quantity)
```

Bases: `CalciumModel`

Simulates intracellular calcium dynamics from spikes. Pieced together from Lütke et al., 2013; Helmchen and Tank, 2015; and Song et al., 2021. ([code](#))

Method generated by attrs for class `DynamicCalcium`.

**B\_T: Quantity**

total indicator (buffer) concentration (molar)

**Ca\_rest: Quantity**

resting Ca<sup>2+</sup> concentration (molar)

**dCa\_T: Quantity**

total Ca<sup>2+</sup> concentration increase per spike (molar)

**gamma: Quantity**

clearance/extrusion rate (1/sec)

**init\_syn\_vars**(*syn: Synapses*) → None

**kappa\_S: float**

Ca<sup>2+</sup> binding ratio of the endogenous buffer

**model: str**

from eq 8 in Lütke et al., 2013

**on\_pre: str**

from eq 9 in Lütke et al., 2013

```
class cleo.imaging.ExcitationModel(model: str)
```

Bases: `object`

Defines `exc_factor`

Method generated by attrs for class `ExcitationModel`.

**model:** str

```
class cleo.imaging.GECI(extra_namespace: dict = _Nothing.NOTHING, *, name: str = _Nothing.NOTHING,
                        save_history: bool = True, sigma_noise: float, dFF_1AP: float, cal_model:
                        CalciumModel, bind_act_model: CalBindingActivationModel, exc_model:
                        ExcitationModel, K_d: Quantity, n_H: float, dFF_max: float)
```

Bases: [Sensor](#)

GECI model based on Song et al., 2021, with interchangeable components.

See [geci\(\)](#) for a convenience function for creating GECI models.

As a potentially simpler alternative for future work, see the phenomenological S2F model from [Zhang et al., 2023](#). While parameter count looks similar, at least they have parameters fit already, and directly to data, rather than to biophysical processes before the data.

Method generated by attrs for class GECI.

**K\_d:** Quantity

indicator dissociation constant (binding affinity) (molar)

**bind\_act\_model:** [CalBindingActivationModel](#)

**cal\_model:** [CalciumModel](#)

**dFF\_max:** float

amplitude of Hill equation for conversion from Ca2+ to F/F, Fmax/F0. May be approximated from 'dynamic range' in literature Fmax/Fmin

**exc\_model:** [ExcitationModel](#)

**fluor\_model:** str

Uses a Hill equation to convert from Ca2+ to F/F, as in Song et al., 2021

**get\_state()** → dict[[NeuronGroup](#), ndarray]

Returns a {neuron\_group: fluorescence} dict of dFF values.

**init\_syn\_vars**(syn: [Synapses](#)) → None

Initializes appropriate variables in Synapses implementing the model

Also called on `reset()`.

#### Parameters

**syn** ([Synapses](#)) – The synapses object implementing this model

**location:** str

cytoplasm or membrane

#### Type

where sensor is expressed

**n\_H:** float

Hill coefficient for conversion from Ca2+ to F/F

**property params:** dict

Returns a dictionary of all parameters from model/submodels

**class** cleo.imaging.**LightExcitation**

Bases: [ExcitationModel](#)

Models light-dependent excitation (not implemented yet)

Method generated by attrs for class LightExcitation.

**model:** **str**

**class** cleo.imaging.**NullBindingActivation**

Bases: [CalBindingActivationModel](#)

Doesn't model binding/activation; i.e., goes straight from [Ca2+] to F/F

Method generated by attrs for class NullBindingActivation.

**model:** **str**

**class** cleo.imaging.**NullExcitation**

Bases: [ExcitationModel](#)

Models excitation as a constant factor

Method generated by attrs for class NullExcitation.

**model:** **str**

**class** cleo.imaging.**PreexistingCalcium**

Bases: [CalciumModel](#)

Calcium concentration is pre-existing in neuron model

Method generated by attrs for class PreexistingCalcium.

**model:** **str**

**class** cleo.imaging.**Scope**(*sensor: [Sensor](#), img\_width: [Quantity](#), focus\_depth: [Quantity](#) = None, location: [Quantity](#) = array([0., 0., 0.]) \* metre, direction: [Quantity](#) = (0, 0, 1), soma\_radius: [Quantity](#) = 10. \* umetre, snr\_cutoff: float = 1, rand\_seed: int = None, \*, name: str = [\\_Nothing.NOTHING](#), save\_history: bool = True)*)

Bases: [Recorder](#)

Two-photon microscope.

**Injection kwargs**

- **rho\_rel\_generator** (*Callable[[int], NDArray[(Any,), float]]*, optional) – A function assigning expression levels. Takes n as an arg, outputs float array. `lambda n: np.ones(n)` by default.
- **focus\_depth** (*Quantity*, optional) – The depth of the focal plane, by default that of the scope.
- **soma\_radius** (*Quantity*, optional) – The radius of the soma of the neuron, by default that of the scope. Used to compute noise focus factor, since smaller ROIs will have a noisier distribution of fluorescence, averaged over fewer pixels.

Method generated by attrs for class Scope.

**add\_self\_to\_plot**(*ax: [Axes3D](#), axis\_scale\_unit: [Unit](#), \*\*kwargs*) → list[[Artist](#)]

Add device to an existing plot

Should only be called by [plot\(\)](#).

**Parameters**

- **ax** (*Axes3D*) – The existing matplotlib Axes object
- **axis\_scale\_unit** (*Unit*) – The unit used to label axes and define chart limits
- **\*\*kwargs** (*optional*) –

**Returns**

A list of artists used to render the device. Needed for use in conjunction with [VideoVisualizer](#).

**Return type**

list[Artist]

**connect\_to\_neuron\_group**(*neuron\_group*: [NeuronGroup](#), **\*\*kwargs**) → None

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a [NeuronGroup](#), [Synapses](#), or [Monitor](#), make sure to add these to [brian\\_objects](#).

**Parameters**

- **neuron\_group** (*NeuronGroup*) –
- **\*\*kwargs** (*optional*) – Passed from *inject*

**dFF**: list[NDArray[Any, float]]

F/F from every call to [get\\_state\(\)](#). Shape is (n\_samples, n\_ROIs). Stored if [save\\_history](#)

**property dFF\_1AP**: NDArray[Any, Float]

dFF\_1AP for all targets, in order injected. Varies with expression levels.

**direction**: NDArray[3, float]

Direction in which the microscope is pointing. By default straight down (+z direction)

**focus\_coords\_per\_injct**: list[NDArray[Any, float]]

coordinates on the focal plane of neurons selected from each injection

**focus\_depth**: Quantity

The depth of the focal plane, with distance units

**get\_state()** → NDArray[Any, Float]

Returns a 1D array of F/F values for all targets, in order injected.

**Returns**

Fluorescence values for all targets

**Return type**

NDArray[(Any,), float]

**i\_targets\_for\_neuron\_group**(*neuron\_group*)

can handle multiple injections into same ng

**i\_targets\_per\_injct**: list[NDArray[Any, int]]

targets of neurons selected from each injection

**img\_width**: Quantity

The width (diameter) of the (circular) image captured by the microscope. Specified in distance units.

**inject\_sensor\_for\_targets**(**\*\*kwargs**) → None

**location**: Quantity

Location of the objective lens.

**property n: int**  
 Number of imaged ROIs

**neuron\_groups: list[NeuronGroup]**  
 neuron groups the scope has been injected into, in order of injection

**rand\_seed: int**

**reset(\*\*kwargs) → None**  
 Reset the device to a neutral state

**rho\_rel\_per\_injct: list[NDArray[Any, float]]**  
 relative expression levels of neurons selected from each injection

**sensor: [Sensor](#)**

**property sigma\_noise: NDArray[Any, Float]**  
 noise std dev (in terms of F/F) for all targets, in order injected.

**sigma\_per\_injct: list[NDArray[Any, float]]**  
*sigma\_noise* of neurons selected from each injection

**snr\_cutoff: float**  
 SNR below which neurons are discarded. Applied only when not *focus\_depth* is not None

**soma\_radius: Quantity**  
 Assumed radius of neurons, used to compute noise focus factor. Smaller neurons have noisier signals.

**t\_ms: list[float]**  
 Times at which sensor traces are recorded, in ms, stored if *save\_history*

**target\_neurons\_in\_plane**(*ng*, *focus\_depth*: *Quantity* = None, *soma\_radius*: *Quantity* = None) →  
 tuple[NDArray[Any, Int], NDArray[Any, Float], NDArray[Any, 3, Float]]  
 calls [target\\_neurons\\_in\\_plane\(\)](#) with scope parameter defaults. *focus\_depth* and *soma\_radius* can be overridden here.

#### Parameters

- **ng** (*NeuronGroup*) – The neuron group to target.
- **focus\_depth** (*Quantity*) – The depth of the focal plane, by default that of the microscope.
- **soma\_radius** (*Quantity*, *optional*) – The radius of the soma of the neuron, by default that of the microscope. Used to compute noise focus factor, since smaller ROIs will have a noisier distribution of fluorescence, averaged over fewer pixels.

#### Returns

A tuple of (i\_targets, noise\_focus\_factor, coords\_on\_plane)

#### Return type

Tuple[NDArray[(Any,), int], NDArray[(Any,), float], NDArray[(Any, 3), float]]

**class** cleo.imaging.**Sensor**(*extra\_namespace*: dict = *Nothing.NOTHING*, \*, *name*: str = *Nothing.NOTHING*, *save\_history*: bool = True, *sigma\_noise*: float, *dFF\_IAP*: float, *location*: str)

Bases: [SynapseDevice](#)

Base class for sensors

Method generated by attrs for class Sensor.



**dFF\_1AP: float**

F/F for 1 AP, only used for scope SNR cutoff

**property exc\_spectrum: list[tuple[float, float]]**

Excitation spectrum, alias for spectrum

**get\_state()** → dict[NeuronGroup, ndarray]

Returns a {neuron\_group: fluorescence} dict of dFF values.

**location: str**

cytoplasm or membrane

**Type**

where sensor is expressed

**sigma\_noise: float**

standard deviation of Gaussian noise in F/F measurement

**property snr: float**

Signal-to-noise ratio for 1 AP

```
cleo.imaging.gcamp3(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=287. *
nmolar, n_H=2.52, dFF_max=12, sigma_noise=0.018571428571428572,
dFF_1AP=0.039, ca_amp=0.05, t_on=1, t_off=1, Ca_rest=50. * nmolar, kappa_S=110,
gamma=292.3 * hertz, B_T=200. * umolar, dCa_T=7.6 * umolar, name='gcamp3',
**kwargs) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is None, then we don't have it fit yet.

ca\_amp, t\_on, and t\_off are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = ca\_amp / (second / 100)$  and  $\tau_{[on|off]} = second / t_{[on|off]}$ .

```
cleo.imaging.gcamp6f(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=290. *
nmolar, n_H=2.7, dFF_max=25.2, sigma_noise=0.03748181818181818,
dFF_1AP=0.09775500000000001, ca_amp=76.1251, t_on=0.8535, t_off=98.6173,
Ca_rest=50. * nmolar, kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar,
dCa_T=7.6 * umolar, name='gcamp6f', **kwargs) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is None, then we don't have it fit yet.

ca\_amp, t\_on, and t\_off are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = ca\_amp / (second / 100)$  and  $\tau_{[on|off]} = second / t_{[on|off]}$ .

```
cleo.imaging.gcamp6rs06(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=0.32 *
umolar, n_H=3, dFF_max=15, sigma_noise=0.030227272727272728,
dFF_1AP=None, ca_amp=None, t_on=None, t_off=None, Ca_rest=50. * nmolar,
kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar, dCa_T=7.6 * umolar,
name='gcamp6rs06', **kwargs) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = \text{ca\_amp} / (\text{second} / 100)$  and  $\text{tau}_{[\text{on}|\text{off}]} = \text{second} / \text{t}_{[\text{on}|\text{off}]}$ .

*Don't know sigma\_noise. Default is the GCaMP6s value.*

```
cleo.imaging.gcamp6rs09(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=0.52 *
    umolar, n_H=3.2, dFF_max=25, sigma_noise=0.030227272727272728,
    dFF_IAP=None, ca_amp=None, t_on=None, t_off=None, Ca_rest=50. * nmolar,
    kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar, dCa_T=7.6 * umolar,
    name='gcamp6rs09', **kwparams) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from NAOMi's code (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = \text{ca\_amp} / (\text{second} / 100)$  and  $\text{tau}_{[\text{on}|\text{off}]} = \text{second} / \text{t}_{[\text{on}|\text{off}]}$ .

*Don't know sigma\_noise. Default is the GCaMP6s value.*

```
cleo.imaging.gcamp6s(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=147. *
    nmolar, n_H=2.45, dFF_max=27.2, sigma_noise=0.030227272727272728,
    dFF_IAP=0.133, ca_amp=54.6943, t_on=0.4526, t_off=68.5461, Ca_rest=50. *
    nmolar, kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar, dCa_T=7.6 *
    umolar, name='gcamp6s', **kwparams) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from NAOMi's code (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = \text{ca\_amp} / (\text{second} / 100)$  and  $\text{tau}_{[\text{on}|\text{off}]} = \text{second} / \text{t}_{[\text{on}|\text{off}]}$ .

```
cleo.imaging.geci(light_dependent: bool, doub_exp_conv: bool, pre_existing_cal: bool, **kwparams) →
    GECI
```

Initializes a GECI model with given parameters.

#### Parameters

- **light\_dependent** (*bool*) – Whether the indicator is light-dependent.
- **doub\_exp\_conv** (*bool*) – Whether to use double exponential convolution for binding/activation.
- **pre\_existing\_cal** (*bool*) – Whether to use calcium concentrations already simulated in the neuron model.
- **\*\*kwparams** – Keyword parameters for GECI and sub-models.

#### Returns

A (LightDependent)GECI model specified submodels and parameters.

#### Return type

GECI

```
cleo.imaging.jgcamp7b(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=82. *
                      nmolar, n_H=3.06, dFF_max=22.1, sigma_noise=0.0075568181818182,
                      dFF_1AP=0.61712, ca_amp=None, t_on=None, t_off=None, Ca_rest=50. * nmolar,
                      kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar, dCa_T=7.6 * umolar,
                      name='jgcamp7b', **kwargs) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = ca\_amp / (second / 100)$  and  $\tau_{[on|off]} = second / t_{[on|off]}$ .

```
cleo.imaging.jgcamp7c(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=298. *
                      nmolar, n_H=2.44, dFF_max=145.6, sigma_noise=0.011788636363636364,
                      dFF_1AP=0.24605000000000002, ca_amp=None, t_on=None, t_off=None,
                      Ca_rest=50. * nmolar, kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar,
                      dCa_T=7.6 * umolar, name='jgcamp7c', **kwargs) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = ca\_amp / (second / 100)$  and  $\tau_{[on|off]} = second / t_{[on|off]}$ .

```
cleo.imaging.jgcamp7f(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=174. *
                      nmolar, n_H=2.3, dFF_max=30.2, sigma_noise=0.021763636363636363,
                      dFF_1AP=0.22743000000000002, ca_amp=None, t_on=None, t_off=None,
                      Ca_rest=50. * nmolar, kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar,
                      dCa_T=7.6 * umolar, name='jgcamp7f', **kwargs) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = ca\_amp / (second / 100)$  and  $\tau_{[on|off]} = second / t_{[on|off]}$ .

```
cleo.imaging.jgcamp7s(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=68. *
                      nmolar, n_H=2.49, dFF_max=40.4, sigma_noise=0.009975000000000001,
                      dFF_1AP=0.65968, ca_amp=None, t_on=None, t_off=None, Ca_rest=50. * nmolar,
                      kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar, dCa_T=7.6 * umolar,
                      name='jgcamp7s', **kwargs) → GECI
```

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = ca\_amp / (second / 100)$  and  $\tau_{[on|off]} = second / t_{[on|off]}$ .

`t_[on|off]`.

`cleo.imaging.ogb1(light_dependent=False, doub_exp_conv=True, pre_existing_cal=False, K_d=250. * nmolar, n_H=1, dFF_max=14, sigma_noise=0.0302272727272728, dFF_IAP=None, ca_amp=None, t_on=None, t_off=None, Ca_rest=50. * nmolar, kappa_S=110, gamma=292.3 * hertz, B_T=200. * umolar, dCa_T=7.6 * umolar, name='ogb1', **kwparams) → GECI`

Returns a (light-dependent) GECI model with specified submodel choices. Default parameters are taken from [NAOMi's code](#) (Song et al., 2021) as well as Dana et al., 2019 and Zhang et al., 2023.

Only those parameters used in chosen model components apply. If the default is `None`, then we don't have it fit yet.

`ca_amp`, `t_on`, and `t_off` are given as in NAOMi, but are converted to the proper scale and units for the double exponential convolution model. Namely,  $A = ca\_amp / (second / 100)$  and  $tau_{[on|off]} = second / t_{[on|off]}$ .

*Don't know `sigma_noise`. Default is the GCaMP6s value.*

`cleo.imaging.target_neurons_in_plane(ng: NeuronGroup, scope_focus_depth: Quantity, scope_img_width: Quantity, scope_location: Quantity = array([0., 0., 0.]) * metre, scope_direction: tuple = (0, 0, 1), soma_radius: Quantity = 10. * umetre, sensor_location: str = 'cytoplasm') → tuple[NDArray[Any, Int], NDArray[Any, Float], NDArray[Any, 3, Float]]`

Returns a tuple of (i\_targets, noise\_focus\_factor, coords\_on\_plane)

#### Parameters

- **ng** (*NeuronGroup*) – The neuron group to target.
- **scope\_focus\_depth** (*Quantity*) – The depth of the focal plane of the microscope.
- **scope\_img\_width** (*Quantity*) – The width of the image captured by the microscope.
- **scope\_location** (*Quantity*, *optional*) – The location of the microscope, by default (0, 0, 0) \* mm.
- **scope\_direction** (*tuple*, *optional*) – The direction of the microscope, by default (0, 0, 1).
- **soma\_radius** (*Quantity*, *optional*) – The radius of the soma of the neuron, by default 10 \* um. Used to compute noise focus factor, since smaller ROIs will have a noisier distribution of fluorescence, averaged over fewer pixels.
- **sensor\_location** (*str*, *optional*) – The location of the sensor, by default “cytoplasm”.

#### Returns

A tuple of (i\_targets, noise\_focus\_factor, coords\_on\_plane)

#### Return type

`Tuple[NDArray[(Any,), int], NDArray[(Any,), float], NDArray[(Any, 3), float]]`

### 6.3.5 cleo.ioproc module

Classes and functions for constructing and configuring an *IOProcessor*.

**class** cleo.ioproc.**ConstantDelay**(*delay\_ms: float*)

Bases: *Delay*

Simply adds a constant delay to the computation

**Parameters**

**delay\_ms** (*float*) – Desired delay in milliseconds

**compute**()

Compute delay.

**class** cleo.ioproc.**Delay**

Bases: ABC

Abstract base class for computing delays.

**abstract compute**() → float

Compute delay.

**class** cleo.ioproc.**FiringRateEstimator**(*tau\_ms: float, sample\_period\_ms: float, \*\*kwargs*)

Bases: *ProcessingBlock*

Exponential filter to estimate firing rate.

Requires *sample\_time\_ms* kwarg when process is called.

**Parameters**

- **tau\_ms** (*float*) – Time constant of filter
- **sample\_period\_ms** (*float*) – Sampling period in milliseconds

**compute\_output**(*input: NDArray[Any, UInt], \*\*kwargs*) → NDArray[Any, Float]

Estimate firing rate given past and current spikes.

**Parameters**

**input** (*NDArray[(n,), np.uint]*) – n-length vector of spike counts

**Keyword Arguments**

**sample\_time\_ms** (*float*) – Time measurement was taken in milliseconds

**Returns**

n-length vector of firing rates

**Return type**

NDArray[(n,), float]

**class** cleo.ioproc.**GaussianDelay**(*loc: float, scale: float*)

Bases: *Delay*

Generates normal-distributed delay.

Will return 0 when a negative value is sampled.

**Parameters**

- **loc** (*float*) – Center of distribution
- **scale** (*float*) – Standard deviation of delay distribution

**compute()** → float

Compute delay.

**class** cleo.ioproc.**LatencyIOProcessor**(*sample\_period\_ms: float = 1, sampling: str = 'fixed', processing: str = 'parallel'*)

Bases: *IOProcessor*

IOProcessor capable of delivering stimulation some time after measurement.

---

**Note:** It doesn't make much sense to combine parallel computation with "when idle" sampling, because "when idle" sampling only produces one sample at a time to process.

---

Method generated by attrs for class LatencyIOProcessor.

**get\_ctrl\_signals**(*query\_time\_ms*)

Get per-stimulator control signal from the *IOProcessor*.

**Parameters**

**query\_time\_ms** (*float*) – Current simulation time.

**Returns**

A {'stimulator\_name': ctrl\_signal} dictionary for updating stimulators.

**Return type**

dict

**is\_sampling\_now**(*query\_time\_ms*)

Determines whether the processor will take a sample at this timestep.

**Parameters**

**time** (*Brian 2 temporal Unit*) – Current timestep.

**Return type**

bool

**out\_buffer:** deque[Tuple[dict, float]]

**abstract process**(*state\_dict: dict, sample\_time\_ms: float*) → Tuple[dict, float]

Process network state to generate output to update stimulators.

This is the function the user must implement to define the signal processing pipeline.

**Parameters**

- **state\_dict** (*dict*) – {recorder\_name: state} dictionary from *get\_state()*

- **time\_ms** (*float*) –

**Returns**

{'stim\_name': ctrl\_signal} dictionary and output time in milliseconds.

**Return type**

Tuple[dict, float]

**processing:** str

"serial" or "parallel".

"parallel" computes the output time by adding the delay for a sample onto the sample time, so if the delay is 2 ms, for example, while the sample period is only 1 ms, some of the processing is happening in parallel. Output order matches input order even if the computed output time for a sample is sooner than that for a previous sample.

“serial” computes the output time by adding the delay for a sample onto the output time of the previous sample, rather than the sampling time. Note this may be of limited utility because it essentially means the *entire* round trip cannot be in parallel at all. More realistic is that simply each block or phase of computation must be serial. If anyone cares enough about this, it will have to be implemented in the future.

#### Type

Processing scheme

**put\_state**(*state\_dict: dict, sample\_time\_ms: float*)

Deliver network state to the IOProcessor.

#### Parameters

- **state\_dict** (*dict*) – A dictionary of recorder measurements, as returned by [get\\_state\(\)](#)
- **sample\_time\_ms** (*float*) – The current simulation timestep. Essential for simulating control latency and for time-varying control.

**sampling:** **str**

“fixed” or “when idle”.

“fixed” sampling means samples are taken on a fixed schedule, with no exceptions.

“when idle” sampling means no samples are taken before the previous sample’s output has been delivered. A sample is taken ASAP after an over-period computation: otherwise remains on schedule.

#### Type

Sampling scheme

**t\_samp\_ms:** **list[float]**

Record of sampling times—each time [put\\_state\(\)](#) is called.

**class** `cleo.ioproc.PIDcontroller`(*ref\_signal: callable, Kp: float, Ki: float = 0, sample\_period\_ms: float = 0, \*\*kwargs: Any*)

Bases: [ProcessingBlock](#)

Simple PI controller.

[compute\\_output\(\)](#) requires a `sample_time_ms` keyword argument. Only tested on controlling scalar values, but could be easily adapted to controlling a multi-dimensional state.

#### Parameters

- **ref\_signal** (*callable*) – Must return the target as a function of time in ms
- **Kp** (*float*) – Gain on the proportional error
- **Ki** (*float, optional*) – Gain on the integral error, by default 0
- **sample\_period\_ms** (*float, optional*) – Rate at which processor takes samples, by default 0. Only used to compute integrated error on first sample

**compute\_output**(*input: float, \*\*kwargs*) → float

Compute control input to the system using previously specified gains.

#### Parameters

**input** (*Any*) – Current system state

#### Returns

Control signal

#### Return type

float

**ref\_signal:** `callable[[float], Any]`

Callable returning the target as a function of time in ms

**class** `cleo.ioproc.ProcessingBlock(**kwargs)`

Bases: `ABC`

Abstract signal processing stage or control block.

It's important to use `super().__init__(**kwargs)` in the base class to use the parent-class logic here.

**Keyword Arguments**

**delay** (`Delay`) – Delay object which adds to the compute time

**Raises**

**TypeError** – When `delay` is not a `Delay` object.

**abstract compute\_output** (`input: Any, **kwargs`) → `Any`

Computes output for given input.

This is where the user will implement the desired functionality of the *ProcessingBlock* without regard for latency.

**Parameters**

- **input** (`Any`) – Data to be processed. Passed in from `process()`.
- **\*\*kwargs** (`Any`) – optional key-value argument pairs passed from `process()`. Could be used to pass in such values as the IO processor's walltime or the measurement time for time- dependent functions.

**Returns**

output

**Return type**

`Any`

**delay:** `Delay`

The delay object determining compute latency for the block

**process** (`input: Any, t_in_ms: float, **kwargs`) → `Tuple[Any, float]`

Computes and saves output and output time given input and input time.

The user should implement `compute_output()` for their child classes, which performs the computation itself without regards for timing or saving variables.

**Parameters**

- **input** (`Any`) – Data to be processed
- **t\_in\_ms** (`float`) – Time the block receives the input data
- **\*\*kwargs** (`Any`) – Key-value list of arguments passed to `compute_output()`

**Returns**

output, out time in milliseconds

**Return type**

`Tuple[Any, float]`

**save\_history:** `bool`

Whether to record `t_in_ms`, `t_out_ms`, and `values` with every timestep



**t\_in\_ms:** list[float]

The walltime the block received each input. Only recorded if *save\_history*

**t\_out\_ms:** list[float]

The walltime of each of the block's outputs. Only recorded if *save\_history*

**values:** list[Any]

Each of the block's outputs. Only recorded if *save\_history*

**class** cleo.ioproc.**RecordOnlyProcessor**(sample\_period\_ms, \*\*kwargs)

Bases: *LatencyIOProcessor*

Take samples without performing any control.

Use this if all you are doing is recording.

Method generated by attrs for class LatencyIOProcessor.

**out\_buffer:** deque[Tuple[dict, float]]

**process**(state\_dict: dict, sample\_time\_ms: float) → Tuple[dict, float]

Process network state to generate output to update stimulators.

This is the function the user must implement to define the signal processing pipeline.

#### Parameters

- **state\_dict** (dict) – {recorder\_name: state} dictionary from *get\_state()*
- **time\_ms** (float) –

#### Returns

{'stim\_name': ctrl\_signal} dictionary and output time in milliseconds.

#### Return type

Tuple[dict, float]

**processing:** str

“serial” or “parallel”.

“parallel” computes the output time by adding the delay for a sample onto the sample time, so if the delay is 2 ms, for example, while the sample period is only 1 ms, some of the processing is happening in parallel. Output order matches input order even if the computed output time for a sample is sooner than that for a previous sample.

“serial” computes the output time by adding the delay for a sample onto the output time of the previous sample, rather than the sampling time. Note this may be of limited utility because it essentially means the *entire* round trip cannot be in parallel at all. More realistic is that simply each block or phase of computation must be serial. If anyone cares enough about this, it will have to be implemented in the future.

#### Type

Processing scheme

**sampling:** str

“fixed” or “when idle”.

“fixed” sampling means samples are taken on a fixed schedule, with no exceptions.

“when idle” sampling means no samples are taken before the previous sample's output has been delivered. A sample is taken ASAP after an over-period computation: otherwise remains on schedule.

#### Type

Sampling scheme

**t\_samp\_ms:** `list[float]`

Record of sampling times—each time `put_state()` is called.

### 6.3.6 cleo.light module

**class** `cleo.light.GaussianEllipsoid`(*sigma\_axial*: *Quantity* = 18. \* *umetre*, *sigma\_lateral*: *Quantity* = 8. \* *umetre*)

Bases: *LightModel*

Method generated by attrs for class `GaussianEllipsoid`.

**sigma\_axial:** *Quantity*

Standard deviation distance along the focal axis.

Standard deviations estimated by taking point where response is ~60% of peak:

Publication	axial	lateral	measure
Prakash et al., 2012	13 m	7 m	photocurrent
Rickgauer et al., 2014	8 m	4 m	Ca2+ dF/F response
Packer et al., 2015	18 m	8 m	AP probability
Chen et al., 2019	18/11 m	8/? m	AP probability/photocurrent

**sigma\_lateral:** *Quantity*

Standard deviation distance along the focal plane.

**transmittance**(*source\_coords*: *Quantity*, *source\_dir\_uvec*: *NDArray*[*Any*, 3, *Any*], *target\_coords*: *Quantity*) → *NDArray*[*Any*, *Any*, *Float*]

Output must be between 0 and 1 with shape (n\_sources, n\_targets).

**viz\_params**(*coords*: *Quantity*, *direction*: *NDArray*[*Any*, 3, *Any*], *T\_threshold*: *float*, *n\_points\_per\_source*: *int* = 4000, *\*\*kwargs*) → *Quantity*

Returns info needed for visualization. Output is ((m, n\_points\_per\_source, 3) viz\_points array, marker-size\_um, intensity\_scale).

For best-looking results, implementations should scale *markersize\_um* and *intensity\_scale*.

**class** `cleo.light.Koehler`(*radius*: *Quantity*, *zmax*: *Quantity* = 0.5 \* *mmetre*)

Bases: *LightModel*

Even illumination over a circular area, with no scattering.

Method generated by attrs for class `Koehler`.

**radius:** *Quantity*

The radius of the Köhler beam

**transmittance**(*source\_coords*, *source\_dir\_uvec*, *target\_coords*)

Output must be between 0 and 1 with shape (n\_sources, n\_targets).

**viz\_params**(*coords*, *direction*, *T\_threshold*, *n\_points\_per\_source*=4000, *\*\*kwargs*)

Returns info needed for visualization. Output is ((m, n\_points\_per\_source, 3) viz\_points array, marker-size\_um, intensity\_scale).

For best-looking results, implementations should scale *markersize\_um* and *intensity\_scale*.

**zmax: Quantity**

The maximum extent of the Köhler beam, 500 m by default (i.e., no thicker than necessary to go through a slice or culture).

```
class cleo.light.Light(coords=array([0., 0., 0.]) * metre, direction: Quantity = (0, 0, 1), *, name: str =
    _Nothing.NOTHING, save_history: bool = True, light_model: LightModel,
    wavelength: Quantity = 0.473 * umetre, max_Irr0_mW_per_mm2: float = None,
    max_Irr0_mW_per_mm2_viz: float = None, default_value: NDArray[Any, Float] =
    _Nothing.NOTHING)
```

Bases: *Stimulator*

Delivers light to the network for photostimulation and (when implemented) imaging.

Requires neurons to have 3D spatial coordinates already assigned.

**Visualization kwargs**

- **n\_points\_per\_source** (*int, optional*) – The number of points per light source used to represent light intensity in space. Default varies by *light\_model*. Alias *n\_points*.
- **T\_threshold** (*float, optional*) – The transmittance below which no points are plotted. By default 1e-3.
- **intensity** (*float, optional*) – How bright the light appears, should be between 0 and 1. By default 0.5.
- **rasterized** (*bool, optional*) – Whether to render as rasterized in vector output, True by default. Useful since so many points makes later rendering and editing slow.

Method generated by attrs for class Light.

**add\_self\_to\_plot** (*ax, axis\_scale\_unit, \*\*kwargs*) → list[PathCollection]

Add device to an existing plot

Should only be called by *plot()*.

**Parameters**

- **ax** (*Axes3D*) – The existing matplotlib Axes object
- **axis\_scale\_unit** (*Unit*) – The unit used to label axes and define chart limits
- **\*\*kwargs** (*optional*) –

**Returns**

A list of artists used to render the device. Needed for use in conjunction with *VideoVisualizer*.

**Return type**

list[Artist]

**property color**

Color of light

**connect\_to\_neuron\_group** (*neuron\_group: NeuronGroup, \*\*kwparams: Any*) → None

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a *NeuronGroup*, *Synapses*, or *Monitor*, make sure to add these to *brian\_objects*.

**Parameters**

- **neuron\_group** (*NeuronGroup*) –

- **\*\*kwargs** (*optional*) – Passed from *inject*

**coords:** [Quantity](#)

(x, y, z) coords with Brian unit specifying where to place the base of the light source, by default (0, 0, 0)\*mm. Can also be an nx3 array for multiple sources.

**default\_value:** `NDArray[Any, Float]`

The default value of the device—used on initialization and on `reset()`

**direction:** `NDArray[Any, 3, Any]`

(x, y, z) vector specifying direction in which light source is pointing, by default (0, 0, 1).

Will be converted to unit magnitude.

**init\_for\_simulator**(*sim*: [CLSimulator](#)) → None

Initialize device for simulator on initial injection.

This function is called only the first time a device is injected into a simulator and performs any operations that are independent of the individual neuron groups it is connected to.

#### Parameters

**simulator** ([CLSimulator](#)) – simulator being injected into

**light\_model:** [LightModel](#)

Defines how light is emitted. See [OpticFiber](#) for an example.

**max\_Irr0\_mW\_per\_mm2:** `float`

The maximum irradiance the light source can emit.

Usually determined by hardware in a real experiment.

**max\_Irr0\_mW\_per\_mm2\_viz:** `float`

Maximum irradiance for visualization purposes.

i.e., the level at or above which the light appears maximally bright. Only relevant in video visualization.

**property n**

Number of light sources

**property source:** [Subgroup](#)

Returns the “neuron(s)” representing the light source(s).

**to\_neo()**

Return a Neo signal object with the device’s data

#### Returns

Neo object representing exported data

#### Return type

`neo.core.BaseNeo`

**transmittance**(*target\_coords*) → `ndarray`

Returns [light\\_model](#) transmittance given light’s coords and direction.

**update**(*value*: `float` | `ndarray`) → None

Set the light intensity, in mW/mm2 (without unit) for 1P excitation or laser power (mW) for 2P excitation (GaussianEllipsoid light model).

#### Parameters

**Irr0\_mW\_per\_mm2** (`float`) – Desired light intensity for light source

**update\_artists**(*artists*: list[Artist], *value*, \**args*, \*\**kwargs*) → list[Artist]

Update the artists used to render the device

Used to set the artists' state at every frame of a video visualization. The current state would be passed in \**args* or \*\**kwargs*

**Parameters**

**artists** (list[Artist]) – the artists used to render the device originally, i.e., which were returned from the first `add_self_to_plot()` call.

**Returns**

The artists that were actually updated. Needed for efficient blit rendering, where only updated artists are re-rendered.

**Return type**

list[Artist]

**wavelength:** Quantity

light wavelength with unit (usually nmeter)

```
class cleo.light.LightDependent(spectrum: list[tuple[float, float]] = _Nothing.NOTHING,
                                spectrum_interpolator: ~typing.Callable = <function
                                cubic_interpolator>)
```

Bases: object

Mix-in class for opsin and light-dependent indicator. Light-dependent devices are connected to light sources (and vice-versa) on injection via the registry.

We approximate dynamics under multiple wavelengths using a weighted sum of photon fluxes, where the factor indicates the activation relative to the peak-sensitivity wavelength for a equivalent power, which most papers report. When they report the action spectrum for equivalent photon flux instead (see Mager et al, 2018), use `equal_photon_flux_spectrum()`. This weighted sum is an approximation of a nonlinear peak-non-peak wavelength relation; see `notebooks/multi_wavelength_model.ipynb` for details.

Method generated by attrs for class LightDependent.

**epsilon**(*lambda\_new*) → float

Returns the  $\epsilon$  value for a given lambda (in nm) representing the relative sensitivity of the opsin to that wavelength.

**property light\_agg\_ngs**

**spectrum:** list[tuple[float, float]]

List of (wavelength, epsilon) tuples representing the action (opsin) or excitation (indicator) spectrum.

**spectrum\_interpolator:** Callable

Function of signature (lambdas\_nm, epsilons, lambda\_new\_nm) that interpolates the action spectrum data and returns  $\epsilon \in [0, 1]$  for the new wavelength.

```
class cleo.light.LightModel
```

Bases: ABC

Defines how light propagates given a source location and direction.

Method generated by attrs for class LightModel.

**abstract transmittance**(*source\_coords*: Quantity, *source\_direction*: NDArray[Any, 3, Any],  
*target\_coords*: Quantity) → NDArray[Any, Any, Float]

Output must be between 0 and 1 with shape (n\_sources, n\_targets).

**abstract viz\_params**(*coords: Quantity, direction: NDArray[Any, 3, Any], T\_threshold: float, n\_points\_per\_source: int = None, \*\*kwargs*) → tuple[NDArray[Any, Any, 3, Any], float, float]

Returns info needed for visualization. Output is ((m, n\_points\_per\_source, 3) viz\_points array, marker-size\_um, intensity\_scale).

For best-looking results, implementations should scale *markersize\_um* and *intensity\_scale*.

**class** cleo.light.OpticFiber(*R0: Quantity = 100. \* umetre, NAfib: float = 0.37, K: Quantity = 125. \* metre\*\*-1, S: Quantity = 7370. \* metre\*\*-1, ntis: float = 1.36*)

Bases: [LightModel](#)

Optic fiber light model from Foutz et al., 2012.

Defaults are from paper for 473 nm wavelength.

Method generated by attrs for class OpticFiber.

**K: Quantity**

absorbance coefficient (wavelength/tissue dependent)

**NAfib: float**

optical fiber numerical aperture

**R0: Quantity**

optical fiber radius

**S: Quantity**

scattering coefficient (wavelength/tissue dependent)

**ntis: float**

tissue index of refraction (wavelength/tissue dependent)

**transmittance**(*source\_coords: Quantity, source\_dir\_uvec: NDArray[Any, 3, Any], target\_coords: Quantity*) → NDArray[Any, Any, Float]

Output must be between 0 and 1 with shape (n\_sources, n\_targets).

**viz\_params**(*coords: Quantity, direction: NDArray[Any, 3, Any], T\_threshold: float, n\_points\_per\_source: int = 4000, \*\*kwargs*) → Quantity

Returns info needed for visualization. Output is ((m, n\_points\_per\_source, 3) viz\_points array, marker-size\_um, intensity\_scale).

For best-looking results, implementations should scale *markersize\_um* and *intensity\_scale*.

cleo.light.cubic\_interpolator(*lambdas\_nm, epsilons, lambda\_new\_nm*)

cleo.light.equal\_photon\_flux\_spectrum(*spectrum: list[tuple[float, float]]*) → list[tuple[float, float]]

Converts an equival photon flux spectrum to an equal power density spectrum.

cleo.light.fiber473nm(*R0=100. \* umetre, NAfib=0.37, K=125. \* metre\*\*-1, S=7370. \* metre\*\*-1, ntis=1.36*) → [OpticFiber](#)

Returns an [OpticFiber](#) model with parameters for 473 nm light.

Parameters from Foutz et al., 2012.

cleo.light.linear\_interpolator(*lambdas\_nm, epsilons, lambda\_new\_nm*)

cleo.light.plot\_spectra(*\*ldds: LightDependent*) → tuple[plt.Figure, plt.Axes]

Plots the action/excitation spectra for multiple light-dependent devices.

`cleo.light.tp_light_from_scope(scope, wavelength=1.06 * umetre, **kwargs) → Light`

Creates a light object from a scope object with 2P focused laser points at each target.

#### Parameters

- **scope** ([Scope](#)) – The scope object containing the laser spots.
- **wavelength** (*Quantity, optional*) – The wavelength of the laser, by default 1060 \* nmeter.

### 6.3.7 cleo.opto module

Contains opsin models, light sources, and some parameters

```
class cleo.opto.BansalFourStateOpsin(extra_namespace: dict = _Nothing.NOTHING, spectrum:  
list[tuple[float, float]] = _Nothing.NOTHING,  
spectrum_interpolator: Callable = <function cubic_interpolator>,  
Gd1: Quantity = 66. * hertz, Gd2: Quantity = 10. * hertz, Gr0:  
Quantity = 0.333 * hertz, g0: Quantity = 3.2 * nsiemens, phim:  
Quantity = 1.e+22 * metre ** -2 * second ** -1, k1: Quantity = 0.4  
* khertz, k2: Quantity = 120. * hertz, Gf0: Quantity = 18. * hertz,  
Gb0: Quantity = 8. * hertz, kf: Quantity = 10. * hertz, kb: Quantity  
= 8. * hertz, gamma: Quantity = 0.05, p: Quantity = 1, q: Quantity  
= 1, E: Quantity = 0. * volt, *, name: str = _Nothing.NOTHING,  
save_history: bool = True)
```

Bases: [MarkovOpsin](#)

4-state model from Bansal et al. 2020.

The difference from the PyRhO model is that there is no voltage dependence.

rho\_rel is channel density relative to standard model fit; modifying it post-injection allows for heterogeneous opsin expression.

IOPTO\_VAR\_NAME and V\_VAR\_NAME are substituted on injection.

Method generated by attrs for class BansalFourStateOpsin.

**E:** [Quantity](#)

**Gb0:** [Quantity](#)

**Gd1:** [Quantity](#)

**Gd2:** [Quantity](#)

**Gf0:** [Quantity](#)

**Gr0:** [Quantity](#)

**g0:** [Quantity](#)

**gamma:** [Quantity](#)

**init\_syn\_vars**(*opto\_syn: Synapses*) → None

Initializes appropriate variables in Synapses implementing the model

Also called on `reset()`.

#### Parameters

**syn** (*Synapses*) – The synapses object implementing this model

**k1:** Quantity

**k2:** Quantity

**kb:** Quantity

**kf:** Quantity

**model:** str

Basic Brian model equations string.

Should contain a *rho\_rel* term reflecting relative expression levels. Will likely also contain special NeuronGroup-dependent symbols such as V\_VAR\_NAME to be replaced on injection in `modify_model_and_params_for_ng()`.

**p:** Quantity

**phim:** Quantity

**q:** Quantity

```
class cleo.opto.BansalThreeStatePump(extra_namespace: dict = _Nothing.NOTHING, spectrum:
    list[tuple[float, float]] = _Nothing.NOTHING,
    spectrum_interpolator: Callable = <function cubic_interpolator>,
    Gd: Quantity = 250. * hertz, Gr: Quantity = 50. * hertz, ka:
    Quantity = msecond ** -1, p: Quantity = 0.7, q: Quantity = 0.1,
    phim: Quantity = 1.2e+24 * metre ** -2 * second ** -1, E: Quantity
    = -0.4 * volt, g0: Quantity = 22.34 * nsiemens, a: Quantity =
    2.e+08 * metre ** -3 * second ** -1 * amp ** -1 * mole, b: float =
    12, vartheta_max: Quantity = 5. * katal / (metre ** 3), kd: Quantity
    = 16. * mmolar, g_Cl: Quantity = 2.3 * nsiemens, Cl_out: Quantity
    = 124. * mmolar, Psi0: Quantity = 4.4286 * katal / (metre ** 3),
    E_Cl0: Quantity = -70. * mvolt, vmin: Quantity = -0.4 * volt, vmax:
    Quantity = 50. * mvolt, *, name: str = _Nothing.NOTHING,
    save_history: bool = True)
```

Bases: MarkovOpsin

3-state model from [Bansal et al. 2020](#). Defaults are for eNpHR3.0.

*rho\_rel* is channel density relative to standard model fit; modifying it post-injection allows for heterogeneous opsin expression.

IOPTO\_VAR\_NAME and V\_VAR\_NAME are substituted on injection.

Method generated by attrs for class BansalThreeStatePump.

**Cl\_out:** Quantity

**E:** Quantity

**E\_Cl0:** Quantity

**Gd:** Quantity

**Gr:** Quantity

**Psi0:** Quantity

**a:** Quantity



**b:** float

**g0:** Quantity

**g\_C1:** Quantity

**init\_syn\_vars**(*opto\_syn*: *Synapses*) → None

Initializes appropriate variables in Synapses implementing the model

Also called on `reset()`.

**Parameters**

**syn** (*Synapses*) – The synapses object implementing this model

**ka:** Quantity

**kd:** Quantity

**model:** str

Basic Brian model equations string.

Should contain a *rho\_rel* term reflecting relative expression levels. Will likely also contain special NeuronGroup-dependent symbols such as `V_VAR_NAME` to be replaced on injection in `modify_model_and_params_for_ng()`.

**p:** Quantity

**phim:** Quantity

**q:** Quantity

**vartheta\_max:** Quantity

**vmax:** Quantity

Needed to avoid jumps in `[Cl_in]` for EIF neurons

**vmin:** Quantity

Needed to avoid jumps in `[Cl_in]` for EIF neurons

```
class cleo.opto.FourStateOpsin(spectrum: list[tuple[float, float]] = _Nothing.NOTHING,
                             spectrum_interpolator: Callable = <function cubic_interpolator>, g0:
                             Quantity = 114. * nsiemens, gamma: Quantity = 0.00742, phim: Quantity =
                             2.33e+23 * metre ** -2 * second ** -1, k1: Quantity = 4.15 * khertz, k2:
                             Quantity = 0.868 * khertz, p: Quantity = 0.833, Gf0: Quantity = 37.3 *
                             hertz, kf: Quantity = 58.1 * hertz, Gb0: Quantity = 16.1 * hertz, kb:
                             Quantity = 63. * hertz, q: Quantity = 1.94, Gd1: Quantity = 105. * hertz,
                             Gd2: Quantity = 13.8 * hertz, Gr0: Quantity = 0.33 * hertz, E: Quantity =
                             0. * volt, v0: Quantity = 43. * mvolt, v1: Quantity = 17.1 * mvolt, *, name:
                             str = _Nothing.NOTHING, save_history: bool = True)
```

Bases: MarkovOpsin

4-state model from PyRhO (Evans et al. 2016).

`rho_rel` is channel density relative to standard model fit; modifying it post-injection allows for heterogeneous opsin expression.

`IOPTO_VAR_NAME` and `V_VAR_NAME` are substituted on injection.

Defaults are for Chr2.

Method generated by attrs for class FourStateOpsin.

**E:** Quantity

**Gb0:** Quantity

**Gd1:** Quantity

**Gd2:** Quantity

**Gf0:** Quantity

**Gr0:** Quantity

**extra\_namespace:** dict[str, Any]

Additional items (beyond parameters) to be added to the opto synapse namespace

**g0:** Quantity

**gamma:** Quantity

**init\_syn\_vars**(*opto\_syn*: Synapses) → None

Initializes appropriate variables in Synapses implementing the model

Also called on `reset()`.

**Parameters**

**syn** (Synapses) – The synapses object implementing this model

**k1:** Quantity

**k2:** Quantity

**kb:** Quantity

**kf:** Quantity

**model:** str

Basic Brian model equations string.

Should contain a *rho\_rel* term reflecting relative expression levels. Will likely also contain special NeuronGroup-dependent symbols such as `V_VAR_NAME` to be replaced on injection in `modify_model_and_params_for_ng()`.

**p:** Quantity

**phim:** Quantity

**q:** Quantity

**v0:** Quantity

**v1:** Quantity

```
class cleo.opto.Opsin(extra_namespace: dict = _Nothing.NOTHING, spectrum: list[tuple[float, float]] =  
    _Nothing.NOTHING, spectrum_interpolator: Callable = <function  
    cubic_interpolator>, *, name: str = _Nothing.NOTHING, save_history: bool = True)
```

Bases: [LightDependent](#), [SynapseDevice](#)

Base class for opsin model.

Requires that the neuron model has a current term (by default `Iopto`) which is assumed to be positive (unlike the convention in many opsin modeling papers, where the current is described as negative).

We approximate dynamics under multiple wavelengths using a weighted sum of photon fluxes, where the factor indicates the activation relative to the peak-sensitivity wavelength for an equivalent number of photons (see Mager et al, 2018). This weighted sum is an approximation of a nonlinear peak-non-peak wavelength relation; see `notebooks/multi_wavelength_model.ipynb` for details.

Method generated by attrs for class Opsin.

#### property `action_spectrum`

Alias for `light_receptor.spectrum`

```
class cleo.opto.ProportionalCurrentOpsin(extra_namespace: dict = _Nothing.NOTHING, spectrum:  
list[tuple[float, float]] = _Nothing.NOTHING,  
spectrum_interpolator: Callable = <function  
cubic_interpolator>, *, name: str = _Nothing.NOTHING,  
save_history: bool = True, I_per_Irr: Quantity)
```

Bases: [Opsin](#)

A simple model delivering current proportional to light intensity

Method generated by attrs for class ProportionalCurrentOpsin.

#### `I_per_Irr`: Quantity

How much current (in amps or unitless, depending on neuron model) to deliver per mW/mm2.

#### model: str

Basic Brian model equations string.

Should contain a *rho\_rel* term reflecting relative expression levels. Will likely also contain special NeuronGroup-dependent symbols such as `V_VAR_NAME` to be replaced on injection in `modify_model_and_params_for_ng()`.

#### required\_vars: list[Tuple[str, Unit]]

Default names of state variables required in the neuron group, along with units, e.g., [('Iopto', amp)].

It is assumed that non-default values can be passed in on injection as a keyword argument `[default_name]_var_name=[non_default_name]` and that these are found in the model string as `[DEFAULT_NAME]_VAR_NAME` before replacement.

`cleo.opto.chr2_4s()` → [FourStateOpsin](#)

Returns a 4-state Chr2 model.

Params taken from [try.projectpyrho.org](#)'s default 4-state configuration. Action spectrum from [Nagel et al., 2003, Fig. 4a](#), extracted using [Plot Digitizer](#).

Parameters can be changed after initialization but *before injection*.

`cleo.opto.chr2_b4s()` → [BansalFourStateOpsin](#)

Returns a 4-state Chr2 model.

Params given in [Bansal et al., 2020](#). Action spectrum from [Nagel et al., 2003, Fig. 4a](#), extracted using [Plot Digitizer](#).

Parameters can be changed after initialization but *before injection*.

`cleo.opto.chr2_h134r_4s()` → [BansalFourStateOpsin](#)

Returns a 4-state Chr2(H134R) model.

Params given in [Bansal et al., 2020](#). Action spectrum is same as for [chr2\\_4s\(\)](#), but blue-shifted 20 nm (I cannot find it directly in the literature).

Parameters can be changed after initialization but *before injection*.

`cleo.opto.chrimson_4s()` → *BansalFourStateOpsin*

Returns a 4-state Chrimson model.

Params given in Bansal et al., 2020. Action spectrum from Mager et al., 2018, Supp. Fig. 1a, extracted using Plot Digitizer.

Parameters can be changed after initialization but *before injection*.

`cleo.opto.enphr3_3s()`

Returns a 3-state model of eNpHR3, a chloride pump.

Params given in Bansal et al., 2020. Action spectrum from Gradinaru et al., 2010, Figure 3F, extracted using Plot Digitizer.

`cleo.opto.gtacr2_4s()` → *BansalFourStateOpsin*

Returns a 4-state model of GtACR2, an anion channel.

Params given in Bansal et al., 2020. Action spectra from Govorunova et al., 2015, Fig. 1f, extracted using Plot Digitizer.

Parameters can be changed after initialization but *before injection*.

`cleo.opto.vfchromson_4s()` → *BansalFourStateOpsin*

Returns a 4-state vf-Chrimson model.

Params given in Bansal et al., 2020. Action spectrum from Mager et al., 2018, Supp. Fig. 1a, extracted using Plot Digitizer.

Parameters can be changed after initialization but *before injection*.

### 6.3.8 cleo.recorders module

Contains basic recorders.

**class** `cleo.recorders.GroundTruthSpikeRecorder`(\* , name: str = *\_Nothing.NOTHING*, save\_history: bool = True)

Bases: *Recorder*

Reports the number of spikes seen since last queried for each neuron.

This amounts effectively to the number of spikes per control period. Note: this will only work for one neuron group at the moment.

Method generated by attrs for class GroundTruthSpikeRecorder.

**connect\_to\_neuron\_group**(neuron\_group)

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a NeuronGroup, Synapses, or Monitor, make sure to add these to *brian\_objects*.

#### Parameters

- **neuron\_group** (*NeuronGroup*) –
- **\*\*kwargs** (*optional*) – Passed from *inject*

**get\_state**() → *NDArray[Any, UInt]*

#### Returns

*n\_neurons*-length array with spike counts over the latest control period.

**Return type**

NDArray[(n\_neurons,), np.uint]

**neuron\_group:** [NeuronGroup](#)

**class** cleo.recorders.**RateRecorder**(*i: int, \*, name: str = \_Nothing.NOTHING, save\_history: bool = True*)

Bases: [Recorder](#)

Records firing rate from a single neuron.

Firing rate comes from Brian's [PopulationRateMonitor](#)

Method generated by attrs for class RateRecorder.

**connect\_to\_neuron\_group**(*neuron\_group*)

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a NeuronGroup, Synapses, or Monitor, make sure to add these to [brian\\_objects](#).

**Parameters**

- **neuron\_group** (*NeuronGroup*) –
- **\*\*kwargs** (*optional*) – Passed from *inject*

**get\_state**()

Return current measurement.

**i: int**

index of neuron to record

**mon:** [PopulationRateMonitor](#)

**class** cleo.recorders.**VoltageRecorder**(*voltage\_var\_name: str = 'v', \*, name: str = \_Nothing.NOTHING, save\_history: bool = True*)

Bases: [Recorder](#)

Records the voltage of a single neuron group.

Method generated by attrs for class VoltageRecorder.

**connect\_to\_neuron\_group**(*neuron\_group*)

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a NeuronGroup, Synapses, or Monitor, make sure to add these to [brian\\_objects](#).

**Parameters**

- **neuron\_group** (*NeuronGroup*) –
- **\*\*kwargs** (*optional*) – Passed from *inject*

**get\_state**() → [Quantity](#)

**Returns**

Current voltage of target neuron group

**Return type**

Quantity

**mon:** [StateMonitor](#)

**voltage\_var\_name:** str

Name of variable representing membrane voltage

### 6.3.9 cleo.registry module

Code for orchestrating inter-device interactions.

This should only be relevant for developers, not users, as this code is used under the hood when interacting devices are injected (e.g., light and opsin).

**class** cleo.registry.DeviceInteractionRegistry(*sim*: CLSimulator)

Bases: object

Facilitates the creation and maintenance of ‘neurons’ and ‘synapses’ implementing many-to-many light-opsin/indicator relationships

Method generated by attrs for class DeviceInteractionRegistry.

**brian\_objects:** set

Stores all Brian objects created (and injected into the network) by this registry

**connect\_light\_to\_ldd\_for\_ng**(*light*: Light, *ldd*: LightDependent, *ng*: NeuronGroup) → None

Connects a light to a light-dependent device for a given neuron group.

**Parameters**

- **light** (Light) – Light being injected
- **ldd** (LightDependent) – Light-dependent device the light will affect
- **ng** (NeuronGroup) – Neurons affected by the light-dependent device

**Raises**

**ValueError** – if the connection has already been made

**connections:** set[Tuple['Light', 'LightDependent', NeuronGroup]]

Set of (light, light-dependent device, neuron group) tuples representing previously created connections.

**init\_register\_light**(*light*: Light) → Subgroup

Creates neurons for the light source, if they don’t already exist

**ldds\_for\_ng:** dict[NeuronGroup, set['LightDependent']]

Maps neuron group to the light-dependent devices injected into it

**light\_prop\_model** = '\n T : 1\n epsilon : 1\n Ephoton : joule\n Irr\_post = epsilon \* T \* Irr0\_pre : watt/meter\*\*2 (summed)\n phi\_post = Irr\_post / Ephoton : 1/second/meter\*\*2 (summed)\n '

Model used in light propagation synapses

**light\_prop\_syms:** dict[Tuple['LightDependent', NeuronGroup], Synapses]

Maps (light-dependent device, neuron group) to the synapses implementing light propagation

**light\_source\_ng:** NeuronGroup

Represents ALL light sources (multiple devices)

**lights\_for\_ng:** dict[NeuronGroup, set['Light']]

Maps neuron group to the lights injected into it

**register**(*device*: [InterfaceDevice](#), *ng*: *NeuronGroup*) → None

Registers a device injection with the registry.

**Parameters**

- **device** ([InterfaceDevice](#)) – Device being injected
- **ng** (*NeuronGroup*) – Neurons being injected into

**register\_ldd**(*ldd*: [LightDependent](#), *ng*: *NeuronGroup*)

Connects lights previously injected into this neuron group to this light-dependent device

**register\_light**(*light*: [Light](#), *ng*: *NeuronGroup*)

Connects light to light-dependent devices already injected into this neuron group

**sim**: [CLSimulator](#)

**source\_for\_light**(*light*: [Light](#)) → Subgroup

Returns the subgroup representing the given light source

**subgroup\_idx\_for\_light**: dict['Light', slice]

Maps light to its indices in the [light\\_source\\_ng](#)

`cleo.registry.registries`: dict['CLSimulator', [DeviceInteractionRegistry](#)] = {}

Maps simulator to its registry

`cleo.registry.registry_for_sim`(*sim*: [CLSimulator](#)) → [DeviceInteractionRegistry](#)

Returns the registry for the given simulator

### 6.3.10 cleo.stimulators module

Contains basic stimulators.

**class** `cleo.stimulators.StateVariableSetter`(*default\_value*: Any = 0, \*, *name*: str = [\\_Nothing.NOTHING](#),  
*save\_history*: bool = True, *variable\_to\_ctrl*: str, *unit*: Unit)

Bases: [Stimulator](#)

Sets the given state variable of target neuron groups.

Method generated by attrs for class StateVariableSetter.

**connect\_to\_neuron\_group**(*neuron\_group*)

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a *NeuronGroup*, *Synapses*, or *Monitor*, make sure to add these to [brian\\_objects](#).

**Parameters**

- **neuron\_group** (*NeuronGroup*) –
- **\*\*kwargs** (*optional*) – Passed from *inject*

**neuron\_groups**: list[*NeuronGroup*]

**unit**: Unit

will be used in [update\(\)](#)

**Type**

Unit of controlled variable

**update**(*ctrl\_signal: float*) → None

Set state variable of target neuron groups

**Parameters**

**ctrl\_signal** (*float*) – Value to update variable to, without unit. The unit provided on initialization is automatically multiplied.

**variable\_to\_ctrl:** **str**

Name of state variable to control

### 6.3.11 cleo.utilities module

Assorted utilities for developers.

**cleo.utilities.add\_to\_neo\_segment**(*segment: Segment, \*objects: DataObject*)

Conveniently adds multiple objects to a segment.

Taken from `neo.core.group.Group`.

**cleo.utilities.analog\_signal**(*t\_ms, values\_no\_unit, units=""*) → BaseSignal

**cleo.utilities.brian\_safe\_name**(*name: str*) → str

**cleo.utilities.get\_orth\_vectors\_for\_V**(*V*)

For nx3 block of row vectors V, return nx3 W1, W2 orthogonal vector blocks

**cleo.utilities.modify\_model\_with\_eqs**(*neuron\_group, eqs\_to\_add*)

Adapted from `_create_variables()` from `neurongroup.py` from Brian2 source code v2.3.0.2

**cleo.utilities.normalize\_coords**(*coords: Quantity*) → Quantity

Normalize coordinates to unit vectors.

**cleo.utilities.rng = Generator(PCG64) at 0x7FF4F2EFD380**

supposed to be the central random number generator, but not yet used everywhere

**cleo.utilities.style\_plots\_for\_docs**(*dark=True*)

**cleo.utilities.times\_are\_regular**(*times*)

**cleo.utilities.uniform\_cylinder\_rz**(*n, rmax, zmax*)

uniformly fills a cylinder with radius rmax and height zmax with n points.

Does so by generating a Fibonacci spiral cylinder by rotating around axis and up and down cylinder simultaneously, using different angle steps.

**cleo.utilities.unit\_safe\_append**(*q1: Quantity, q2: Quantity, axis=0*)

**cleo.utilities.wavelength\_to\_rgb**(*wavelength\_nm, gamma=0.8*) → tuple[float, float, float]

taken from [http://www.noah.org/wiki/Wavelength\\_to\\_RGB\\_in\\_Python](http://www.noah.org/wiki/Wavelength_to_RGB_in_Python) This converts a given wavelength of light to an approximate RGB color value. The wavelength must be given in nanometers in the range from 380 nm through 750 nm (789 THz through 400 THz).

Based on code by Dan Bruton <http://www.physics.sfasu.edu/astro/color/spectra.html>

**cleo.utilities.xyz\_from\_rz**(*rs, thetas, zs, xyz\_start, xyz\_end*)

Convert from cylindrical to Cartesian coordinates.



### 6.3.12 cleo.viz module

Tools for visualizing models and simulations

```
class cleo.viz.VideoVisualizer(devices: Iterable[InterfaceDevice | Tuple[InterfaceDevice, dict]] =  
    _Nothing.NOTHING, dt: Quantity = 1. * msecond, *, name: str =  
    _Nothing.NOTHING, save_history: bool = True)
```

Bases: [InterfaceDevice](#)

Device for visualizing a simulation.

Must be injected after all other devices and before the simulation is run.

Method generated by attrs for class VideoVisualizer.

**ax:** [Axes](#)

```
connect_to_neuron_group(neuron_group: NeuronGroup, **kwargs) → None
```

Connect device to given *neuron\_group*.

If your device introduces any objects which Brian must keep track of, such as a NeuronGroup, Synapses, or Monitor, make sure to add these to [brian\\_objects](#).

#### Parameters

- **neuron\_group** (*NeuronGroup*) –
- **\*\*kwargs** (*optional*) – Passed from *inject*

```
devices: Iterable\[InterfaceDevice | Tuple\[InterfaceDevice, dict\]\]
```

list of devices or (device, vis\_kwargs) tuples to include in the plot, just as in the [plot\(\)](#) function, by default “all”, which will include all recorders and stimulators currently injected when this visualizer is injected into the simulator.

**dt:** [Quantity](#)

length of each frame—that is, every dt the visualizer takes a snapshot of the network, by default 1\*ms

**fig:** [Figure](#)

```
generate_Animation(plotargs: dict, slowdown_factor: float = 10, **figargs: Any) → Animation
```

Create a matplotlib Animation object from the recorded simulation

#### Parameters

- **plotargs** (*dict*) – dictionary of arguments as taken by [plot\(\)](#). can include *xlim*, *ylim*, *zlim*, *colors*, *axis\_scale\_unit*, *invert\_z*, and/or *scatterargs*. neuron groups and devices are automatically added and *\*\*figargs* are specified separately.
- **slowdown\_factor** (*float, optional*) – how much slower the animation will be rendered, as a multiple of real-time, by default 10
- **\*\*figargs** (*Any, optional*) – keyword arguments passed to `plt.figure()`, such as `figsize`

#### Returns

An Animation object capturing the desired visualization. See matplotlib’s docs for saving and rendering options.

#### Return type

[matplotlib.animation.Animation](#)

**init\_for\_simulator**(*simulator*: CLSimulator)

Initialize device for simulator on initial injection.

This function is called only the first time a device is injected into a simulator and performs any operations that are independent of the individual neuron groups it is connected to.

**Parameters**

**simulator** (CLSimulator) – simulator being injected into

**neuron\_groups**: list[NeuronGroup]

`cleo.viz.plot(*neuron_groups: NeuronGroup, xlim: Tuple[float, float] = None, ylim: Tuple[float, float] = None, zlim: Tuple[float, float] = None, colors: Iterable = None, axis_scale_unit: Unit = umetre, devices: Iterable[InterfaceDevice | Tuple[InterfaceDevice, dict]] = [], invert_z: bool = True, scatterargs: dict = {}, sim: CLSimulator = None, **figargs: Any) → None`

Visualize neurons and interface devices

**Parameters**

- **xlim** (Tuple[float, float], optional) – xlim for plot, determined automatically by default
- **ylim** (Tuple[float, float], optional) – ylim for plot, determined automatically by default
- **zlim** (Tuple[float, float], optional) – zlim for plot, determined automatically by default
- **colors** (Iterable, optional) – colors, one for each neuron group, automatically determined by default
- **axis\_scale\_unit** (Unit, optional) – Brian unit to scale lim params, by default mm
- **devices** (Iterable[Union[InterfaceDevice, Tuple[InterfaceDevice, dict]]], optional) – devices to add to the plot or (device, kwargs) tuples. `add_self_to_plot()` is called for each, using the kwargs dict if given. By default []
- **invert\_z** (bool, optional) – whether to invert z-axis, by default True to reflect the convention that +z represents depth from cortex surface
- **scatterargs** (dict, optional) – arguments passed to `plt.scatter()` for each neuron group, such as marker
- **sim** (CLSimulator, optional) – Optional shortcut to include all neuron groups and devices
- **\*\*figargs** (Any, optional) – keyword arguments passed to `plt.figure()`, such as `figsize`

**Raises**

**ValueError** – When neuron group doesn't have x, y, and z already defined

## INDICES AND TABLES

- `genindex`
- `modindex`
- `search`



## PYTHON MODULE INDEX

### C

- `cleo`, [108](#)
- `cleo.coords`, [115](#)
- `cleo.ephys`, [117](#)
- `cleo.imaging`, [127](#)
- `cleo.ioproc`, [137](#)
- `cleo.light`, [142](#)
- `cleo.opto`, [147](#)
- `cleo.recorders`, [152](#)
- `cleo.registry`, [154](#)
- `cleo.stimulators`, [155](#)
- `cleo.utilities`, [156](#)
- `cleo.viz`, [157](#)



## A

A (*cleo.imaging.DoubExpCalBindingActivation* attribute), 128  
a (*cleo.opto.BansalThreeStatePump* attribute), 148  
action\_spectrum (*cleo.opto.Opsin* property), 151  
add\_self\_to\_plot() (*cleo.ephys.Probe* method), 118  
add\_self\_to\_plot() (*cleo.imaging.Scope* method), 130  
add\_self\_to\_plot() (*cleo.InterfaceDevice* method), 110  
add\_self\_to\_plot() (*cleo.light.Light* method), 143  
add\_signals() (*cleo.ephys.Probe* method), 118  
add\_to\_neo\_segment() (in module *cleo.utilities*), 156  
amp\_func (*cleo.ephys.RWSLFPSignalBase* attribute), 120  
analog\_signal() (in module *cleo.utilities*), 156  
assign\_coords() (in module *cleo.coords*), 115  
assign\_coords\_grid\_rect\_prism() (in module *cleo.coords*), 115  
assign\_coords\_rand\_cylinder() (in module *cleo.coords*), 115  
assign\_coords\_rand\_rect\_prism() (in module *cleo.coords*), 115  
assign\_coords\_uniform\_cylinder() (in module *cleo.coords*), 115  
assign\_xyz() (in module *cleo.coords*), 116  
ax (*cleo.viz.VideoVisualizer* attribute), 157

## B

b (*cleo.opto.BansalThreeStatePump* attribute), 148  
B\_T (*cleo.imaging.DynamicCalcium* attribute), 128  
BansalFourStateOpsin (class in *cleo.opto*), 147  
BansalThreeStatePump (class in *cleo.opto*), 148  
bind\_act\_model (*cleo.imaging.GECI* attribute), 129  
brian\_objects (*cleo.ephys.LFPSignalBase* attribute), 117  
brian\_objects (*cleo.ephys.Signal* attribute), 122  
brian\_objects (*cleo.InterfaceDevice* attribute), 111  
brian\_objects (*cleo.registry.DeviceInteractionRegistry* attribute), 154  
brian\_safe\_name() (in module *cleo.utilities*), 156

## C

Ca\_rest (*cleo.imaging.DoubExpCalBindingActivation* attribute), 128  
Ca\_rest (*cleo.imaging.DynamicCalcium* attribute), 128  
cal\_model (*cleo.imaging.GECI* attribute), 129  
CalBindingActivationModel (class in *cleo.imaging*), 127  
CalciumModel (class in *cleo.imaging*), 127  
chr2\_4s() (in module *cleo.opto*), 151  
chr2\_b4s() (in module *cleo.opto*), 151  
chr2\_h134r\_4s() (in module *cleo.opto*), 151  
chromson\_4s() (in module *cleo.opto*), 151  
Cl\_out (*cleo.opto.BansalThreeStatePump* attribute), 148  
cleo  
    module, 108  
cleo.coords  
    module, 115  
cleo.ephys  
    module, 117  
cleo.imaging  
    module, 127  
cleo.ioproc  
    module, 137  
cleo.light  
    module, 142  
cleo.opto  
    module, 147  
cleo.recorders  
    module, 152  
cleo.registry  
    module, 154  
cleo.stimulators  
    module, 155  
cleo.utilities  
    module, 156  
cleo.viz  
    module, 157  
CLSimulator (class in *cleo*), 108  
color (*cleo.light.Light* property), 143  
compute() (*cleo.ioproc.ConstantDelay* method), 137  
compute() (*cleo.ioproc.Delay* method), 137  
compute() (*cleo.ioproc.GaussianDelay* method), 137

- `compute_output()` (*cleo.ioproc.FiringRateEstimator* method), 137  
`compute_output()` (*cleo.ioproc.PIController* method), 139  
`compute_output()` (*cleo.ioproc.ProcessingBlock* method), 140  
`concat_coords()` (in module *cleo.coords*), 116  
`connect_light_to_ldd_for_ng()` (*cleo.registry.DeviceInteractionRegistry* method), 154  
`connect_to_neuron_group()` (*cleo.ephys.MultiUnitSpiking* method), 117  
`connect_to_neuron_group()` (*cleo.ephys.Probe* method), 118  
`connect_to_neuron_group()` (*cleo.ephys.RWSLFPSignalFromPSCs* method), 121  
`connect_to_neuron_group()` (*cleo.ephys.RWSLFPSignalFromSpikes* method), 122  
`connect_to_neuron_group()` (*cleo.ephys.Signal* method), 122  
`connect_to_neuron_group()` (*cleo.ephys.SortedSpiking* method), 123  
`connect_to_neuron_group()` (*cleo.ephys.Spiking* method), 123  
`connect_to_neuron_group()` (*cleo.ephys.TKLFPSignal* method), 125  
`connect_to_neuron_group()` (*cleo.imaging.Scope* method), 131  
`connect_to_neuron_group()` (*cleo.InterfaceDevice* method), 111  
`connect_to_neuron_group()` (*cleo.light.Light* method), 143  
`connect_to_neuron_group()` (*cleo.recorders.GroundTruthSpikeRecorder* method), 152  
`connect_to_neuron_group()` (*cleo.recorders.RateRecorder* method), 153  
`connect_to_neuron_group()` (*cleo.recorders.VoltageRecorder* method), 153  
`connect_to_neuron_group()` (*cleo.stimulators.StateVariableSetter* method), 155  
`connect_to_neuron_group()` (*cleo.SynapseDevice* method), 113  
`connect_to_neuron_group()` (*cleo.viz.VideoVisualizer* method), 157  
`connections` (*cleo.registry.DeviceInteractionRegistry* attribute), 154  
`ConstantDelay` (class in *cleo.ioproc*), 137  
`coords` (*cleo.ephys.Probe* attribute), 119  
`coords` (*cleo.light.Light* attribute), 144  
`coords_from_ng()` (in module *cleo.coords*), 116  
`coords_from_xyz()` (in module *cleo.coords*), 116  
`cubic_interpolator()` (in module *cleo.light*), 146  
`cutoff_probability` (*cleo.ephys.Spiking* attribute), 124
- ## D
- `dCa_T` (*cleo.imaging.DynamicCalcium* attribute), 128  
`default_value` (*cleo.light.Light* attribute), 144  
`default_value` (*cleo.Stimulator* attribute), 112  
`Delay` (class in *cleo.ioproc*), 137  
`delay` (*cleo.ioproc.ProcessingBlock* attribute), 140  
`DeviceInteractionRegistry` (class in *cleo.registry*), 154  
`devices` (*cleo.CLSimulator* attribute), 108  
`devices` (*cleo.viz.VideoVisualizer* attribute), 157  
`dFF` (*cleo.imaging.Scope* attribute), 131  
`dFF_1AP` (*cleo.imaging.Scope* property), 131  
`dFF_1AP` (*cleo.imaging.Sensor* attribute), 132  
`dFF_max` (*cleo.imaging.GECI* attribute), 129  
`direction` (*cleo.imaging.Scope* attribute), 131  
`direction` (*cleo.light.Light* attribute), 144  
`DoubExpCalBindingActivation` (class in *cleo.imaging*), 127  
`dt` (*cleo.viz.VideoVisualizer* attribute), 157  
`DynamicCalcium` (class in *cleo.imaging*), 128
- ## E
- `E` (*cleo.opto.BansalFourStateOpsin* attribute), 147  
`E` (*cleo.opto.BansalThreeStatePump* attribute), 148  
`E` (*cleo.opto.FourStateOpsin* attribute), 149  
`E_Cl0` (*cleo.opto.BansalThreeStatePump* attribute), 148  
`enphr3_3s()` (in module *cleo.opto*), 152  
`epsilon()` (*cleo.light.LightDependent* method), 145  
`equal_photon_flux_spectrum()` (in module *cleo.light*), 146  
`exc_model` (*cleo.imaging.GECI* attribute), 129  
`exc_spectrum` (*cleo.imaging.Sensor* property), 133  
`ExcitationModel` (class in *cleo.imaging*), 128  
`extra_namespace` (*cleo.opto.FourStateOpsin* attribute), 150  
`extra_namespace` (*cleo.SynapseDevice* attribute), 113
- ## F
- `fiber473nm()` (in module *cleo.light*), 146  
`fig` (*cleo.viz.VideoVisualizer* attribute), 157  
`FiringRateEstimator` (class in *cleo.ioproc*), 137  
`fluor_model` (*cleo.imaging.GECI* attribute), 129  
`focus_coords_per_injct` (*cleo.imaging.Scope* attribute), 131  
`focus_depth` (*cleo.imaging.Scope* attribute), 131  
`FourStateOpsin` (class in *cleo.opto*), 149



## G

- `g0` (*cleo.opto.BansalFourStateOpsin* attribute), 147
  - `g0` (*cleo.opto.BansalThreeStatePump* attribute), 149
  - `g0` (*cleo.opto.FourStateOpsin* attribute), 150
  - `g_C1` (*cleo.opto.BansalThreeStatePump* attribute), 149
  - `gamma` (*cleo.imaging.DynamicCalcium* attribute), 128
  - `gamma` (*cleo.opto.BansalFourStateOpsin* attribute), 147
  - `gamma` (*cleo.opto.FourStateOpsin* attribute), 150
  - `GaussianDelay` (class in *cleo.ioproc*), 137
  - `GaussianEllipsoid` (class in *cleo.light*), 142
  - `Gb0` (*cleo.opto.BansalFourStateOpsin* attribute), 147
  - `Gb0` (*cleo.opto.FourStateOpsin* attribute), 150
  - `gcamp3()` (in module *cleo.imaging*), 133
  - `gcamp6f()` (in module *cleo.imaging*), 133
  - `gcamp6rs06()` (in module *cleo.imaging*), 133
  - `gcamp6rs09()` (in module *cleo.imaging*), 134
  - `gcamp6s()` (in module *cleo.imaging*), 134
  - `Gd` (*cleo.opto.BansalThreeStatePump* attribute), 148
  - `Gd1` (*cleo.opto.BansalFourStateOpsin* attribute), 147
  - `Gd1` (*cleo.opto.FourStateOpsin* attribute), 150
  - `Gd2` (*cleo.opto.BansalFourStateOpsin* attribute), 147
  - `Gd2` (*cleo.opto.FourStateOpsin* attribute), 150
  - `GECI` (class in *cleo.imaging*), 129
  - `geci()` (in module *cleo.imaging*), 134
  - `generate_Animation()` (*cleo.viz.VideoVisualizer* method), 157
  - `get_ctrl_signals()` (*cleo.ioproc.LatencyIOProcessor* method), 138
  - `get_ctrl_signals()` (*cleo.IOProcessor* method), 109
  - `get_intersample_ctrl_signal()` (*cleo.IOProcessor* method), 109
  - `get_orth_vectors_for_V()` (in module *cleo.utilities*), 156
  - `get_state()` (*cleo.CLSimulator* method), 108
  - `get_state()` (*cleo.ephys.MultiUnitSpiking* method), 117
  - `get_state()` (*cleo.ephys.Probe* method), 119
  - `get_state()` (*cleo.ephys.RWSLFPSignalBase* method), 120
  - `get_state()` (*cleo.ephys.Signal* method), 122
  - `get_state()` (*cleo.ephys.SortedSpiking* method), 123
  - `get_state()` (*cleo.ephys.Spiking* method), 124
  - `get_state()` (*cleo.ephys.TKLFPSignal* method), 125
  - `get_state()` (*cleo.imaging.GECI* method), 129
  - `get_state()` (*cleo.imaging.Scope* method), 131
  - `get_state()` (*cleo.imaging.Sensor* method), 133
  - `get_state()` (*cleo.Recorder* method), 112
  - `get_state()` (*cleo.recorders.GroundTruthSpikeRecorder* method), 152
  - `get_state()` (*cleo.recorders.RateRecorder* method), 153
  - `get_state()` (*cleo.recorders.VoltageRecorder* method), 153
  - `get_stim_values()` (*cleo.IOProcessor* method), 109
  - `Gf0` (*cleo.opto.BansalFourStateOpsin* attribute), 147
  - `Gf0` (*cleo.opto.FourStateOpsin* attribute), 150
  - `Gr` (*cleo.opto.BansalThreeStatePump* attribute), 148
  - `Gr0` (*cleo.opto.BansalFourStateOpsin* attribute), 147
  - `Gr0` (*cleo.opto.FourStateOpsin* attribute), 150
  - `GroundTruthSpikeRecorder` (class in *cleo.recorders*), 152
  - `gtacr2_4s()` (in module *cleo.opto*), 152
- ## I
- `i` (*cleo.ephys.Spiking* attribute), 124
  - `i` (*cleo.recorders.RateRecorder* attribute), 153
  - `I_per_Irr` (*cleo.opto.ProportionalCurrentOpsin* attribute), 151
  - `i_probe_by_i_ng` (*cleo.ephys.Spiking* attribute), 124
  - `i_targets_for_neuron_group()` (*cleo.imaging.Scope* method), 131
  - `i_targets_per_injct` (*cleo.imaging.Scope* attribute), 131
  - `I_threshold` (*cleo.ephys.RWSLFPSignalFromSpikes* attribute), 122
  - `img_width` (*cleo.imaging.Scope* attribute), 131
  - `init_for_probe()` (*cleo.ephys.Signal* method), 122
  - `init_for_simulator()` (*cleo.InterfaceDevice* method), 111
  - `init_for_simulator()` (*cleo.light.Light* method), 144
  - `init_for_simulator()` (*cleo.viz.VideoVisualizer* method), 157
  - `init_register_light()` (*cleo.registry.DeviceInteractionRegistry* method), 154
  - `init_syn_vars()` (*cleo.imaging.DoubExpCalBindingActivation* method), 128
  - `init_syn_vars()` (*cleo.imaging.DynamicCalcium* method), 128
  - `init_syn_vars()` (*cleo.imaging.GECI* method), 129
  - `init_syn_vars()` (*cleo.opto.BansalFourStateOpsin* method), 147
  - `init_syn_vars()` (*cleo.opto.BansalThreeStatePump* method), 149
  - `init_syn_vars()` (*cleo.opto.FourStateOpsin* method), 150
  - `init_syn_vars()` (*cleo.SynapseDevice* method), 113
  - `inject()` (*cleo.CLSimulator* method), 108
  - `inject_sensor_for_targets()` (*cleo.imaging.Scope* method), 131
  - `InterfaceDevice` (class in *cleo*), 110
  - `io_processor` (*cleo.CLSimulator* attribute), 108
  - `IOProcessor` (class in *cleo*), 109
  - `is_sampling_now()` (*cleo.ioproc.LatencyIOProcessor* method), 138
  - `is_sampling_now()` (*cleo.IOProcessor* method), 109

## J

jgcamp7b() (in module *cleo.imaging*), 134  
 jgcamp7c() (in module *cleo.imaging*), 135  
 jgcamp7f() (in module *cleo.imaging*), 135  
 jgcamp7s() (in module *cleo.imaging*), 135

## K

K (*cleo.light.OpticFiber* attribute), 146  
 k1 (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 k1 (*cleo.opto.FourStateOpsin* attribute), 150  
 k2 (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 k2 (*cleo.opto.FourStateOpsin* attribute), 150  
 K\_d (*cleo.imaging.GECI* attribute), 129  
 ka (*cleo.opto.BansalThreeStatePump* attribute), 149  
 kappa\_S (*cleo.imaging.DynamicCalcium* attribute), 128  
 kb (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 kb (*cleo.opto.FourStateOpsin* attribute), 150  
 kd (*cleo.opto.BansalThreeStatePump* attribute), 149  
 kf (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 kf (*cleo.opto.FourStateOpsin* attribute), 150  
 Koehler (class in *cleo.light*), 142

## L

LatencyIOProcessor (class in *cleo.ioproc*), 138  
 latest\_ctrl\_signal (*cleo.IOProcessor* attribute), 110  
 ldds\_for\_ng (*cleo.registry.DeviceInteractionRegistry* attribute), 154  
 lfp (*cleo.ephys.LFPSignalBase* attribute), 117  
 LFPSignalBase (class in *cleo.ephys*), 117  
 Light (class in *cleo.light*), 143  
 light\_agg\_ngs (*cleo.light.LightDependent* property), 145  
 light\_model (*cleo.light.Light* attribute), 144  
 light\_prop\_model (*cleo.registry.DeviceInteractionRegistry* attribute), 154  
 light\_prop\_syng (*cleo.registry.DeviceInteractionRegistry* attribute), 154  
 light\_source\_ng (*cleo.registry.DeviceInteractionRegistry* attribute), 154  
 LightDependent (class in *cleo.light*), 145  
 LightExcitation (class in *cleo.imaging*), 129  
 LightModel (class in *cleo.light*), 145  
 lights\_for\_ng (*cleo.registry.DeviceInteractionRegistry* attribute), 154  
 linear\_interpolator() (in module *cleo.light*), 146  
 linear\_shank\_coords() (in module *cleo.ephys*), 125  
 location (*cleo.imaging.GECI* attribute), 129  
 location (*cleo.imaging.Scope* attribute), 131  
 location (*cleo.imaging.Sensor* attribute), 133

## M

max\_Irr0\_mW\_per\_mm2 (*cleo.light.Light* attribute), 144

max\_Irr0\_mW\_per\_mm2\_viz (*cleo.light.Light* attribute), 144  
 model (*cleo.imaging.CalBindingActivationModel* attribute), 127  
 model (*cleo.imaging.CalciumModel* attribute), 127  
 model (*cleo.imaging.DoubExpCalBindingActivation* attribute), 128  
 model (*cleo.imaging.DynamicCalcium* attribute), 128  
 model (*cleo.imaging.ExcitationModel* attribute), 128  
 model (*cleo.imaging.LightExcitation* attribute), 130  
 model (*cleo.imaging.NullBindingActivation* attribute), 130  
 model (*cleo.imaging.NullExcitation* attribute), 130  
 model (*cleo.imaging.PreexistingCalcium* attribute), 130  
 model (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 model (*cleo.opto.BansalThreeStatePump* attribute), 149  
 model (*cleo.opto.FourStateOpsin* attribute), 150  
 model (*cleo.opto.ProportionalCurrentOpsin* attribute), 151  
 model (*cleo.SynapseDevice* attribute), 113  
 modify\_model\_and\_params\_for\_ng() (*cleo.SynapseDevice* method), 113  
 modify\_model\_with\_eqs() (in module *cleo.utilities*), 156  
 module  
   *cleo*, 108  
   *cleo.coords*, 115  
   *cleo.ephys*, 117  
   *cleo.imaging*, 127  
   *cleo.ioproc*, 137  
   *cleo.light*, 142  
   *cleo.opto*, 147  
   *cleo.recorders*, 152  
   *cleo.registry*, 154  
   *cleo.stimulators*, 155  
   *cleo.utilities*, 156  
   *cleo.viz*, 157  
 mon (*cleo.recorders.RateRecorder* attribute), 153  
 mon (*cleo.recorders.VoltageRecorder* attribute), 153  
 MultiUnitSpiking (class in *cleo.ephys*), 117

## N

n (*cleo.ephys.Probe* property), 119  
 n (*cleo.imaging.Scope* property), 131  
 n (*cleo.light.Light* property), 144  
 n\_H (*cleo.imaging.GECI* attribute), 129  
 NAFib (*cleo.light.OpticFiber* attribute), 146  
 name (*cleo.ephys.LFPSignalBase* attribute), 117  
 name (*cleo.ephys.Signal* attribute), 123  
 name (*cleo.InterfaceDevice* attribute), 111  
 NeoExportable (class in *cleo*), 112  
 network (*cleo.CLSimulator* attribute), 108  
 neuron\_group (*cleo.recorders.GroundTruthSpikeRecorder* attribute), 153

neuron\_groups (*cleo.imaging.Scope* attribute), 132  
 neuron\_groups (*cleo.stimulators.StateVariableSetter* attribute), 155  
 neuron\_groups (*cleo.viz.VideoVisualizer* attribute), 158  
 normalize\_coords() (in module *cleo.utilities*), 156  
 ntis (*cleo.light.OpticFiber* attribute), 146  
 NullBindingActivation (class in *cleo.imaging*), 130  
 NullExcitation (class in *cleo.imaging*), 130

## O

ogb1() (in module *cleo.imaging*), 136  
 on\_pre (*cleo.imaging.CalciumModel* attribute), 127  
 on\_pre (*cleo.imaging.DynamicCalcium* attribute), 128  
 on\_pre (*cleo.SynapseDevice* attribute), 114  
 Opsin (class in *cleo.opto*), 150  
 OpticFiber (class in *cleo.light*), 146  
 out\_buffer (*cleo.ioproc.LatencyIOProcessor* attribute), 138  
 out\_buffer (*cleo.ioproc.RecordOnlyProcessor* attribute), 141

## P

p (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 p (*cleo.opto.BansalThreeStatePump* attribute), 149  
 p (*cleo.opto.FourStateOpsin* attribute), 150  
 params (*cleo.imaging.GECI* property), 129  
 params (*cleo.SynapseDevice* property), 114  
 per\_ng\_unit\_replacements (*cleo.SynapseDevice* attribute), 114  
 phim (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 phim (*cleo.opto.BansalThreeStatePump* attribute), 149  
 phim (*cleo.opto.FourStateOpsin* attribute), 150  
 PIController (class in *cleo.ioproc*), 139  
 plot() (in module *cleo.viz*), 158  
 plot\_spectra() (in module *cleo.light*), 146  
 poly2\_shank\_coords() (in module *cleo.ephys*), 125  
 poly3\_shank\_coords() (in module *cleo.ephys*), 126  
 pop\_aggregate (*cleo.ephys.RWSLFPSignalBase* attribute), 120  
 PreexistingCalcium (class in *cleo.imaging*), 130  
 preprocess\_ctrl\_signals() (*cleo.IOProcessor* method), 110  
 Probe (class in *cleo.ephys*), 118  
 probe (*cleo.ephys.LFPSignalBase* attribute), 117  
 probe (*cleo.ephys.Probe* attribute), 119  
 probe (*cleo.ephys.Signal* attribute), 123  
 process() (*cleo.ioproc.LatencyIOProcessor* method), 138  
 process() (*cleo.ioproc.ProcessingBlock* method), 140  
 process() (*cleo.ioproc.RecordOnlyProcessor* method), 141  
 processing (*cleo.ioproc.LatencyIOProcessor* attribute), 138

processing (*cleo.ioproc.RecordOnlyProcessor* attribute), 141  
 ProcessingBlock (class in *cleo.ioproc*), 140  
 ProportionalCurrentOpsin (class in *cleo.opto*), 151  
 Psi0 (*cleo.opto.BansalThreeStatePump* attribute), 148  
 put\_state() (*cleo.ioproc.LatencyIOProcessor* method), 139  
 put\_state() (*cleo.IOProcessor* method), 110

## Q

q (*cleo.opto.BansalFourStateOpsin* attribute), 148  
 q (*cleo.opto.BansalThreeStatePump* attribute), 149  
 q (*cleo.opto.FourStateOpsin* attribute), 150

## R

R0 (*cleo.light.OpticFiber* attribute), 146  
 r\_half\_detection (*cleo.ephys.Spiking* attribute), 124  
 r\_perfect\_detection (*cleo.ephys.Spiking* attribute), 124  
 radius (*cleo.light.Koehler* attribute), 142  
 rand\_seed (*cleo.imaging.Scope* attribute), 132  
 RateRecorder (class in *cleo.recorders*), 153  
 Recorder (class in *cleo*), 112  
 recorders (*cleo.CLSimulator* attribute), 108  
 RecordOnlyProcessor (class in *cleo.ioproc*), 141  
 ref\_signal (*cleo.ioproc.PIController* attribute), 139  
 register() (*cleo.registry.DeviceInteractionRegistry* method), 154  
 register\_ldd() (*cleo.registry.DeviceInteractionRegistry* method), 155  
 register\_light() (*cleo.registry.DeviceInteractionRegistry* method), 155  
 registries (in module *cleo.registry*), 155  
 registry\_for\_sim() (in module *cleo.registry*), 155  
 required\_vars (*cleo.opto.ProportionalCurrentOpsin* attribute), 151  
 required\_vars (*cleo.SynapseDevice* attribute), 114  
 reset() (*cleo.CLSimulator* method), 108  
 reset() (*cleo.ephys.Probe* method), 119  
 reset() (*cleo.ephys.RWSLFPSignalBase* method), 120  
 reset() (*cleo.ephys.RWSLFPSignalFromPSCs* method), 121  
 reset() (*cleo.ephys.Signal* method), 123  
 reset() (*cleo.ephys.Spiking* method), 124  
 reset() (*cleo.ephys.TKLFPSignal* method), 125  
 reset() (*cleo.imaging.Scope* method), 132  
 reset() (*cleo.InterfaceDevice* method), 111  
 reset() (*cleo.IOProcessor* method), 110  
 reset() (*cleo.Stimulator* method), 112  
 reset() (*cleo.SynapseDevice* method), 114  
 rho\_rel\_per\_injct (*cleo.imaging.Scope* attribute), 132  
 rng (in module *cleo.utilities*), 156  
 run() (*cleo.CLSimulator* method), 108

RWSLFPSignalBase (class in *cleo.ephys*), 120  
 RWSLFPSignalFromPSCs (class in *cleo.ephys*), 120  
 RWSLFPSignalFromSpikes (class in *cleo.ephys*), 121

## S

S (*cleo.light.OpticFiber* attribute), 146  
 sample\_period\_ms (*cleo.IOProcessor* attribute), 110  
 sampling (*cleo.ioproc.LatencyIOProcessor* attribute), 139  
 sampling (*cleo.ioproc.RecordOnlyProcessor* attribute), 141  
 save\_history (*cleo.InterfaceDevice* attribute), 111  
 save\_history (*cleo.ioproc.ProcessingBlock* attribute), 140  
 Scope (class in *cleo.imaging*), 130  
 Sensor (class in *cleo.imaging*), 132  
 sensor (*cleo.imaging.Scope* attribute), 132  
 set\_io\_processor() (*cleo.CLSimulator* method), 108  
 sigma\_axial (*cleo.light.GaussianEllipsoid* attribute), 142  
 sigma\_lateral (*cleo.light.GaussianEllipsoid* attribute), 142  
 sigma\_noise (*cleo.imaging.Scope* property), 132  
 sigma\_noise (*cleo.imaging.Sensor* attribute), 133  
 sigma\_per\_injct (*cleo.imaging.Scope* attribute), 132  
 Signal (class in *cleo.ephys*), 122  
 signals (*cleo.ephys.Probe* attribute), 119  
 sim (*cleo.InterfaceDevice* attribute), 111  
 sim (*cleo.registry.DeviceInteractionRegistry* attribute), 155  
 snr (*cleo.imaging.Sensor* property), 133  
 snr\_cutoff (*cleo.imaging.Scope* attribute), 132  
 soma\_radius (*cleo.imaging.Scope* attribute), 132  
 SortedSpiking (class in *cleo.ephys*), 123  
 source (*cleo.light.Light* property), 144  
 source\_for\_light() (*cleo.registry.DeviceInteractionRegistry* method), 155  
 source\_ngs (*cleo.SynapseDevice* attribute), 114  
 spectrum (*cleo.light.LightDependent* attribute), 145  
 spectrum\_interpolator (*cleo.light.LightDependent* attribute), 145  
 Spiking (class in *cleo.ephys*), 123  
 StateVariableSetter (class in *cleo.stimulators*), 155  
 Stimulator (class in *cleo*), 112  
 stimulators (*cleo.CLSimulator* attribute), 109  
 style\_plots\_for\_docs() (in module *cleo.utilities*), 156  
 subgroup\_idx\_for\_light (*cleo.registry.DeviceInteractionRegistry* attribute), 155  
 syn\_delay (*cleo.ephys.RWSLFPSignalFromSpikes* attribute), 122  
 SynapseDevice (class in *cleo*), 113  
 synapses (*cleo.SynapseDevice* attribute), 114

## T

t\_in\_ms (*cleo.ioproc.ProcessingBlock* attribute), 140  
 t\_ms (*cleo.ephys.LFPSignalBase* attribute), 117  
 t\_ms (*cleo.ephys.Spiking* attribute), 124  
 t\_ms (*cleo.imaging.Scope* attribute), 132  
 t\_ms (*cleo.Stimulator* attribute), 112  
 t\_out\_ms (*cleo.ioproc.ProcessingBlock* attribute), 141  
 t\_samp\_ms (*cleo.ephys.Spiking* attribute), 124  
 t\_samp\_ms (*cleo.ioproc.LatencyIOProcessor* attribute), 139  
 t\_samp\_ms (*cleo.ioproc.RecordOnlyProcessor* attribute), 141  
 target\_neurons\_in\_plane() (*cleo.imaging.Scope* method), 132  
 target\_neurons\_in\_plane() (in module *cleo.imaging*), 136  
 tau1\_ampa (*cleo.ephys.RWSLFPSignalFromSpikes* attribute), 122  
 tau1\_gaba (*cleo.ephys.RWSLFPSignalFromSpikes* attribute), 122  
 tau2\_ampa (*cleo.ephys.RWSLFPSignalFromSpikes* attribute), 122  
 tau2\_gaba (*cleo.ephys.RWSLFPSignalFromSpikes* attribute), 122  
 tau\_off (*cleo.imaging.DoubExpCalBindingActivation* attribute), 128  
 tau\_on (*cleo.imaging.DoubExpCalBindingActivation* attribute), 128  
 tetrode\_shank\_coords() (in module *cleo.ephys*), 126  
 tile\_coords() (in module *cleo.ephys*), 127  
 times\_are\_regular() (in module *cleo.utilities*), 156  
 TKLFPSignal (class in *cleo.ephys*), 124  
 to\_neo() (*cleo.CLSimulator* method), 109  
 to\_neo() (*cleo.ephys.LFPSignalBase* method), 117  
 to\_neo() (*cleo.ephys.MultiUnitSpiking* method), 118  
 to\_neo() (*cleo.ephys.Probe* method), 119  
 to\_neo() (*cleo.ephys.RWSLFPSignalBase* method), 120  
 to\_neo() (*cleo.ephys.Spiking* method), 124  
 to\_neo() (*cleo.light.Light* method), 144  
 to\_neo() (*cleo.NeoExportable* method), 112  
 to\_neo() (*cleo.Stimulator* method), 112  
 tp\_light\_from\_scope() (in module *cleo.light*), 146  
 transmittance() (*cleo.light.GaussianEllipsoid* method), 142  
 transmittance() (*cleo.light.Koehler* method), 142  
 transmittance() (*cleo.light.Light* method), 144  
 transmittance() (*cleo.light.LightModel* method), 145  
 transmittance() (*cleo.light.OpticFiber* method), 146

## U

uLFP\_threshold\_uV (*cleo.ephys.TKLFPSignal* attribute), 125  
 uniform\_cylinder\_rz() (in module *cleo.utilities*), 156  
 unit (*cleo.stimulators.StateVariableSetter* attribute), 155



`unit_safe_append()` (in module *cleo.utilities*), 156  
`update()` (*cleo.light.Light* method), 144  
`update()` (*cleo.Stimulator* method), 112  
`update()` (*cleo.stimulators.StateVariableSetter* method), 155  
`update_artists()` (*cleo.InterfaceDevice* method), 111  
`update_artists()` (*cleo.light.Light* method), 144  
`update_stimulators()` (*cleo.CLSimulator* method), 109

## V

`v0` (*cleo.opto.FourStateOpsin* attribute), 150  
`v1` (*cleo.opto.FourStateOpsin* attribute), 150  
`value` (*cleo.Stimulator* attribute), 113  
`values` (*cleo.ioproc.ProcessingBlock* attribute), 141  
`values` (*cleo.Stimulator* attribute), 113  
`variable_to_ctrl` (*cleo.stimulators.StateVariableSetter* attribute), 156  
`vartheta_max` (*cleo.opto.BansalThreeStatePump* attribute), 149  
`vfchromson_4s()` (in module *cleo.opto*), 152  
`VideoVisualizer` (class in *cleo.viz*), 157  
`viz_params()` (*cleo.light.GaussianEllipsoid* method), 142  
`viz_params()` (*cleo.light.Koehler* method), 142  
`viz_params()` (*cleo.light.LightModel* method), 145  
`viz_params()` (*cleo.light.OpticFiber* method), 146  
`vmax` (*cleo.opto.BansalThreeStatePump* attribute), 149  
`vmin` (*cleo.opto.BansalThreeStatePump* attribute), 149  
`voltage_var_name` (*cleo.recorders.VoltageRecorder* attribute), 153  
`VoltageRecorder` (class in *cleo.recorders*), 153

## W

`wavelength` (*cleo.light.Light* attribute), 145  
`wavelength_to_rgb()` (in module *cleo.utilities*), 156  
`weight` (*cleo.ephys.RWSLFPSignalFromSpikes* attribute), 122

## X

`xs` (*cleo.ephys.Probe* property), 119  
`xyz_from_rz()` (in module *cleo.utilities*), 156

## Y

`ys` (*cleo.ephys.Probe* property), 119

## Z

`zmax` (*cleo.light.Koehler* attribute), 142  
`zs` (*cleo.ephys.Probe* property), 120